TOXICOLOGICAL PROFILE FOR
METHYLENEDIANILINE

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES
Public Health Service
Agency for Toxic Substances and Disease Registry

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UPDATE STATEMENT

A Toxicological Profile for methylenedianiline was released in October 1995. This edition supersedes any previously released draft or final profile.

Toxicological profiles are revised and republished as necessary, but no less than once every three years. For information regarding the update status of previously released profiles, contact ATSDR at:

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Division of Toxicology/Toxicology Information Branch
1600 Clifton Road NC, E-29
Atlanta, Georgia 30333
FOREWORD

This toxicological profile is prepared in accordance with guidelines developed by the Agency for Toxic Substances and Disease Registry (ATSDR) and the Environmental Protection Agency (EPA). The original guidelines were published in the Federal Register on April 17, 1987. Each profile will be revised and republished as necessary.

The ATSDR toxicological profile succinctly characterizes the toxicologic and adverse health effects information for the hazardous substance described therein. Each peer-reviewed profile identifies and reviews the key literature that describes a hazardous substance's toxicologic properties. Other pertinent literature is also presented, but is described in less detail than the key studies. The profile is not intended to be an exhaustive document; however, more comprehensive sources of specialty information are referenced.

The focus of the profiles is on health and toxicologic information; therefore, each toxicological profile begins with a public health statement that describes, in nontechnical language, a substance's relevant toxicological properties. Following the public health statement is information concerning levels of significant human exposure and, where known, significant health effects. The adequacy of information to determine a substance's health effects is described in a health effects summary. Data needs that are of significance to protection of public health are identified by ATSDR and EPA.

Each profile includes the following:

(A) The examination, summary, and interpretation of available toxicologic information and epidemiologic evaluations on a hazardous substance to ascertain the levels of significant human exposure for the substance and the associated acute, subacute, and chronic health effects;

(B) A determination of whether adequate information on the health effects of each substance is available or in the process of development to determine levels of exposure that present a significant risk to human health of acute, subacute, and chronic health effects; and

(C) Where appropriate, identification of toxicologic testing needed to identify the types or levels of exposure that may present significant risk of adverse health effects in humans.

The principal audiences for the toxicological profiles are health professionals at the Federal, State, and local levels; interested private sector organizations and groups; and members of the public.

This profile reflects ATSDR's assessment of all relevant toxicologic testing and information that has been peer-reviewed. Staff of the Centers for Disease Control and Prevention and other Federal scientists have also reviewed the profile. In addition, this profile has been peer-reviewed by a nongovernmental panel and was made available for public review. Final responsibility for the contents and views expressed in this toxicological profile resides with ATSDR.

Claire V. Broome, M.D.
Acting Administrator
Agency for Toxic Substances and Disease Registry
The toxicological profiles are developed in response to the Super-fund Amendments and Reauthorization Act (SARA) of 1986 (Public Law 99-499) which amended the Comprehensive Environmental Response, Compensation, and Liability Act of 1980 (CERCLA or Super-fund). Section 211 of SARA also amended Title 10 of the U. S. Code, creating the Defense Environmental Restoration Program. Section 2704(a) of Title 10 of the U. S. Code directs the Secretary of Defense to notify the Secretary of Health and Human Services of not less than 25 of the most commonly found unregulated hazardous substances at defense facilities. Section 2704(b) of Title 10 of the U. S. Code directs the Administrator of the Agency for Toxic Substances and Disease Registry (ATSDR) to prepare a toxicological profile for each substance on the list provided by the Secretary of Defense under subsection (b).
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THE PROFILE HAS UNDERGONE THE FOLLOWING ATSDR INTERNAL REVIEWS:


2. Health Effects Review. The Health Effects Review Committee examines the health effects chapter of each profile for consistency and accuracy in interpreting health effects and classifying end points.

3. Minimal Risk Level Review. The Minimal Risk Level Workgroup considers issues relevant to substance-specific minimal risk levels (MRLs), reviews the health effects database of each profile, and makes recommendations for derivation of MRLs.

4. Data Needs Review. The Research Implementation Branch reviews data needs sections to assure consistency across profiles and adherence to instructions in the Guidance.
A peer review panel was assembled for 4,4'-methylenedianiline. The panel consisted of the following members:

1. Dr. G.A.S. Ansari, Professor, Department of Human Biological Chemistry and Genetics and Pathology, University of Texas Medical Branch, Galveston, TX 77555;

2. Dr. W. Decker, Private Consultant, El Paso, TX 79904; and

3. Dr. E. Sowinski, Vice President, Environmental Health Management and Science, Hudson, OH 44236.

These experts collectively have knowledge of 4,4'-methylenedianiline’s physical and chemical properties, toxicokinetics, key health end points, mechanisms of action, human and animal exposure, and quantification of risk to humans. All reviewers were selected in conformity with the conditions for peer review specified in Section 104(i)(13) of the Comprehensive Environmental Response, Compensation, and Liability Act, as amended.

Scientists from the Agency for Toxic Substances and Disease Registry (ATSDR) have reviewed the peer reviewers’ comments and determined which comments will be included in the profile. A listing of the peer reviewers’ comments not incorporated in the profile, with a brief explanation of the rationale for their exclusion, exists as part of the administrative record for this compound. A list of databases reviewed and a list of unpublished documents cited are also included in the administrative record.

The citation of the peer review panel should not be understood to imply its approval of the profile’s final content. The responsibility for the content of this profile lies with the ATSDR.
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1. PUBLIC HEALTH STATEMENT

This public health statement tells you about 4,4′-methylenedianiline and the effects of exposure.

The Environmental Protection Agency (EPA) identifies the most serious hazardous waste sites in the nation. These sites make up the National Priorities List (NPL) and are the sites targeted for long-term federal clean-up activities. 4,4′-methylenedianiline has not been found in any of the 1,445 current or former NPL sites. However, the total number of NPL sites evaluated is not known. As more sites are evaluated, the number of sites at which 4,4′-methylenedianiline is found may increase. This is important because exposure to this substance may harm you and because these sites may be sources of exposure.

When a substance is released from a large area, such as an industrial plant, or from a container, such as a drum or bottle, it enters the environment. This release does not always lead to exposure. You can be exposed to a substance only when you come in contact with it by breathing, eating, touching, or drinking.

If you are exposed to 4,4′-methylenedianiline many factors determine whether you’ll be harmed. These factors include the dose (how much), the duration (how long), and how you come in contact with it. You must also consider the other chemicals you’re exposed to and your age, sex, diet, family traits, lifestyle, and state of health.

1.1 WHAT IS METHYLENEDIANILINE?

Methylenedianiline exists in several very similar forms. Of these various forms, 4,4′-methylenedianiline is the form used by industries. This profile discusses this important form of methylenedianiline. Other forms of methylenedianiline are only used as laboratory chemicals and have no commercial uses at this time. 4,4′-Methylenedianiline is also commonly known as diaminodiphenylmethane or MDA. It is a colorless (like an ice cube) to pale yellow solid with a faint amine odor (sharp odor). The taste of the compound is not
known. It is very slightly soluble in water (does not mix well with water) and does not readily evaporate at room temperature. If left in an open container, it slowly turns brown because of chemical reactions with components present in air.

4,4’-methylenedianiline is an industrially produced compound that is not known to occur naturally. It is produced by industries mainly for making polyurethane foams (such as insulating materials in mailing containers). Smaller amounts are used for making coating materials, epoxy resins (glues), Spandex fiber, and dyes and for other purposes. You can find more information on the properties, sources, and uses of 4,4’-methylenedianiline in Chapters 3 and 4.

1.2 WHAT HAPPENS TO METHYLENEDIANILINE WHEN IT ENTERS THE ENVIRONMENT?

Most 4,4’-methylenedianiline enters the environment when it’s produced or when it’s used to make other compounds. Of the total environmental release of at least 20,000 pounds per year, 52.6% is released to the air and 45% is released to deep soil during underground injection. Only 2.4% of the total is released to land and water. In addition, about 2,000 pounds per year are transferred to public sewer systems for treatment. No estimate is available for the amounts of 4,4’-methylenedianiline that enter the environment from accidental spills or from identified hazardous waste sites that may contain dianiline.

The lack of experimental data makes it difficult to be certain about what happens to 4,4’-methylenedianiline when it enters the environment. In air, 4,4’-methylenedianiline will mostly be present as tiny particles and will eventually return to land and water by settling and by being brought down in rain and snow. In water, most of the 4,4’-methylenedianiline will tend to attach itself to particles and sediments, and will eventually settle to the bottom. 4,4’-methylenedianiline present in water and sediment will eventually be broken down by bacteria and other microorganisms. This process may take as long as 10–40 days. 4,4’-methylenedianiline does not tend to build up in the food chain, and it is uncertain
whether it accumulates in fish. When deposited on soil, 4,4’-methylenedianiline will become strongly attached to it and, as a result, will not move quickly with rainwater into deeper groundwater. Bacteria and microorganisms present in soil will break down dianiline, but the process may take as long as 10 days. See Chapter 5 for more information about what happens to 4,4’-methylenedianiline in the environment.

1.3 HOW MIGHT I BE EXPOSED TO METHYLENEDIANILINE?

You are most likely to be exposed to 4,4’-methylenedianiline if you work with it. The general population may be exposed to extremely low levels of 4,4’-methylenedianiline through consumer goods such as polyurethane cushioning or products that contain epoxy. The Food and Drug Administration (FDA) reports that the level of exposure to 4,4’-methylenedianiline through food is virtually zero. People who live near hazardous waste sites that contain 4,4’-methylenedianiline are susceptible to exposure if dust particles containing the substance are released from the waste site, enter the air, and are breathed into the body. Children playing near these sites may be exposed by touching and eating soil that contains 4,4’-methylenedianiline However, there is no experimental or estimated value for the intake of 4,4’-methylenedianiline by the general population.

People who work in the following industries can be exposed to 4,4’-methylenedianiline by breathing in the dust or aerosol, or by getting it on their skin: manufacture, formulation, and packaging of 4,4’-methylenedianiline certain paint making industries that use epoxy materials; pattern and tool with polyurethane; potting and encapsulation with polyurethane; and casting and molding with resins made with 4,4’-methylenedianiline 4,4’-methylenedianiline has been detected in workplace air, in skin patches worn by workers, and in the urine of workers in these industries. However, the level in work atmospheres rarely exceeds the level of 0.8 milligram (mg) of 4,4’-methylenedianiline in a cubic meter of air (1 mg is equivalent to a thousandth of a gram), which is suggested as a safe level by the American Conference of Governmental Industrial Hygienists (ACGIH). The maximum exposure was found to occur in workers in the manufacturing and formulating industries. People with diseases or who need frequent blood transfusions may be exposed to tiny
amounts of 4,4'-methylenedianiline during their treatment by machines like dialyzers. The compound is released from polyurethane parts of equipment when they are sterilized with radiation or heat. See Chapter 5 for more information about 4,4'-methylenedianiline exposure.

1.4 HOW CAN METHYLENEDIANILINE ENTER AND LEAVE MY BODY?

If you breathe air containing dust contaminated with 4,4'-methylenedianiline, it can enter your body through your lungs and pass into the bloodstream. We do not know how much of the 4,4'-methylenedianiline will pass into your bloodstream or how fast this will happen. If you swallow food, water, or soil contaminated with 4,4'-methylenedianiline, some of it will probably enter your body and pass from the stomach into the bloodstream, but we do not know how much or how fast this will occur. If you touch soil containing 4,4'-methylenedianiline (for example, at a hazardous waste site), some 4,4'-methylenedianiline will pass through the skin into the bloodstream, but we do not know how much or how fast. For people living around waste sites, or processing, or storage facilities, the most likely way it will enter their bodies is from skin contact with contaminated soil. For people who work with or around 4,4'-methylenedianiline, skin contact with contaminated dirt particles is the most likely way it will enter the body. Once 4,4'-methylenedianiline is in your body, it may change into other related chemicals called metabolites. We think that some metabolites of 4,4'-methylenedianiline may be more harmful than unchanged 4,4'-methylenedianiline, but there is no conclusive experimental evidence to support this assumption. Some of the metabolites may leave your body in the urine within hours. We do not know if 4,4'-methylenedianiline is stored in the body. For more information on how 4,4'-methylenedianiline can enter and leave your body, see Chapter 2.

1.5 HOW CAN METHYLENEDIANILINE AFFECT MY HEALTH?

Studies in the workplace suggest that exposure to 4,4'-methylenedianiline may cause skin irritation and can damage your liver. People who ate bread that was accidentally contaminated with 4,4'-methylenedianiline also experienced liver damage. We do not know,
however, how much 4,4´-methylenedianiline was in the bread. We do not know if breathing air contaminated with 4,4´-methylenedianiline, or eating contaminated food, or skin contact affects human reproduction or development. We also do not know if 4,4´-methylenedianiline affects the nervous system or the ability to fight disease in humans.

To protect the public from the harmful effects of toxic chemicals and to find ways to treat people who have been harmed, scientists use many tests.

One way to see if a chemical will hurt people is to learn how the chemical is absorbed, used, and released by the body; for some chemicals, animal testing may be necessary. Animal testing may also be used to identify health effects such as cancer or birth defects. Without laboratory animals, scientists would lose a basic method to get information needed to make wise decisions to protect public health. Scientists must treat research animals with care and compassion. Laws today protect the welfare of research animals, and scientists must comply with strict animal care guidelines.

Some mice that drank water containing large amounts of 4,4´-methylenedianiline for a short period died as a result. Rats that ate food or drank water containing smaller amounts of 4,4´-methylenedianiline for months or years had liver damage and thyroid gland injuries. Only a small amount of information exists on the health effects in animals exposed to 4,4´-methylenedianiline by breathing or skin contact. This information indicates that guinea pigs that breathed air contaminated with a very high amount of 4,4´-methylenedianiline for 2 weeks suffered damage to their eyes. Rabbits that received repeated skin exposure to relatively high amounts of 4,4´-methylenedianiline also had liver damage. There is not enough information to determine if exposure to 4,4´-methylenedianiline affects reproduction, development, the nervous system, or the ability to fight disease in animals.

There is not enough information on workers to determine if 4,4´-methylenedianiline is carcinogenic (causes cancer) in people. Rats and mice that drank water containing 4,4´-methylenedianiline throughout their lives developed cancer in their liver and thyroid glands. The International Agency for Research on Cancer (IARC) has determined that
4,4´-methyleneedianiline is possibly carcinogenic to humans. For more information on how 4,4´-methyleneedianiline can affect your health, see Chapter 2.

1.6 IS THERE A MEDICAL TEST TO DETERMINE WHETHER I HAVE BEEN EXPOSED TO METHYLENEDIANILINE?

Samples of your urine can be tested to find out if you have recently been exposed to 4,4´-methyleneedianiline. These tests will only confirm if you have been exposed. They cannot estimate how much of it has entered your body and will not tell you whether your health will be affected by exposure to 4,4´-methyleneedianiline. The exposure tests are not routinely available in hospitals and clinics because they require special analytical equipment and must be specially requested by your physician. See Chapters 2 and 6 for more information about tests for exposure to 4,4´-methyleneedianiline.

1.7 WHAT RECOMMENDATIONS HAS THE FEDERAL GOVERNMENT MADE TO PROTECT HUMAN HEALTH?

The federal government develops regulations and recommendations to protect public health. Regulations can be enforced by law. Federal agencies that develop regulations for toxic substances include the Environmental Protection Agency (EPA), the Occupational Safety and Health Administration (OSHA), and the Food and Drug Administration (FDA). Recommendations provide valuable guidelines to protect public health but cannot be enforced by law. Federal organizations that develop recommendations for toxic substances include the Agency for Toxic Substances and Disease Registry (ATSDR) and the National Institute for Occupational Safety and Health (NIOSH).

Regulations and recommendations can be expressed in not-to-exceed levels in air, water, soil, or food that are usually based on levels that affect animals; then they are adjusted to help protect people. Sometimes these not-to-exceed levels differ among federal organizations because of different exposure times (an 8-hour workday or a 24-hour day), the use of different animal studies, or other factors.
Recommendations and regulations are also periodically updated as more information becomes available. For the most current information, check with the federal agency or organization that provides it. Some regulations and recommendations for 4,4’-methylenedianiline include the following:

NIOSH recommends that workers should not breathe air that contains more than 0.03 milligram of 4,4’-methylenedianiline per cubic meter of air (0.03 mg/m³) during a 10-hour workday, 40-hour workweek.

OSHA recommends that workers should not be exposed to more than 0.081 mg/m³ 4,4’-methylenedianiline during an 8-hour workday.

ACGIH recommends that workers should not be exposed to more than 0.81 mg/m³ 4,4’-methylenedianiline for an 8-hour workday, 40-hour workweek.

See Chapter 7 for more information on regulations and guidelines concerning 4,4’-methylenedianiline.

1.8 WHERE CAN I GET MORE INFORMATION?

If you have any more questions or concerns, please contact your community or state health or environmental quality department or

Agency for Toxic Substances and Disease Registry
Division of Toxicology
1600 Clifton Road NE, Mailstop E-29
Atlanta, GA 30333

Information line and technical assistance

Phone: (404) 639-6000
Fax: (404) 639-6315 or 6324

ATSDR can also tell you the location of occupational and environmental health clinics. These clinics specialize in recognizing, evaluation and treating illnesses resulting from exposure to hazardous substances.
To order toxicological profiles, contact

National Technical Information Service
5285 Port Royal Road
Springfield, VA 22161
Phone: (800) 553-6847 or (703) 487-4650
2. HEALTH EFFECTS

2.1 INTRODUCTION

The primary purpose of this chapter is to provide public health officials, physicians, toxicologists, and other interested individuals and groups with an overall perspective of the toxicology of 4,4´-methylenedianiline. It contains descriptions and evaluations of toxicological studies and epidemiological investigations and provides conclusions, where possible, on the relevance of toxicity and toxicokinetic data to public health.

Methylenedianilines can exist in five isomeric forms: 2,2´-methylenedianiline; 2,4´-methylenedianiline; 3,3´-methylenedianiline; 3,4´-methylenedianiline; and 4,4´-methylenedianiline. Of the various isomers, 2,2´-methylenedianiline, 3,4´-methylenedianiline, 3,3´-methylenedianiline, and 2,4´-methylenedianiline are produced on a very small scale as a research chemical (HSDB 1996). The isomer 4,4´-methylenedianiline is produced in the United States for industrial use. Therefore, this profile will limit its discussion to 4,4´-methylenedianiline.

A glossary and list of acronyms, abbreviations, and symbols can be found at the end of this profile.

2.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE

To help public health professionals and others address the needs of persons living or working near hazardous waste sites, the information in this section is organized first by route of exposure-inhalation, oral, and dermal; and then by health effect-death, systemic, immunological, neurological, reproductive, developmental, genotoxic, and carcinogenic effects. These data are discussed in terms of three exposure periods-acute (14 days or less), intermediate (15-364 days), and chronic (365 days or more).

Levels of significant exposure for each route and duration are presented in tables and illustrated in figures. The points in the figures showing no-observed-adverse-effect levels (NOAELs) or lowest-observed-adverse-effect levels (LOAELs) reflect the actual doses (levels of exposure) used in the studies. LOAELs have been classified into “less serious” or “serious” effects. “Serious” effects are those that evoke failure in a biological system and can lead to morbidity or mortality (e.g., acute
respiratory distress or death). “Less serious” effects are those that are not expected to cause significant dysfunction or death, or those whose significance to the organism is not entirely clear. ATSDR acknowledges that a considerable amount of judgment may be required in establishing whether an end point should be classified as a NOAEL, “less serious” LOAEL, or “serious” LOAEL, and that in some cases, there will be insufficient data to decide whether the effect is indicative of significant dysfunction. However, the Agency has established guidelines and policies that are used to classify these end points. ATSDR believes that there is sufficient merit in this approach to warrant an attempt at distinguishing between “less serious” and “serious” effects. The distinction between “less serious” effects and “serious” effects is considered to be important because it helps the users of the profiles to identify levels of exposure at which major health effects start to appear. LOAELs or NOAELs should also help in determining whether or not the effects vary with dose and/or duration, and place into perspective the possible significance of these effects to human health.

The significance of the exposure levels shown in the Levels of Significant Exposure (LSE) tables and figures may differ depending on the user’s perspective. Public health officials and others concerned with appropriate actions to take at hazardous waste sites may want information on levels of exposure associated with more subtle effects in humans or animals (LOAEL) or exposure levels below which no adverse effects (NOAELs) have been observed. Estimates of levels posing minimal risk to humans (Minimal Risk Levels or MRLs) may be of interest to health professionals and citizens alike.

Levels of exposure associated with carcinogenic effects (Cancer Effect Levels, CELs) of 4,4´-methylenedianiline are indicated in Tables 2-2 and 2-3 and Figure 2-2.

Estimates of exposure levels posing minimal risk to humans (Minimal Risk Levels or MRLs) have been made for 4,4´-methylenedianiline. An MRL is defined as an estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncancerogenic) over a specified duration of exposure. MRLs are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. MRLs are based on noncancerous health effects only and do not consider carcinogenic effects. MRLs can be derived for acute, intermediate, and chronic duration exposures for inhalation and oral routes. Appropriate methodology does not exist to develop MRLs for dermal exposure.
Although methods have been established to derive these levels (Barnes and Dourson 1988; EPA 1990), uncertainties are associated with these techniques. Furthermore, ATSDR acknowledges additional uncertainties inherent in the application of the procedures to derive less than lifetime MRLs. As an example, acute inhalation MRLs may not be protective for health effects that are delayed in development or are acquired following repeated acute insults, such as hypersensitivity reactions, asthma, or chronic bronchitis. As these kinds of health effects data become available and methods to assess levels of significant human exposure improve, these MRLs will be revised.

A User’s Guide has been provided at the end of this profile (see Appendix B). This guide should aid in the interpretation of the tables and figures for Levels of Significant Exposure and the MRLs.

### 2.2.1 Inhalation Exposure

Occupational exposure to 4,4´-methylenedianiline probably involves both inhalation of dust particles containing the chemical, and dermal contact with these particles. In addition, direct ingestion of contaminated dust or ingestion of particles that are expelled from the respiratory tree cannot be ruled out. It is generally agreed, however, that dermal contact is the main contributing route of exposure in occupational settings. For this reason, health effects in humans attributed to occupational exposure to 4,4´-methylenedianiline are discussed in Section 2.2.3, Dermal Exposure.

#### 2.2.1.1 Death

No studies were located regarding death in humans or animals after inhalation exposure to 4,4´-methylenedianiline.

#### 2.2.1.2 Systemic Effects

No studies were located regarding systemic effects in humans after inhalation exposure to 4,4´-methylenedianiline. In addition, no studies were located regarding cardiovascular, gastrointestinal, hematological, or musculoskeletal effects in animals after inhalation exposure to 4,4´-methylenedianiline. Only one study was located that provided some information on systemic effects of inhaled 4,4´-methylenedianiline in animals; this limited information is summarized below.
The highest NOAEL values and all LOAEL values for each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1.

**Respiratory Effects.** Guinea pigs exposed nose-only to 440 mg/m$^3$ of an aerosol of 4,4’-methylenedianiline in propylene glycol 4 hours per day, 5 days per week for 2 weeks experienced no respiratory distress during exposures (Leong et al. 1987). The mean particle diameter was 2.4 µm. Two weeks after exposures terminated and prior to sacrifice, tests were conducted to detect possible changes in the distensibility of the lungs from an intratracheal challenge dose of 4,4’-methylenedianiline; the results were unremarkable. Histopathologic examination of the lungs revealed mild to slight pneumonia and pulmonary granulomas.

**Hepatic Effects.** No histopathological alterations were observed in the liver of guinea pigs exposed nose-only to 440 mg/m$^3$ of an aerosol of 4,4’-methylenedianiline 4 hours per day, 5 days per week for 2 weeks (Leong et al. 1987). The animals were sacrificed 2 weeks after the exposure period terminated. No further information regarding hepatic effects after inhalation exposure to 4,4’-methylenedianiline was located.

**Renal Effects.** No histopathological alterations were observed in the kidneys of guinea pigs exposed nose-only to 440 mg/m$^3$ of an aerosol of 4,4’-methylenedianiline 4 hours per day, 5 days per week for 2 weeks (Leong et al. 1987). Sacrifices were conducted 2 weeks after the exposure period terminated. No further information regarding renal effects following inhalation exposure to 4,4’-methylenedianiline was located.

**Dermal Effects.** The possibility that inhalation exposure to 4,4’-methylenedianiline could induce dermal sensitization was explored in guinea pigs (Leong et al. 1987) (see Section 2.2.3.2). The animals were exposed nose-only to 440 mg/m$^3$ of an aerosol of 4,4’-methylenedianiline 4 hours per day, 5 days per week for 2 weeks. Two weeks after exposure ceased, 4,4’-methylenedianiline (0.2-22 mg/kg) was applied to shaved sites of skin and observations were made for up to 24 hours. Neither erythema nor edema were observed suggesting that under the conditions of the experiment, 4,4’-methylenedianiline was not a dermal sensitizer. No further information was located regarding dermal effects after inhalation exposure to 4,4’-methylenedianiline.
Table 2-1. Levels of Significant Exposure to 4,4'-Methylenedianiline - Inhalation

<table>
<thead>
<tr>
<th>Key to figure</th>
<th>Species (strain)</th>
<th>Exposure/duration/ frequency</th>
<th>System</th>
<th>NOAEL (mg/m³)</th>
<th>LOAEL Less serious (mg/m³)</th>
<th>Serious (mg/m³)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Gn Pig</td>
<td>2 wk</td>
<td>Resp</td>
<td>440 M</td>
<td>(mild to slight pneumonia in 3/16 animals; pulmonary granulomas in 7/16)</td>
<td></td>
<td>Laong et al. 1987</td>
</tr>
<tr>
<td>(Albino Hartley)</td>
<td>5 d/wk</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>4 hr/d</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Hepatic</td>
<td>440 M</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Renal</td>
<td>440 M</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Dermal</td>
<td>440 M</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Ocular</td>
<td></td>
<td></td>
<td>440 M (degeneration of photoreceptor cells in retina)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bd Wt</td>
<td>440 M</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*The number corresponds to entries in Figure 2-1.

Bd Wt = Body Weight; d = day(s); Gn Pig = guinea pig; hr = hour; LOAEL = lowest-observed-adverse-effect level; M = male; NOAEL = no-observed-adverse-effect level; Resp = respiratory; wk = week(s)
Figure 2-1. Levels of Significant Exposure to 4,4-Methylenedianiline - Inhalation
Acute (≤14 days)

Systemic

(mg/m³)

1000

100

10

1

Respiratory

Hepatic

Renal

Dermal

Ocular

Body Weight

1g

1g

1g

1g

1g

1g

Key

g guinea pig

- LOAEL for serious effects (animals)

- LOAEL for less serious effects (animals)

- NOAEL (animals)

The number next to each point corresponds to entries in Table 2-1.
Ocular Effects. The role of melanin in ocular toxicity of 4,4′-methylenedianiline was examined in guinea pigs (Leong et al. 1987). Two strains were used, albino (lacking melanin) and pigmented. The animals were exposed nose-only to 440 mg/m³ of an aerosol of 4,4′-methylenedianiline 4 hours per day, 5 days per week for 2 weeks. Two weeks after exposure terminated, the animals were sacrificed and the retinas examined. The retinas of both strains showed marked alterations ranging from retraction and thickening of the outer segments of the photoreceptor cells to swelling and retraction extended through the inner segments of the photoreceptors to the outer nuclear layer. There were also degenerative changes in the pigmented epithelial cells. Since the retinal lesions were similar in both strains, the authors concluded that these changes were not related to the affinity of 4,4′-methylenedianiline for melanin. No further information was located regarding ocular effects after inhalation exposure to 4,4′-methylenedianiline.

Body Weight Effects. Guinea pigs exposed nose-only to 440 mg/m³ of an aerosol of 4,4′-methylenedianiline 4 hours per day, 5 days per week for 2 weeks experienced a slight weight loss during exposure days, but recovery was apparent during the 2 resting weekend days (Leong et al. 1987). The authors attributed this temporary loss in weight to the stress of being restrained during exposure since the trend over the entire experimental period was similar between exposed and control animals. No further information was located regarding body weight effects after inhalation exposure to 4,4′-methylenedianiline.

No studies were located regarding the following health effects in humans or animals after inhalation exposure to 4,4′-methylenedianiline:

2.2.1.3 Immunological and Lymphoreticular Effects
2.2.1.4 Neurological Effects
2.2.1.5 Reproductive Effects
2.2.1.6 Developmental Effects
2.2.1.7 Genotoxic Effects

Genotoxicity studies are discussed in Section 2.5.
2.2.1.8 Cancer

No studies were located regarding carcinogenic effects in humans or animals after inhalation exposure to 4,4′-methylenedianiline.

2.2.2 Oral Exposure

2.2.2.1 Death

No cases of human deaths attributed to oral exposure to 4,4′-methylenedianiline were located in the literature reviewed. The only relevant information was found in a study that examined the potential long-term health effects in a population that had consumed bread contaminated with 4,4′-methylenedianiline in 1965 in the Epping district of Essex, England (Hall et al. 1992). Liver toxicity was the main adverse effect reported at the time of the accident (Kopelman et al. 1966). Of the original 84 cases, 55 people were alive, 18 had died, and 16 could not be traced (Hall et al. 1992). The causes of death (neoplastic and non-neoplastic diseases) were, by and large, unremarkable, and the observed/expected ratios for death from all causes were well below 1.0. Thus, there was no obvious relationship between ingestion of 4,4′-methylenedianiline in that particular episode and death in humans.

Several studies have reported death in animals after oral administration of 4,4′-methylenedianiline. In Wistar rats, oral LD₅₀ values of 335 mg/kg (Schmidt et al. 1980) and 830 mg/kg (Pludro et al. 1969) were reported. In the former study, the test material was administered in propylene glycol, whereas peanut oil was used as vehicle in the latter. Two of 5 male mice died in a 14-day study after receiving daily doses of 207 mg 4,4′-methylenedianiline/kg/day in the drinking water, 1 of 5 females died at a 220 mg/kg/day dose level; all males and females (5/5) died at 829 mg/kg/day and 882 mg/kg/day, respectively (NTP 1983). The cause of death was not reported. The LD₅₀ values in male guinea pigs and male rabbits administered 4,4′-methylenedianiline in peanut oil were 260 mg/kg and 620 mg/kg, respectively (Schmidt et al. 1974). In an intermediate-duration study, female Sprague-Dawley rats were treated by gavage intermittently for 30 days with 36 mg 4,4′-methylenedianiline/kg/day in sesame oil (Griswold et al. 1968). Forty-five days after treatment started, survival in treated rats was reduced 16% relative to untreated controls; the cause of death was not reported. In a chronic-duration study, survival rate was reduced by approximately 20% in male B6C3F₁ mice.
administered doses of 57 mg 4,4′-methylenedianiline/kg/day in the drinking water for 103 weeks (Lamb et al. 1986; NTP 1983).

LOAEL and LD$_{50}$ values for death for each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.2 Systemic Effects

Little information is available regarding systemic effects in humans after oral exposure to 4,4′-methylenedianiline. In contrast, numerous studies have examined the effects of oral administration of 4,4′-methylenedianiline in animals, particularly in rats. The overall evidence suggests that the liver and perhaps the thyroid are target organs for 4,4′-methylenedianiline toxicity.

The highest NOAEL values and all reliable LOAEL values for systemic effects for each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

Respiratory Effects. Abnormal respiratory rhythm was reported in a man on arrival to the hospital after drinking an unspecified amount of a liquid containing 4,4′-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). Because of the simultaneous ingestion of other chemicals, the role of 4,4′-methylenedianiline, if any, in causing this respiratory effect cannot be ascertained.

No gross or histopathogical alterations were observed in the lungs, trachea, bronchi, or nasal cavity of rats administered up to 141 mg 4,4′-methylenedianiline/kg/day in drinking water for 13 weeks or up to 19 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice administered up to 116 mg 4,4′-methylenedianiline/kg/day in the drinking water for 13 weeks or up to 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). An earlier study reported lung congestion and hyperemia, pneumonia, and pulmonary edema in female beagle dogs treated with doses of approximately 2.7 mg 4,4′-methylenedianiline/kg/day for 54-84 months (Deichmann et al. 1978). The test material was dissolved in corn oil and administered in a gelatin capsule 3 times per week. Because this study used only a total of 9 animals and no concurrent controls were-used, the validity of the findings is unclear.
<table>
<thead>
<tr>
<th>Key to figure</th>
<th>Species (Strain)</th>
<th>Exposure/duration/frequency (Specific route)</th>
<th>System</th>
<th>NOAEL (mg/kg/day)</th>
<th>Less serious (mg/kg/day)</th>
<th>Serious (mg/kg/day)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Rat (Wistar)</td>
<td>once (G)</td>
<td></td>
<td></td>
<td></td>
<td>830 (LD₅₀)</td>
<td>Pludro et al. 1969</td>
</tr>
<tr>
<td>2</td>
<td>Rat (Wistar)</td>
<td>once (GO)</td>
<td></td>
<td></td>
<td></td>
<td>335 M (LD₅₀)</td>
<td>Schmidt et al. 1980</td>
</tr>
<tr>
<td>3</td>
<td>Mouse (B6C3F1)</td>
<td>14 d (ad libitum) (W)</td>
<td></td>
<td></td>
<td></td>
<td>207 M (2/5 died)</td>
<td>NTP 1983</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>220 F (1/5 died)</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Gn Pig (NS)</td>
<td>once (GO)</td>
<td></td>
<td></td>
<td></td>
<td>260 M (LD₅₀)</td>
<td>Schmidt et al. 1974</td>
</tr>
<tr>
<td>5</td>
<td>Rabbit (NS)</td>
<td>once (GO)</td>
<td></td>
<td></td>
<td></td>
<td>620 M (LD₅₀)</td>
<td>Schmidt et al. 1974</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>Rat (Sprague-Dawley)</td>
<td>once (GO)</td>
<td>Hepatic</td>
<td></td>
<td></td>
<td>25⁵ M (increased serum alanine aminotransferase and relative liver weight)</td>
<td>Baille et al. 1993</td>
</tr>
<tr>
<td>7</td>
<td>Rat (Sprague-Dawley)</td>
<td>once (GO)</td>
<td>Hepatic</td>
<td></td>
<td></td>
<td>50 M (cholestasis, biliary epithelial injury, hepatic parenchymal damage)</td>
<td>Baille et al. 1994</td>
</tr>
<tr>
<td>8</td>
<td>Rat (Sprague-Dawley)</td>
<td>once (G)</td>
<td>Gastro</td>
<td>250 M</td>
<td></td>
<td>250 M (necrosis of bili duct epithelial cells, focal periporal hepatocellular necrosis)</td>
<td>Kanz et al. 1992</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Hepatic</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Key to figure</td>
<td>Species (Strain)</td>
<td>Exposure/duration/ frequency (Specific route)</td>
<td>System</td>
<td>NOAEL (mg/kg/day)</td>
<td>LOAEL</td>
<td></td>
<td></td>
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<tr>
<td>---------------</td>
<td>-----------------</td>
<td>---------------------------------------------</td>
<td>--------</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>Rat (Fischer-344)</td>
<td>14 d (ad libitum) (W)</td>
<td>Gastro</td>
<td>130 F</td>
<td>235 M</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bd Wt</td>
<td>57 M</td>
<td>65 F</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>117 M (10% lower final body</td>
<td>130 F weight)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>Rat (Wistar)</td>
<td>once (GO)</td>
<td>Hepatic</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>50 M (bile duct necrosis; increased serum AP, ALT, glutamate dehydrogenase, and leucine aminopeptidase)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>11</td>
<td>Rat (Sprague-Dawley)</td>
<td>5-14 d 1x/d (GW)</td>
<td>Endocr</td>
<td>110 F</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bd Wt</td>
<td>110 F</td>
<td>(17% reduction in final body weight)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>Mouse (B6C3F1)</td>
<td>14 d (ad libitum) (W)</td>
<td>Bd Wt</td>
<td>207 M</td>
<td>220 F</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>415 M</td>
<td>441 F</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Immunological/Lymphoreticular</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>Rat (Sprague-Dawley)</td>
<td>once (G)</td>
<td></td>
<td></td>
<td>250 M (disintegration of cortical thymocytes; cytolysis in cortical lymphocytes)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Reproductive</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>14</td>
<td>Rat (Sprague-Dawley)</td>
<td>5-14 d 1x/d (GW)</td>
<td></td>
<td></td>
<td>110 F (increased weight of the uterus; atypical folliculoid endometrial response)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
### Table 2-2. Levels of Significant Exposure to 4,4'-Methyleneedianiline - Oral (continued)

<table>
<thead>
<tr>
<th>Key to figure</th>
<th>Species (Strain)</th>
<th>Exposure/duration/frequency (Specific route)</th>
<th>System</th>
<th>NOAEL (mg/kg/day)</th>
<th>Less serious (mg/kg/day)</th>
<th>Serious (mg/kg/day)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>15</td>
<td>Rat (Sprague-Dawley)</td>
<td>30 d 1 x/3d (GO)</td>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>16</td>
<td>Rat (Wistar)</td>
<td>8-40 wk (F)</td>
<td>Hepatic</td>
<td>92 M (hyperplasia of bile ducts, increased relative liver weight)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Renal</td>
<td>92 M</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bd Wt</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>17</td>
<td>Rat (Fischer-344)</td>
<td>34 wk (ad lib) (W)</td>
<td>Hepatic</td>
<td>88 M (bile duct proliferation)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Renal</td>
<td>88 M</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Endocr</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bd Wt</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>18</td>
<td>Rat (Fischer-344)</td>
<td>32 wk (ad lib) (F)</td>
<td>Hepatic</td>
<td>100 M (bile duct proliferation)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>(F)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>19</td>
<td>Rat (Fischer-344)</td>
<td>8 wk (F)</td>
<td>Hepatic</td>
<td>100 M (bile duct proliferation; fatty changes)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Endocr</td>
<td>100 M (hyperplastic goiter of the thyroid)</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**INTERMEDIATE EXPOSURE**

**Death**

36 F (16% decreased survival)  
Griswold et al. 1968

92 M (44% reduction in weight gain after 40 weeks of treatment)  
Fukushima et al. 1979

88 M (40% reduction in body weight gain)  
Fukushima et al. 1981
<table>
<thead>
<tr>
<th>Key to figure</th>
<th>Species (Strain)</th>
<th>Exposure/duration/frequency (Specific route)</th>
<th>System</th>
<th>NOAEL (mg/kg/day)</th>
<th>Serious (mg/kg/day)</th>
<th>Less serious (mg/kg/day)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>20 Rat (Inbred W)</td>
<td>19 wk (F)</td>
<td>Endocr</td>
<td></td>
<td>84 M (thyroid hyperplasia)</td>
<td></td>
<td>79 M (27% reduction in body weight gain)</td>
<td>Hiasa et al. 1984</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Bd Wt</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>21 Rat (Wistar)</td>
<td>12 wk (F)</td>
<td>Hepatic</td>
<td></td>
<td>84 M (bile duct proliferation; fatty infiltration; fibrosis; increased SGOT, SGPT, AP, and BSP retention)</td>
<td></td>
<td>84 M (58% reduction in body weight gain)</td>
<td>Miyamoto et al. 1977</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Bd Wt</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>22 Rat (Fischer-344)</td>
<td>13 wk (ad libitum) (W)</td>
<td>Resp</td>
<td>141 F</td>
<td></td>
<td></td>
<td></td>
<td>NTP 1983</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Cardio</td>
<td>141 F</td>
<td></td>
<td></td>
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<tr>
<td></td>
<td></td>
<td>Gastro</td>
<td>141 F</td>
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<tr>
<td></td>
<td></td>
<td>Musc/skel</td>
<td>141 F</td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td></td>
<td></td>
<td>Hepatic</td>
<td>33 M 35 F</td>
<td>67 M (bile duct hyperplasia in 4/10 males and 3/10 females)</td>
<td>70 F</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Renal</td>
<td>141 F</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Endocr</td>
<td>33 M 35 F</td>
<td>67 M (hyperplastic goiter in 3/10 males and 1/10 females)</td>
<td>70 F</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Dermal</td>
<td>141 F</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Bd Wt</td>
<td>67 M 70 F</td>
<td></td>
<td></td>
<td></td>
<td>134 M (21% reduced final body weight in males, 26% in females)</td>
</tr>
<tr>
<td>Key to figure</td>
<td>Species (Strain)</td>
<td>Exposure/duration/ frequency (Specific route)</td>
<td>System</td>
<td>NOAEL (mg/kg/day)</td>
<td>Less serious (mg/kg/day)</td>
<td>Serious (mg/kg/day)</td>
<td>Reference</td>
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<td>-------------------------</td>
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<td>-----------</td>
</tr>
<tr>
<td>23</td>
<td>Rat (Wistar)</td>
<td>12 wk 1 x/d (G)</td>
<td>Gastro</td>
<td>8.3</td>
<td></td>
<td>83 (intestinal occlusion)</td>
<td>Pludo et al. 1969</td>
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<td>Hemato</td>
<td>83</td>
<td></td>
<td>83 (atrophy of hepatocytes; increased relative liver weight)</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Hepatic</td>
<td>8.3 c</td>
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<td>Renal</td>
<td>83</td>
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<td>Bd Wt</td>
<td>83</td>
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<tr>
<td>24</td>
<td>Rat (Fischer-344)</td>
<td>24 wk (F)</td>
<td>Endocr</td>
<td>97M (hyperplastic goiter)</td>
<td></td>
<td></td>
<td>Tsuda et al. 1987</td>
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<tr>
<td></td>
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<td>Bd Wt</td>
<td></td>
<td></td>
<td>97 M (70% reduction in body weight gain)</td>
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<tr>
<td>25</td>
<td>Mouse (B6C3F1)</td>
<td>13 wk (ad libitum) (W)</td>
<td>Resp</td>
<td>116 F</td>
<td></td>
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<tr>
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<td>Cardio</td>
<td>116 F</td>
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<td>116 F</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Musc/skel</td>
<td>116 F</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Hepatic</td>
<td>58 F</td>
<td></td>
<td>108M (bile duct hyperplasia in 5/10)</td>
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<td></td>
<td></td>
<td>Renal</td>
<td>116 F</td>
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<td></td>
<td></td>
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<td>Endocr</td>
<td>116 F</td>
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<td>Dermal</td>
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<td></td>
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<td></td>
<td>Bd Wt</td>
<td>54 M 116 F</td>
<td></td>
<td>108M (13% reduction in final body weight)</td>
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<td>Immunological/Lymphoreticular</td>
<td>26</td>
<td>Rat (Fischer-344) (ad libitum) (W)</td>
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<td>141 F</td>
<td></td>
<td></td>
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<td>Species (Strain)</td>
<td>Exposure/duration/ frequency (Specific route)</td>
<td>System</td>
<td>LOAEL (mg/kg/day)</td>
<td>Less serious (mg/kg/day)</td>
<td>Serious (mg/kg/day)</td>
<td>Reference</td>
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</tr>
<tr>
<td>27 Mouse</td>
<td>(B6C3F1)</td>
<td>13 wk (ad libitum) (W)</td>
<td></td>
<td>116 F</td>
<td></td>
<td></td>
<td>NTP 1983</td>
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<td>Neurological</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>28 Rat</td>
<td>(Fischer- 344)</td>
<td>13 wk (ad libitum) (W)</td>
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<td>141 F</td>
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<td>NTP 1983</td>
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<td>29 Mouse</td>
<td>(B6C3F1)</td>
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<td>NTP 1983</td>
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<td>Reproductive</td>
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</tr>
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<td>30 Rat</td>
<td>(Fischer- 344)</td>
<td>13 wk (ad libitum) (W)</td>
<td></td>
<td>141 F</td>
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<td>NTP 1983</td>
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<td>31 Mouse</td>
<td>(B6C3F1)</td>
<td>13 wk (ad libitum) (W)</td>
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<td>116 F</td>
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Table 2-2. Levels of Significant Exposure to 4,4'-Methylenedianiline - Oral (continued)

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<th>Species (Strain)</th>
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<th>NOAEL (mg/kg/day)</th>
<th>Less serious (mg/kg/day)</th>
<th>Serious (mg/kg/day)</th>
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<td>Mouse (B6C3F1)</td>
<td>103 wk (ad libitum) (W)</td>
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<td></td>
<td></td>
<td>57 M (20% reduced survival rate)</td>
<td>Lamb et al. 1986; NTP 1993</td>
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<tr>
<td>33</td>
<td>Rat (Fischer- 344)</td>
<td>103 wk (ad libitum) (W)</td>
<td>Resp</td>
<td>19 F</td>
<td></td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Cardio</td>
<td>19 F</td>
<td></td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Gastro</td>
<td>19 F</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Musc/skel</td>
<td>19 F</td>
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<td></td>
<td></td>
<td></td>
<td>Hepatic</td>
<td></td>
<td>9M (fatty metamorphosis, focal cellular change)</td>
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<td></td>
<td></td>
<td></td>
<td>Renal</td>
<td>9 M 19 F</td>
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<td></td>
<td></td>
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<td>Endocr</td>
<td>10 F</td>
<td>19 F (mineralization of the kidney)</td>
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<tr>
<td></td>
<td></td>
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<td>Dermal</td>
<td>19 F</td>
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<td></td>
<td></td>
<td></td>
<td>Bd Wt</td>
<td>19 F</td>
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<td>Key to figure</td>
<td>Species (Strain)</td>
<td>Exposure/ duration/ frequency (Specific route)</td>
<td>System</td>
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<td>Less serious (mg/kg/day)</td>
<td>Serious (mg/kg/day)</td>
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<tr>
<td>34</td>
<td>Mouse (B6C3F1)</td>
<td>103 wk (ad libitum) (W)</td>
<td>Resp</td>
<td>57 M</td>
<td></td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Cardio</td>
<td>57 M</td>
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<tr>
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<td></td>
<td>Gastro</td>
<td>57 M</td>
<td></td>
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<td>Musc/skel</td>
<td>57 M</td>
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<td>Hepatic</td>
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<td>25 M (liver degeneration)</td>
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<td></td>
<td></td>
<td>Endocr</td>
<td>25 M</td>
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<tr>
<td>35</td>
<td>Rat (Fischer- 344)</td>
<td>103 wk (ad libitum) (W)</td>
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<td>19 F</td>
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<td>Mouse (B6C3F1)</td>
<td>103 wk (ad libitum) (W)</td>
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<td>57 M</td>
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<tr>
<td>37</td>
<td>Rat (Fischer- 344)</td>
<td>103 wk (ad libitum) (W)</td>
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<td>19 F</td>
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<tr>
<td>38</td>
<td>Mouse (B6C3F1)</td>
<td>103 wk (ad libitum) (W)</td>
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<td>57 M</td>
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</table>

LOAEL:

- 25 M (nephropathy)
- 19 F

25 M (13-16% reduction in final body weight)
<table>
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<tr>
<th>Key to figure</th>
<th>Species (Strain)</th>
<th>Exposure/ duration/ frequency (Specific route)</th>
<th>System</th>
<th>LOAEL</th>
<th>Serious (mg/kg/day)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>39</td>
<td>Rat (Fischer-344)</td>
<td>103 wk (ad libitum) (W)</td>
<td></td>
<td>19 F</td>
<td></td>
<td>Lamb et al. 1986; NTP 1983</td>
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<tr>
<td>40</td>
<td>Mouse (B6C3F1)</td>
<td>103 wk (ad libitum) (W)</td>
<td></td>
<td>57 M</td>
<td></td>
<td>NTP 1983; Lamb et al. 1986</td>
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</tbody>
</table>

**Reproductive**

**Cancer**

<table>
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<th>Key to figure</th>
<th>Species (Strain)</th>
<th>Exposure/ duration/ frequency (Specific route)</th>
<th>System</th>
<th>LOAEL</th>
<th>Serious (mg/kg/day)</th>
<th>Reference</th>
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<tbody>
<tr>
<td>41</td>
<td>Rat (Fischer-344)</td>
<td>103 wk (ad libitum) (W)</td>
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<td>9 M (CEL: increased incidence of neoplastic nodules in liver)</td>
<td>Lamb et al. 1986; NTP 1983</td>
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<td>42</td>
<td>Mouse (B6C3F1)</td>
<td>103 wk (ad libitum) (W)</td>
<td></td>
<td>10 F</td>
<td>(CEL: malignant lymphoma and adenoma/carcinoma of the liver)</td>
<td>Lamb et al. 1986; NTP 1983</td>
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</tbody>
</table>

a The number corresponds to entries in Figure 2-2.
b Used to derive an acute oral Minimal Risk Level (MRL) of 0.2 mg/kg/day; unadjusted dose divided by an uncertainty factor of 300 (3 for use of a minimal LOAEL, 10 for extrapolation from rats to humans, and 10 for human variability). A modifying factor of 0.5 was used to account for facilitated absorption by the corn oil vehicle.
c Used to derive an intermediate-duration oral MRL of 0.08 mg/kg/day; unadjusted dose divided by an uncertainty factor of 100 (10 for extrapolation from rats to humans and 10 for human variability)

ALT = alanine amino transferase; AP = alkaline phosphatase; Bd Wt = body weight; BSP = bromosulphalein; (C) = capsule; Cardio = cardiovascular; CEL = cancer effect level; d = day(s); Derm = dermal; Endocr = endocrine; F = female; (F) = feed; (G) = gavage; Gastro = gastrointestinal; (GO) = gavage (oil); (GW) = gavage (water); Hemato = hematological; LD<sub>50</sub> = lethal dose, 50% kill; LOAEL = lowest-observed-adverse-effect level; M = male; mo = month(s); Musc/skel = musculoskeletal; NOAEL = no-observed-adverse-effect level; Resp = respiratory; SGOT = serum glutamic oxaloacetic transaminase; SGPT = serum glutamic pyruvic transaminase; (W) = water; wk = week(s); x = time(s)
Figure 2-2. Levels of Significant Exposure to 4,4-Methyleneedianiline - Oral
Acute (≤14 days)

Systemic

(mg/kg/day)

Key

- LD₅₀ (animals)
- LOAEL for serious effects (animals)
- LOAEL for less serious effects (animals)
- NOAEL (animals)
- CEL: cancer effect level (animals)
- Minimal risk level for effects other than cancer

* Doses represent the lowest dose tested per study that produced a tumorigenic response and do not imply the existence of a threshold for the cancer end point.
Figure 2-2. Levels of Significant Exposure to 4,4-Methyleneedianiline - Oral (cont.)

Intermediate (15-364 days)

Systemic

(mg/kg/day)

<table>
<thead>
<tr>
<th>Death</th>
<th>Respiratory</th>
<th>Cardiovascular</th>
<th>Gastrointestinal</th>
<th>Hematological</th>
<th>Muscularkeletal</th>
<th>Hepatic</th>
<th>Renal</th>
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</tbody>
</table>

Key

- r: rat
- m: mouse
- g: guinea pig
- h: rabbit
- LD<sub>50</sub> (animals)
- LOAEL for serious effects (animals)
- LOAEL for less serious effects (animals)
- NOAEL (animals)
- CEL: cancer effect level (animals)
- Minimal risk level for effects other than cancer
- The number next to each point corresponds to entries in Table 2-2.

* Doses represent the lowest dose tested per study that produced a tumorigenic response and do not imply the existence of a threshold for the cancer endpoint.
Figure 2-2. Levels of Significant Exposure to 4,4-Methyleneedianiline - Oral (cont.)
Intermediate (15-364 days)

Systemic

(mg/kg/day)

Key

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<tbody>
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<tr>
<td>g</td>
<td>guinea pig</td>
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<tr>
<td>h</td>
<td>rabbit</td>
</tr>
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<td>■</td>
<td>LD_{50} (animals)</td>
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<td>LOAEL for serious effects (animals)</td>
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<td>○</td>
<td>NOAEL (animals)</td>
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<td>CEL: cancer effect level (animals)</td>
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</table>

* Doses represent the lowest dose tested per study that produced a tumorigenic response and do not imply the existence of a threshold for the cancer end point.
Figure 2-2. Levels of Significant Exposure to 4,4-Methyleneedianiline - Oral (cont.)

Chronic (≥365 days)

Systemic

(mg/kg/day)

Death  Respiratory  Cardiovascular  Gastrointestinal  Musculoskeletal  Hepatic  Renal  Endocrine  Dermal  Body Weight  Immunological/Lymphocytotoxic  Neurological  Reproductive  Cancer*

Key

r  rat  ■ LD₅₀ (animals)
m  mouse  ● LOAEL for serious effects (animals)
g  guinea pig  ○ LOAEL for less serious effects (animals)
h  rabbit  O NOAEL (animals)
  ◇ CEL: cancer effect level (animals)

* Doses represent the lowest dose tested per study that produced a tumorigenic response and do not imply the existence of a threshold for the cancer end point.
Cardiovascular Effects. Very limited and inconclusive information exists regarding cardiovascular effects in humans after ingestion of 4,4´-methyleneedianiline. Bradycardia, hypotension, and abnormal electrocardiogram were reported in a male subject who accidentally drank an undetermined amount of a solution containing 4,4´-methyleneedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). These signs were observed on arrival to the hospital shortly after the accident occurred. Because of the simultaneous ingestion of other chemicals, the role of 4,4´-methyleneedianiline, if any, cannot be determined.

Information on effects in animals is also limited. No gross or histopathological alterations were observed in the heart of rats administered up to 141 mg 4,4´-methyleneedianiline/kg/day in drinking water for 13 weeks or up to 19 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice administered up to 116 mg 4,4´-methyleneedianiline/kg/day in the drinking water for 13 weeks or up to 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983).

Gastrointestinal Effects. Nausea, abdominal pain, and vomiting were reported in one female and five males (ages 17-25) who drank an alcoholic beverage spiked with 4,4´-methyleneedianiline (Tillmann et al. 1997); the amount of 4,4´-methyleneedianiline ingested was not known. No further information was located regarding gastrointestinal effects in humans following oral exposure to 4,4´-methyleneedianiline.

Doses of ≥261 mg 4,4´-methyleneedianiline/kg/day administered in the drinking water for 14 days induced crater-like foci in the cardiac portion of the stomach in female rats (NTP 1983). The NOAEL for this effect in females was 130 mg/kg/day. No such lesions were seen in males treated with up to 235 mg/kg/day for the same period of time (NTP 1983), but lesions were evident at 469 mg/kg/day. Longer-duration studies reported no gross or histopathological alterations in the salivary glands, esophagus, stomach, pancreas, duodenum, jejunum, ileum, and colon from rats administered up to 141 mg 4,4´-methyleneedianiline/kg/day in drinking water for 13 weeks or up to 19 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice administered up to 116 mg 4,4´-methyleneedianiline/kg/day in the drinking water for 13 weeks or up to 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Intestinal occlusion was reported in an earlier study in rats treated by gavage with 83 mg 4,4´-methyleneedianiline/kg/day for 12 weeks; the NOAEL was 8.3 mg/kg/day (Pludro et al. 1969). No further information was provided in this study.
**Hematological Effects.** Limited information was located regarding hematological effects in humans after oral exposure to 4,4´-methylenedianiline. A male subject developed eosinophilia with left shift in neutrophils 7-35 days after accidentally ingesting a solution containing a 4,4´-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). This is consistent with the appearance of erythema multiform which is characterized by eosinophilia. A recent study reports that an 18-year-old male had mild leucocyte elevation 1 day after drinking an alcoholic beverage spiked with 4,4´-methylenedianiline (Tillmann et al. 1997); his blood cell count and thrombocyte rate were normal. The amount of 4,4´-methylenedianiline ingested was not known.

The data in animals are limited to a study that reported no alterations in hemoglobin levels or erythrocyte counts in rats treated daily by gavage for 12 weeks with doses of 83 mg 4,4´-methylenedianiline/kg/day (Pludro et al. 1969).

**Musculoskeletal Effects.** One female and five males (ages 17-25) complained of muscle and joint pain after drinking an alcoholic beverage spiked with 4,4´-methylenedianiline (Tillmann et al. 1997). No further information was located.

Very limited information was found regarding musculoskeletal effects of 4,4´-methylenedianiline in animals after oral exposure. No gross or histopathogical alterations were observed in thigh muscle and costochondral junction (rib) of rats administered up to 141 mg 4,4´-methylenedianiline/kg/day in drinking water for 13 weeks or up to 19 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice administered up to 116 mg 4,4´-methylenedianiline/kg/day in the drinking water for 13 weeks or up to 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983).

**Hepatic Effects.** A local outbreak of jaundice, which was later traced to ingestion of 4,4´-methylenedianiline, occurred in the Epping district of Essex, England, in 1965 (Kopelman et al. 1966). Eighty-four subjects became ill shortly after eating bread prepared with flour that had been contaminated with 4,4´-methylenedianiline. Three general types of clinical presentations were observed. In a majority of the patients, the illness had an acute onset with severe intermittent pain in the upper abdomen and lower chest for 24-36 hours. Over the next few days, the patients in this group improved, but then developed a flu-like condition with fever and increasing jaundice. The liver was enlarged and tender. After a few days, the liver became smaller, but the jaundice persisted for weeks. These patients did not feel completely recovered for a considerable period of time. A second
group of patients only showed mild symptoms of upper abdominal discomfort. A week later, however, they too developed fever and increasing jaundice, which persisted longer than in the first group. A third group, with the least common symptoms, had severe jaundice when first seen, but had minimal preceding symptoms. The liver in these patients was often enlarged, but rarely tender. Clinical chemistry tests showed increases in serum bilirubin, alkaline phosphatase activity, and glutamic oxaloacetic transaminase. Needle biopsy performed in 4 cases within 2-3 weeks of the onset of symptoms showed cellular infiltration and cholestasis, and there was evidence of damage to the liver parenchyma and biliary tree. This, according to the investigators (Kopelman et al. 1966), is the first documented case of human poisoning with 4,4’-methylenedianiline. Forty-three of these subjects were evaluated 2 years later (Kopelman 1968). Aside from slight abnormalities in single liver tests and complaints of subjective nature, there was no evidence of progressive hepatic disease.

Liver toxicity was also observed in the case of a male subject who accidentally drank an unspecified amount of a solution containing 4,4’-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). Clinical tests conducted 2 days after admission to the hospital showed elevated transaminases and hyperbilirubinemia. Slight hepatomegaly developed 6 weeks after admission. One year after the accident, serum transaminases still had not returned to normal levels. Although other chemicals were involved in this case, the signs and symptoms are consistent with those reported by Kopelman et al. (1966) and were most likely caused by 4,4’-methylenedianiline. A case of a young man who drank an alcoholic beverage spiked with 4,4’-methylenedianiline and developed liver toxicity was recently described in the literature (Tillmann et al. 1997). The subject thought that the substance was methylenedioxyamphetamine, a psychoactive drug. Upon admission to the hospital, 24 hours after drinking the beverage, his bilirubin and liver enzyme activities were elevated and increased steadily over 7 days; he also developed jaundice. By the time he was discharged on day 15, the jaundice had vanished and the bilirubin was close to normal. Tillmann et al. (1997) indicates that five other subjects, who also drank the spiked beverage, developed a similar picture of liver toxicity.

Numerous studies of various durations have demonstrated that the liver is a target for 4,4’-methylenedianiline toxicity in animals, particularly in rats. A single gavage dose of 25 mg/kg (range tested 25-225 mg/kg) increased serum alanine aminotransferase activity and relative liver weight in rats (Bailie et al. 1993). Higher single doses (50-250 mg/kg) induced cholestasis, biliary epithelial injury, bile duct necrosis, and periportal hepatocellular necrosis (Bailie et al. 1993, 1994; Kanz et al. 1992; Schmidt et al. 1980). The earliest change identified was bile ductular necrosis 4 hours after dosing (Bailie et al. 1993). These single-
dose studies demonstrated that 4,4’-methylenedianiline is selectively toxic to bile duct in rats and that hepatic lesions appear after the lesions to the bile ducts (Kanz et al. 1992). The minimal effective dose of 25 mg/kg from the Bailie et al. (1993) study is considered a minimal LOAEL and is the basis for derivation of an acute oral MRL of 0.2 mg/kg/day.

Results from intermediate-duration studies support those from single-dose studies. LOAEL values in the range of 67-100 mg 4,4´-methylenedianiline/kg/day have been identified in rats (Fukushima et al. 1979, 1981; Hagiwara et al. 1993; Miyamoto et al. 1977; NTP 1983; Pludro et al. 1969). However, in many of these studies only one dose level was used. Exceptions are the NTP (1983) study in which NOAELs of 35 mg/kg/day and 58 mg/kg/day were identified in rats and mice, respectively, and the earlier report by Pludro et al. (1969) that established a NOAEL of 8.3 mg/kg/day in rats. This NOAEL, 8.3 mg/kg/day, served as the basis for derivation of an intermediate oral MRL of 0.08 mg/kg/day. Gavage, drinking water, or diet were used as vehicles in the studies mentioned above, which suggests that the method of administration of 4,4´-methylenedianiline is not a determining factor in liver toxicity. In addition to elevated serum transaminases, the most commonly seen liver alterations were hyperplasia of the bile ducts, fatty infiltration, fibrosis, and atrophy of the liver parenchyma. In general, it appeared that most of the hepatic lesions were at least partially reversible following cessation of treatment.

Liver dilation, fatty metamorphosis, and focal cellular change were described in rats treated with 9 mg 4,4´-methylenedianiline/kg/day in the drinking water for 103 weeks (Lamb et al. 1986; NTP 1983). In the same study, liver degeneration was seen in mice receiving 25 mg/kg/day. Both dose levels represent the lowest levels tested. In a study in dogs, all 9 of the treated animals had liver lesions that included hepatic cell necrosis, fatty infiltration, and portal fibrosis after treatment with approximately 2.7 mg 4,4´-methylenedianiline/kg/day, 3 days per week for 54-84 months (Deichmann et al. 1978). Because of study limitations, such as a small number of animals used and lack of concurrent controls, it is not possible to conclusively determine whether dogs are more sensitive than rodents.

**Renal Effects.** Only one report was located that described adverse renal effects in humans following oral exposure to 4,4´-methylenedianiline. In this case report (Roy et al. 1985), a male subject accidentally drank an unspecified amount of a liquid containing 4,4´-methylenedianiline, potassium carbonate, and gamma-butyrolactone. Tests conducted two days after the accident showed hematuria and glycosuria. In the presence of normoglycemia, glycosuria indicated renal tubular
dysfunction. Because there was simultaneous ingestion of other chemicals, the role of 4,4′-methyleneedianiline, if any, cannot be conclusively determined. Proteinuria and erythrocyturia were described in a young man who drank an alcoholic beverage spiked with 4,4′-methylenedianiline (Tillmann et al. 1997). No further information was located.

Several reports have investigated renal effects of 4,4′-methylenedianiline in animals after intermediate and chronic-duration exposure. No gross or histopathological alterations have been reported in the kidneys or urinary bladder of rats treated with 4,4′-methylenedianiline in the range of 83-141 mg/kg/day for periods ranging between 8-40 weeks (Pukushima et al. 1979, 1981; NTP 1983; Pludro et al. 1969). Administration vehicles included gavage, drinking water, and diet. Similar lack of effects were reported in mice treated with up to 116 mg 4,4′-methylenedianiline/kg/day in the drinking water for 13 weeks (NTP 1983). In contrast, high incidence of kidney mineralization was observed in male rats treated for 103 weeks with dose of 16 mg 4,4′-methylenedianiline/kg/day in the drinking water (Lamb et al. 1986; NTP 1983). No such effect was observed in females treated with a similar dose (Lamb et al. 1986; NTP 1983). A high incidence of nephropathy was reported in male and female mice treated with 19-25 mg 4,4′-methylenedianiline/kg/day in the drinking water for 103 weeks (Lamb et al. 1986; NTP 1983). Male mice also exhibited renal papilla mineralization at a dose level of 57 mg/kg/day (Lamb et al. 1986; NTP 1983). Various lesions to the kidney and urinary bladder were observed in 9 dogs treated with approximately 2.7 mg 4,4′-methylenedianiline/kg/day, 3 days per week for 54-84 months (Deichmann et al. 1978). Kidney abnormalities included rough surface, congestion, glomerulonephritis, cloudy swellin,, and thickening of the basement membrane. Hyperemia in the urinary bladder was noticed in two dogs, whereas mucosal hyperplasia, edema, lymphocytic infiltration, and marked congestion of the urinary bladder were observed in another dog. This study has severe limitations, such as a very small number of animals (9 female beagle dogs), no concurrent controls, and only one dose level was tested; therefore, the results must be interpreted with caution.

Endocrine Effects. No studies were located regarding endocrine effects in humans after oral exposure to 4,4′-methylenedianiline.

Numerous studies in rats have identified the thyroid as a sensitive organ for 4,4′-methylenedianiline toxicity. Hypertrophy and histopathological alterations of the adrenals and thyroid were reported in rats administered doses of 110-146 mg 4,4′-methylenedianiline/kg/day by gavage for 5-14 days.
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(Tullner 1960). Longer duration studies (8-32 weeks) in rats have established LOAELs for thyroid effects in the range of 67-100 mg 4,4’-methylenedianiline/kg/day (Fukushima et al. 1981; Hagiwara et al. 1993; Hiasa et al. 1984; NTP 1983; Tsuda et al. 1987). The administration vehicle was drinking water or food. With the exception of the NTP (1983) study, only one dose level was tested in these studies. In the NTP (1983) report, a NOAEL of 35 mg/kg/day was identified for rats and 116 mg/kg/day for mice; the latter was the highest dose level tested in mice. The thyroid alterations observed consisted of hyperplasia, decrease of colloid in the follicles, reduced follicle size, and slight reduction in serum T₃ and T₂. In addition to thyroid effects, pituitary basophile hypertrophy was noticed in male and female rats that received approximately 140 mg 4,4’-methylenedianiline/kg/day in the drinking water for 13 weeks (NTP 1983). Results from chronic-duration studies revealed follicular cysts and follicular cell hyperplasia in the thyroid of female rats treated with 19 mg 4,4’-methylenedianiline/kg/day in the drinking water for 103 weeks and thyroid cell hyperplasia in male and female mice treated in the same manner with 57 mg/kg/day and 43 mg/kg/day, respectively (Lamb et al. 1986; NTP 1983).

The overall evidence suggests that thyroid, particularly in rats, may be a sensitive organ for 4,4’-methylenedianiline toxicity. However, until other animal species are tested, it is uncertain whether rats are the most sensitive species.

Dermal Effects. Limited relevant data in humans were located. Roy et al. (1985) described the case of a man who developed erythema multiform after accidentally ingesting a solution containing 4,4’-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). This finding is consistent with an allergic reaction to 4,4’-methylenedianiline (see Section 2.2.3). In a recent case report, an 18-year-old male developed a skin rash 7 days after drinking an alcoholic beverage spiked with 4,4’-methylenedianiline (Tillmann et al. 1997). The rash had cleared by the time he was discharged from the hospital on day 15. No further information was located.

Very limited information was found regarding dermal effects of 4,4’-methylenedianiline in animals after oral exposure. No gross or histopathological alterations were observed in the skin of rats administered up to 141 mg 4,4’-methylenedianiline/kg/day in drinking water for 13 weeks or up to 19 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice administered up to 116 mg 4,4’-methylenedianiline/kg/day in the drinking water for 13 weeks or up to 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983).
Ocular Effects. Ocular effects were described in the case of a male subject who accidentally ingested an unspecified amount of a solution containing 4,4’-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). Four days after admission to the hospital, his vision became blurred and visual acuity was reduced considerably. This condition worsened on subsequent weeks and he developed a coarse pigmented retinopathy similar to that of retinitis pigmentosa. Tests conducted later revealed gross malfunction of the retinal pigment epithelium, a condition which improved little over the next 18 months. Although the subject ingested a mixture of three components, the retina is not known to be a target for gamma-butyrolactone or potassium carbonate toxicity. Nevertheless, there is no conclusive evidence that the effects observed were caused by 4,4’-methylenedianiline.

No studies were located regarding ocular effects in animals after oral exposure to 4,4’-methylenedianiline.

Body Weight Effects. No studies were located regarding body weight effects in humans following oral exposure to 4,4’-methylenedianiline.

Oral exposure to 4,4’-methylenedianiline in animals usually resulted in dose-related reduction in body weight gain, and occasionally, weight loss. Male rats exposed for 14 days in the drinking water to 235 mg 4,4’-methylenedianiline/kg/day had a 31% reduction in final body weight relative to untreated controls (NTP 1983). At the lowest dose level tested, 117 mg/kg/day in males and 130 mg/kg/day in females, final body weights were reduced by about 11%. In mice treated in the same manner, the NOAEL and LOAEL for body weight effects was 220 mg/kg/day and 415 mg/g/day, respectively (NTP 1983). Numerous intermediate-duration studies (8-40 weeks) have reported decreased body weight gain in rats treated with 4,4´-methylenedianiline doses in the range of 84 mg/kg/day to 141 mg/kg/day (Fukushima et al. 1979, 1981; Hagiwara et al. 1993; Hiasa et al. 1984; Miyamoto et al. 1977; NTP 1983; Tsuda et al. 1987). The administration vehicle varied between food and drinking water. In these studies, final body weights were reduced 27-70% relative to untreated controls. The sole exception is a report by Pludro et al. (1969) which identified a NOAEL of 83 mg/kg/day in rats treated by gavage for 12 weeks and no explanation is apparent for this discrepancy. In the studies mentioned above, except for the NTP (1983) study, only one dose level of 4,4´-methylenedianiline was tested. The NOAEL for rats in the NTP (1983) report was 70 mg/kg/day. In mice, the NOAEL and
LOAEL were 54 mg/kg/day and 108 mg/kg/day, respectively (NTP 1983). None of the studies mentioned above provided data regarding food consumption.

Body weights from rats treated for 103 weeks with up to 19 mg 4,4′-methylenedianiline/kg/day in the drinking water were not significantly different than untreated controls (Lamb et al. 1986; NTP 1983). However, in the same study (Lamb et al. 1986; NTP 1983), final body weight was reduced by 13% in male mice treated with 57 mg 4,4′-methylenedianiline/kg/day and by 16% in females treated with 43 mg 4,4′-methylenedianiline/kg/day. The NOAEL was about 20 mg/kg/day. Again, food consumption data were not provided.

2.2.2.3 Immunological and Lymphoreticular Effects

No studies were located regarding immunological and lymphoreticular effects in humans after oral exposure to 4,4′-methylenedianiline.

Data in animals are restricted to histopathological examinations of organs of the lymphoreticular system, but no information is available regarding possible effects on immunocompetence. Focal disintegration of cortical thymocytes was observed in rats 8 hours after a single gavage dose of 250 mg 4,4′-methylenedianiline/kg (Kanz et al. 1992). In some rats, approximately 50% of the thymus cortex was necrotic 24 hours after dosing. However, no histopathological alterations were observed in the spleen (Kanz et al. 1992). A 13-week drinking water study reported no histopathological alterations in the spleen, thymus, and lymph nodes of rats treated with up to 141 mg 4,4′-methylenedianiline/kg/day (NTP 1983). The same findings were reported in mice treated in the same manner with up to 116 mg 4,4′-methylenedianiline/kg/day (NTP 1983). The results from the NTP (1983) study are in conflict with those of Pludro et al. (1969) who reported unspecified lesions in the spleen in rats treated with daily gavage doses of 8.3 mg 4,4′-methylenedianiline/kg. Doses of 83 mg/kg/day induced hyperplasia in lymphatic nodes. A possible explanation for the discrepancy between the results from these two studies is the use of different administration vehicles, drinking water in the NTP (1983) and gavage in propylene glycol in the Pludro et al. (1969) study.

There were no histopathological changes in the spleen, thymus, and lymph nodes from rats treated for 103 weeks with up to 19 mg 4,4′-methylenedianiline/kg/day in the drinking water (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice treated with up to 57 mg 4,4′-methylene-
dianiline/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). In dogs treated for 54-84 months with approximately 2.7 mg 4,4’-methyleneedianiline/kg/day (dose given 3 times/week by gavage in a capsule) the spleens appeared shrunken and the surface had a granular appearance (Deichmann et al. 1978). These latter investigators also reported splenitis with thickening and hyalinization of the capsule, trabeculae and lymphoid corpuscles. Hemosiderosis and spleen congestion, which were also noticed, may have been secondary to hematological effects (not reported) such as hemolytic anemia and methemoglobinemia. This study in dogs was poorly conducted and poorly reported; thus, it is unclear whether dogs are a particularly sensitive species.

The highest NOAEL values and all reliable LOAEL values for immunological and lymphoreticular effects for each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.4 Neurological Effects

No studies were located regarding neurological effects in humans after oral exposure to 4,4’-methyleneedianiline.

Information on neurological effects in animals is very limited. No gross or histopathological alterations were observed in the sciatic nerve, brain, and spinal cord of rats administered up to 141 mg 4,4’-methyleneedianiline/kg/day in drinking water for 13 weeks or up to 19 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice administered up to 116 mg 4,4’-methyleneedianiline/kg/day in the drinking water for 13 weeks or up to 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). No further neurological parameters were evaluated. The limited information available suggests that 4,4’-methyleneedianiline is not a neurotoxicant. The highest NOAEL values and all reliable LOAEL values for neurological effects for each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.5 Reproductive Effects

No studies were located regarding reproductive effects in humans after oral exposure to 4,4’-methyleneedianiline.
Very little information exists regarding reproductive effects of 4,4´-methyleneedianiline in animals. An acute-duration study reported a 71% increase in absolute weight of the uterus in ovariectomized rats administered 110-146 mg 4,4´-methylenedianiline by gavage for 5-14 days (Tullner 1960). Histopathological examination of the uterus revealed an atypical folliculoid response in the endometrium. No further information was provided in this study. No gross or histopathological alterations were observed in the ovaries, uterus, mammary glands, seminal vesicles, prostate, or testes of rats administered up to 141 mg 4,4´-methylenedianiline/kg/day in drinking water for 13 weeks or up to 19 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Similar findings were reported in mice administered up to 116 mg 4,4´-methylenedianiline/kg/day in the drinking water for 13 weeks or up to 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). No further reproductive parameters were evaluated. The limited information available is insufficient to determine whether exposure to 4,4´-methylenedianiline may alter reproductive function.

The highest NOAEL values and all reliable LOAEL values for reproductive effects for each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.6 Developmental Effects

No studies were located regarding developmental effects in humans after oral exposure to 4,4´-methylenedianiline. Only one report was located that provided information on effects in animals. In that study (Bourdelat et al. 1983), fetuses from pregnant rats treated by gavage with 37 mg 4,4´-methylenedianiline/kg/day (as the chlorohydrate) on gestation days 14-20 had liver alterations in the form of fatty infiltration of the parenchyma. This dose level also caused histological alterations in the livers from the dams. Fetuses from 1 of 5 dams administered 219 mg 4,4´-methylenedianiline/kg/day on gestation days 7-20 showed delayed closing of the calvaria, enlarged tongue, and an abnormally large snout (Bourdelat et al. 1983). The 219 mg/kg/day dose level was lethal to 1 of 5 pregnant rats. Because of study limitations such as the use of only one female rat as control and lack of detailed reporting of the results, this study is not presented in Table 2-2.

2.2.2.7 Genotoxic Effects

No studies were located regarding genotoxic effects in humans after oral exposure to 4,4´-methylenedianiline.
Administration of 4, 20, or 50 mg 4,4´-methylenedianiline/kg/day by gavage for 3 days to male Fischer 344 rats resulted in the dose-related formation of adducts with liver DNA, as detected by 32Ppostlabeling analysis (Vock et al. 1996). However, DNA isolated from the bladder and from lymphocytes from these animals did not show any treatment-related DNA-adducts.

Other genotoxicity studies are discussed in Section 2.5.

2.2.2.8 Cancer

A recent study examined the causes of death (neoplastic and non-neoplastic diseases) in a population that had consumed bread contaminated with 4,4´-methylenedianiline in 1965 in the Epping district of Essex, England (Hall et al. 1992). Liver toxicity was the main adverse effect reported at the time of the accident (Kopelman et al. 1966). Of the original 84 cases, 55 people were alive, 18 had died, and 16 could not be traced. Of those alive, 58% completed a health questionnaire. The causes of death were, by and large, unremarkable, with the possible exception of one case of biliary duct carcinoma. The observed/expected ratios for cancer and non-neoplastic diseases were well below 1.0. The results suggested that there was no obvious link between current health status and the poisoning episode. The one case of biliary duct carcinoma was of interest because this tumor, according to the investigators (Hall et al. 1992), is very rare in humans.

The carcinogenic potential of oral administration of 4,4´-methylenedianiline has been examined in dogs (Deichmann et al. 1978), rats, and mice (Griswold et al. 1968; Lamb et al. 1986; NTP 1983). No significant carcinogenic response was reported in rats treated for 30 days with 36 mg 4,4´-methylenedianiline/kg/day by gavage and observed for 9 months (Griswold et al. 1968); however, the observation period may have been too short. Dogs that received 5-6.26 grams 4,4´-methylenedianilinelkg for a period of 54-84 months (the time-weighted dose can be estimated at about 2.7 mg/kg/day) did not show a significant increase in bladder or liver tumors (Deichmann et al. 1978).

This study has severe limitations such as a very small number of animals (9 female beagle dogs), no concurrent controls, and practically only one dose level was tested; therefore, the results must be interpreted with caution. In a 2-year bioassay (Lamb et al. 1986; NTP 1983), Fischer 344 rats were treated with 4,4´-methylenedianiline (as the dihydrochloride) in the drinking water. Doses were 9 and 16 mg/kg/day in males and 10 and 19 mg/kg/day in females. Clear evidence of carcinogenicity was found. In male rats, the incidence of follicular cell carcinomas of the thyroid gland was 0 of
49 (controls), 0 of 47 (low dose), and 7 of 48 (high dose); the incidence of neoplastic nodules of the liver was 1 of 50 (controls), 12 of 50 (low dose), and 25 of 50 (high dose). In females, the incidence of follicular cell adenomas was 0 of 47 (controls), 2 of 47 (low dose), and 17 of 48 (high dose); the incidence of C-cell adenomas was 0 of 47 (controls), 3 of 47 (low dose), and 6 of 48 (high dose). Tumors were also found in other tissues, but the increased incidence was not statistically significant.

4,4´-Methylenedianiline was also carcinogenic in B6C3F1 mice when administered in the drinking water for 103 weeks (Lamb et al. 1986; NTP 1983). The doses were 25 and 57 mg/kg/day for males and 19 and 43 mg/kg/day for females. In males, the incidence of follicular adenomas of the thyroid gland was 0 of 47 (controls), 3 of 49 (low dose), and 16 of 49 (high dose); carcinomas of the liver occurred at an incidence of 10 of 49 (controls), 33 of 50 (low dose), and 29 of 50 (high dose). Also in males, the incidence of pheochromocytoma of the adrenal gland was 2 of 48 (controls), 12 of 49 (low dose), and 14 of 49 (high dose). In females, the following significant increased incidences were observed: follicular cell adenomas of the thyroid gland (0 of 50 controls, 1 of 47 low dose, 13 of 50 high dose), carcinomas/adenomas of the liver (4 of 50 controls, 15 of 50 low dose, 23 of 50 high dose), malignant lymphoma (13 of 50 controls, 28 of 50 low dose, 29 of 50 high dose), and alveolar/bronchiolar adenoma (1 of 50 controls, 2 of 50 low dose, 6 of 49 high dose).

Several studies have examined the effects of 4,4´-methylenedianiline on post-initiation stage carcinogenicity in various organs in rats. For example, administration of 100 mg 4,4´-methylenedianiline/kg/day in the diet for 32 weeks following initiation with N-ethyl-N-hydroxyethylnitrosamine reduced the incidence of hyperplastic nodules in the liver and neoplastic nodules in the kidney induced by the nitrosamine alone (Fukushima et al. 1981). Similar findings were reported regarding neoplastic responses in the urinary bladder when 4,4´-methylenedianiline (88 mg/kg/day for 34 weeks) followed initiation with N-butyl-N-(4-hydroxybutyl) nitrosamine (Fukushima et al. 1981), and regarding hepatocellular carcinomas when 4,4´-methylenedianiline (100 mg/kg/day for 26 weeks) followed initiation with diethylnitrosamine plus 2-acetylamino fluorene (Masui et al. 1986). In these studies, 4,4´-methylenedianiline alone was not carcinogenic. In contrast with the results summarized above, the incidence of thyroid tumors in rats initiated with N-bis(2-hydroxypropyl)nitrosamine and then treated with 4,4´-methylenedianiline (84 mg/kg/day for 19 weeks) was significantly higher (90%) than in rats treated only with the initiator (28%) (Hiasa et al. 1984); no tumors were seen in rats treated with 4,4´-methylenedianiline alone.
In a different type of experiment, administration of 100 mg 4,4´-methylenedianiline/kg/day for 8 weeks to rats followed by treatment with a combination of 3 carcinogens for 4 weeks resulted in a lower incidence of follicular cell hyperplasia and adenomas of the thyroid relative to rats treated with only the carcinogens (Hagiwara et al. 1993). The incidence of preneoplastic/neoplastic lesions observed in other tissues and organs was similar in the two groups.

The long-term bioassays conducted in rats and mice (Lamb et al. 1986; NTP 1983) provide clear evidence of 4,4´-methylenedianiline carcinogenicity in rodents. However, no evidence of carcinogenicity was found in intermediate-duration studies in the groups of rats that were treated with 4,4´-methylenedianiline alone (Fukushima et al. 1981; Hiasa et al. 1984; Masui et al. 1986). These are not necessarily inconsistent results, since exposure duration and observation periods may have been too short in the intermediate-duration studies.

The dose of levels of 9 mg 4,4´-methylenedianiline/kg/day for male rats and 19 mg/kg/day for female mice from the Lamb et al. (1986) and NTP (1983) studies are listed as Cancer Effect Levels (CEL) in Table 2-2 and are plotted in Figure 2-2.

2.2.3 Dermal Exposure

2.2.3.1 Death

No studies were located regarding death in humans following dermal exposure to 4,4´-methylenedianiline. Very limited information exists regarding death in animals after dermal exposure to 4,4´-methylenedianiline.

Four of 9 female mice and 1 of 9 males died after having doses of 168 mg 4,4´-methylenedianiline/kg/day in methanol applied to the clipped skin 5 days per week for 2 weeks (Holland et al. 1987). When the solvent was acetone, 3 of 10 males and 3 of 10 females died. The authors indicated that using acetone as solvent may have provided inaccurate results since 4,4´-methylenedianiline tends to form a Schiff base with acetone. The same group of investigators (Holland et al. 1987) reported a significant dose-related decrease in survival rate in mice applied ≥5.3 mg 4,4´-methylenedianiline/kg/day 3 times per week for 104 weeks. The 24-month survival rate was approximately 60%
in untreated controls and 35.9% in treated mice. LOAEL values for death for each species and duration category are recorded in Table 2-3.

2.2.3.2 Systemic Effects

No studies were located regarding respiratory effects in humans or musculoskeletal effects in humans or animals after dermal exposure to 4,4´-methylenedianiline.

The highest NOAEL values and all reliable LOAEL values for systemic effects for each species and duration category are recorded in Table 2-3.

Respiratory Effects. A single study in animals indicates that no gross or histopathological lesions were detected in the lungs and trachea of rabbits that received daily skin doses of 12,000 mg 4,4´-methylenedianiline/kg as an aqueous paste for 10 consecutive days (DuPont 1975). No further relevant information was located.

Cardiovascular Effects. A case of a male subject with cardiac abnormalities after dermal exposure to 4,4´-methylenedianiline was described by Brooks et al. (1979). The 20-year-old subject had worked for 2 weeks at a chemical plant where he handled large quantities of 4,4´-methylenedianiline. Exposure had occurred mostly by dermal contact with uncovered portions of the arms six days prior to admission to the hospital. Analysis of the electrocardiogram on admission showed normal waves and tests for myocardial damage were unremarkable. However, results from an echocardiogram showed reduced septal motion and reduced left ventricular function. Myocardial abnormalities were still observed three months after exposure, but not one year after the incident. No information was provided regarding possible simultaneous exposure to other chemicals.

No studies were located regarding cardiovascular effects in animals after dermal exposure to 4,4´-methylenedianiline.

Gastrointestinal Effects. No studies were located regarding, gastrointestinal effects in humans after dermal exposure to 4,4´-methylenedianiline.
### Table 2-3. Levels of Significant Exposure to 4,4'-Methyleneedianiline - Dermal

<table>
<thead>
<tr>
<th>Species (Strain)</th>
<th>Exposure/ Duration/ Frequency</th>
<th>System</th>
<th>NOAEL (mg/kg/day)</th>
<th>Serious (mg/kg/day)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ACUTE EXPOSURE</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Death</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mouse (C3H/HeJ)</td>
<td>2 wk</td>
<td></td>
<td></td>
<td>168 F (4/9 died)</td>
<td>Holland et al. 1987</td>
</tr>
<tr>
<td>Mouse (C3H/HeJ)</td>
<td>5 d/wk</td>
<td></td>
<td></td>
<td>168</td>
<td>Holland et al. 1987</td>
</tr>
<tr>
<td><strong>Systemic</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mouse (C3H/HeJ)</td>
<td>2 wk</td>
<td>Hepatic</td>
<td>84</td>
<td>168 (increased absolute liver weight)</td>
<td>Holland et al. 1987</td>
</tr>
<tr>
<td>Mouse (C3H/HeJ)</td>
<td>5 d/wk</td>
<td>Renal</td>
<td>168</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mouse (C3H/HeJ)</td>
<td>5 d/wk</td>
<td>Dermal</td>
<td>168</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mouse (C3H/HeJ)</td>
<td>5 d/wk</td>
<td>Bd Wt</td>
<td>168</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Guinea Pig</td>
<td>10 d</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Guinea Pig</td>
<td>6 x/d</td>
<td>Dermal</td>
<td>22 M</td>
<td></td>
<td>Leong et al. 1987</td>
</tr>
<tr>
<td>Rabbit (Albino)</td>
<td>10 d</td>
<td>Hepatic</td>
<td></td>
<td>700 M (bile duct proliferation, portal cirrhosis, focal parenchymal necrosis in liver)</td>
<td>DuPont 1976a</td>
</tr>
<tr>
<td>Rabbit (Albino)</td>
<td>6 x/d</td>
<td>Renal</td>
<td></td>
<td>700 M (mild acute glomerulonephritis)</td>
<td></td>
</tr>
<tr>
<td>Rabbit (Albino)</td>
<td>6 x/d</td>
<td>Dermal</td>
<td></td>
<td>700 M (acute necrotizing dermatitis)</td>
<td></td>
</tr>
<tr>
<td>Rabbit (Albino)</td>
<td>6 x/d</td>
<td>Bd Wt</td>
<td></td>
<td>700 M (15% reduction in final body weight)</td>
<td></td>
</tr>
<tr>
<td>Rabbit (NS)</td>
<td>10 d</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rabbit (NS)</td>
<td>10 d</td>
<td>Ocular</td>
<td>3.3</td>
<td>(reversible corneal opacity; conjunctivitis; iris congestion)</td>
<td>DuPont 1976b</td>
</tr>
</tbody>
</table>
Table 2-3. Levels of Significant Exposure to 4,4'-Methyleneedianiline - Dermal (continued)

<table>
<thead>
<tr>
<th>Species (Strain)</th>
<th>Exposure/duration/frequency</th>
<th>System</th>
<th>NOAEL (mg/kg/day)</th>
<th>Less serious (mg/kg/day)</th>
<th>Serious (mg/kg/day)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>CHRONIC EXPOSURE</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mouse</td>
<td>104 wk</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(C3H/Bd)</td>
<td>3 d/wk</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Death</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mouse</td>
<td>104 wk</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(C3H/Bd)</td>
<td>3 d/wk</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cancer</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mouse</td>
<td>104 wk</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(C3H/Bd)</td>
<td>3 d/wk</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Bd Wt = body weight; CEL = cancer effect level; d = day(s); Derm = dermal; F = female; LOAEL = lowest-observed-adverse-effect level; M = male; NOAEL = no-observed-adverse-effect level; NS = not specified; wk = week(s); x = time(s)

5.3 F (15.4% survival rate; 60% in controls) Holland et al. 1987
5.3 F (CEL-increased incidence of hepatic tumors) Holland et al. 1987
A single study in animals indicate that no gross or histopathological lesions were detected in the stomach and intestines of rabbits that received daily skin doses of up to 2,000 mg 4,4´-methylenedianiline/kg as an aqueous paste for 10 consecutive days (DuPont 1975). No further relevant information was located.

**Hematological Effects.** Very limited information was located regarding hematological effects in humans following dermal exposure to 4,4´-methylenedianiline. Normal blood counts were reported in a man following accidental contact with 4,4´-methylenedianiline (Van Joost et al. 1987). A second case concerned six men who came in contact with 4,4´-methylenedianiline while mixing it with an epoxy resin at work (Williams et al. 1974). Four of the six men were reported to have elevated eosinophil count, which is consistent with an immunological allergic reaction (although none was described) (see Section 2.5).

No studies were located regarding hematological effects in animals after dermal exposure to 4,4´-methylenedianiline.

**Hepatic Effects.** Several studies have described adverse hepatic effects in humans after dermal exposure to 4,4´-methylenedianiline. Thirteen cases of toxic hepatitis were reported in a factory that manufactured hard plastic (McGill and Motto 1974). 4,4´-Methylenedianiline was used as a curing agent in the process. The illness began between one and three weeks after employment started and all the reported signs and symptoms were consistent with liver disease (right upper quadrant pain and fever, jaundice, elevated transaminases, hyperbilirubinemia). Similar findings have also been described by others (Bastian 1984; Brooks et al. 1979; Williams et al. 1974). All these cases shared common signs and symptoms that included pain, elevated serum transaminases, jaundice, and hyperbilirubinemia. Although simultaneous exposure to other chemicals cannot be totally ruled out, the overall evidence and results from animal studies suggest that 4,4´-methylenedianiline was a major contributor to liver toxicity.

Limited data from animal studies suggest that the liver may also be a target for 4,4´-methylenedianiline after dermal exposure. Mice that received daily applications of 168 mg 4,4´-methylenedianiline/kg in methanol or acetone 5 days per week for 2 weeks exhibited an increase in liver relative to vehicle controls (Holland et al. 1987). No effects were seen at 84 mg/kg/day. Bile duct proliferation, portal cirrhosis, and focal parenchymal necrosis were observed in the livers of rabbits.
which received skin doses of 700 mg 4,4'-methylenedianiline/kg/day in ethanol for 10 consecutive days (DuPont 1976a). These changes were not noticed in rabbits treated with ethanol alone. In contrast, no adverse hepatic effects were noticed in the livers of rabbits also treated for 10 consecutive days with up to 2,000 mg 4,4'-methylenedianiline/kg/day, but applied as an aqueous paste (DuPont 1975), suggesting that the vehicle plays an important role in dermal absorption. No further information was located regarding hepatic effects after dermal exposure.

**Renal Effects.** No studies were located regarding renal effects in humans after dermal exposure to 4,4'-methylenedianiline.

No adverse kidney effects were reported in mice which received up to 168 mg 4,4'-methylenedianiline/kg/day in methanol or acetone applied 5 days per week for 2 weeks (Holland et al. 1987). Mild acute glomerulonephritis was reported in rabbits treated with 700 mg 4,4'-methylenedianiline/kg/day in ethanol for 10 consecutive days (DuPont 1976a). However, no such effect was noticed when 2,000 mg 4,4'-methylenedianiline/kg/day was applied as an aqueous paste (DuPont 1975), indicating that the vehicle plays a role in dermal absorption. No further information was located regarding renal effects after dermal exposure.

**Endocrine Effects.** No studies were located regarding endocrine effects in humans after dermal exposure to 4,4'-methylenedianiline.

The only information regarding endocrine effects in animals is that provided in a study in which no gross or histopathological alterations were seen in the adrenals and thyroid of rabbits which received doses of up to 2,000 mg 4,4'-methylenedianiline/kg/day as an aqueous paste to the skin for 10 consecutive days (DuPont 1975).

**Dermal Effects.** Skin rash was one of the physical findings among a group of 13 individuals who came in contact with 4,4'-methylenedianiline at work (McGill and Motto 1974). However, since there was clinical evidence of liver disease, the rash may have been another sign of toxic hepatitis and not due to direct contact with the chemical. A similar clinical picture was described by Brooks et al. (1979) in a male who handled large amounts of 4,4'-methylenedianiline at a chemical plant. Dermatitis without evidence of liver damage has also been reported (Emmett 1976; Van Joost et al. 1987). It appeared that in these cases dermal sensitization had occurred since patch testing with
4,4′-methyleneedianiline gave positive reactions. A case of photosensitivity to 4,4′-methylenedianiline was reported in a male subject who developed erythematous, pruritic dermatitis on his arms and forearms during four consecutive summers (Levine 1983). The rash appeared 60 minutes after a 30 minute exposure to sunlight even if filtered by window glass. Photopatch tests conducted with 24 contact allergens were positive for 4,4′-methylenedianiline.

No dermal irritation at the application site was observed in mice treated with up to 168 mg 4,4′-methylenedianiline/kg/day in ethanol or acetone 5 days per week for 2 weeks (Holland et al. 1987). Ten daily doses of 700 mg 4,4′-methylenedianiline/kg in ethanol produced acute necrotizing dermatitis in rabbits (DuPont 1976a). However, when the test material was applied as an aqueous paste, ten doses of 1,000 mg 4,4′-methylenedianiline/kg produced only minimal irritation (DuPont 1975). Application of a single 22 mg 4,4′-methylenedianiline/kg in polyethylene glycol to the back of guinea pigs produced neither dermal irritation nor sensitization (Leong et al. 1987). In this study, the guinea pigs had been previously exposed to an aerosol of 4,4′-methylenedianiline intermittently for 2 weeks in order to determine whether dermal sensitization to 4,4′-methylenedianiline occurred across routes of exposure.

**Ocular Effects.** No studies were located regarding ocular effects in humans after dermal exposure to 4,4′-methylenedianiline.

No gross or histopathological lesions were observed in the eyes of rabbits after receiving daily skin applications of up to 2,000 mg 4,4′-methylenedianiline/kg as an aqueous paste for 10 consecutive days (DuPont 1975). Moderate to mild reversible ocular effects were seen in the eyes of rabbits after solid 4,4′-methylenedianiline (3.3 or 33.3 mg/kg) was placed into the conjunctival sac for 20 seconds (DuPont 1976b). Effects observed included corneal opacity, congestion of the iris, and redness and swelling of the conjunctiva. The severity of the effects was dose-related and washing with water for 3.5 minutes after the 20-second treatment considerably lessened the severity.

**Body Weight Effects.** No studies were located regarding body weight effects in humans following dermal exposure to 4,4′-methylenedianiline.

No significant alterations in body weight were observed in mice treated 5 days per week for 2 weeks with up to 168 mg 4,4′-methylenedianiline/kg/day in acetone or ethanol (Holland et al. 1987). Similar
results were reported in rabbits treated with up to 2,000 mg 4,4'–methyleneedianiline/kg as an aqueous paste for 10 consecutive days (DuPont 1975). However, when the solvent was ethanol, there was a 15% reduction in final body weight after 10 days of treatment with 700 mg 4,4’-methyleneedianiline/kg/day, which suggests that ethanol facilitates dermal absorption of this chemical (DuPont 1976a).

2.2.3.3 Immunological and Lymphoreticular Effects

Several cases of dermal sensitization have been described in individuals who came in contact with 4,4’-methyleneedianiline in the workplace (Emmett 1976; Levine 1983; Van Joost et al. 1987) (see Dermal Effects). However, no information is available regarding possible effects of 4,4’-methyleneedianiline on human immunocompetence.

No dermal sensitization was observed in guinea pigs after a 2-week nose-only exposure period to 4,4’-methyleneedianiline aerosol was followed with a single topical application of up to 22 mg 4,4’-methyleneedianiline/kg (Leong et al. 1987). Increased spleen weight was reported in mice that received topical applications of up to 168 mg 4,4’-methyleneedianiline/kg/day in ethanol for 2 weeks (Holland et al. 1987); no further information was provided in that report. No gross or histopathological alterations were observed in the spleen and thymus of rabbits treated dermally with 2,000 mg 4,4’-methyleneedianiline/kg as an aqueous paste for 10 consecutive days (DuPont 1975). The information available is insufficient to draw any conclusions regarding immunological effects of 4,4’-methyleneedianiline after dermal exposure and, therefore, no entries are presented in Table 2-3.

2.2.3.4 Neurological Effects

No studies were located regarding neurological effects in humans after dermal exposure to 4,4’-methyleneedianiline.

The information regarding neurological effects in animals is limited to a study that reported no treatment-related gross or histopathological alterations in the brains of rabbits after receiving 10 consecutive daily skin applications of up to 2,000 mg 4,4’-methyleneedianiline/kg as an aqueous paste (DuPont 1975). No further neurological parameters were examined. This brief information is not considered a reliable indicator for neurological effect and, therefore, is not listed in Table 2-3.
2.2.3.5 Reproductive Effects

No studies were located regarding reproductive effects in humans following dermal exposure to 4,4’-methyleneedianiline.

The only information regarding reproductive effects in animals is that provided in a study in which no gross or histopathological alterations were seen in the testes and epididymis of rabbits which received up to 2,000 mg 4,4’-methyleneedianiline/kg/day applied as an aqueous paste to the skin for 10 consecutive days (DuPont 1975). No other reproductive parameters were evaluated. Because this information is not considered a reliable indicator of reproductive function, it is not listed in Table 2-3.

2.2.3.6 Developmental Effects

No studies were located regarding developmental effects in humans or animals after dermal exposure to 4,4’-methyleneedianiline.

2.2.3.7 Genotoxic Effects

No studies were located regarding genotoxic effects in humans or animals after dermal exposure to 4,4’-methyleneedianiline. Genotoxicity studies are discussed in Section 2.5.

2.2.3.8 Cancer

A retrospective assessment of exposure and cancer morbidity was conducted in power generation workers exposed to an epoxy resin containing 35% 4,4’-methyleneedianiline in Sweden (Selden et al. 1992). The cohort was composed of 550 males and 45 females. Based on company records, the individuals were subdivided into three categories: exposed, possibly exposed, and unexposed. Information on the cancer incidence of the cohort was obtained by computerized matching with the national cancer register for the period 1964-1985. Standardized incidence ratios (SIR) were obtained from the ratio of the observed to the expected number of cases. Exposure was considered to be primarily by the dermal route. In all three male groups, the observed number of cancers for all sites and for urinary bladder cancer was lower than the observed number. The overall SIR was 0.52 based on 5 observed cases. In the male exposed subgroup, no single cancer case appeared throughout the
observation period; the expected number was 3. Among the female workers, 2 cancer cases were identified (none in the urinary bladder); 2.7 cancer cases (all cancers) had been expected from the national rates. The authors indicate that the results should be interpreted with caution since the cohort was small, the majority of the subjects were quite young and had not reached cancer-prone age, and the follow-up period was short and may have not covered the latency period for bladder cancer (20 years).

Additional information is available from a morbidity study of employees who had worked in the gas centrifuge process at Oak Ridge Gaseous Diffusion Plant (Cragle et al. 1992). In addition to potential exposure to 4,4’-methylenedianiline, the workers may have been exposed to m-phenylenediamine, bis(2,3-epoxycyclopentylether), diglycidyl ether of bisphenol A, and the solvents trichloroethylene and methylene chloride. The cohort consisted of 263 workers who had worked closest to the process for the longest amount of time and a comparison group of 271 workers who did not work in the process. The most significant finding of the study was the report of five bladder cancers among centrifuge workers and none among the comparison worker group. However, interviews with these five workers revealed that none of them was working closely with the epoxy resin materials during any part of their employment in the process. The authors concluded that no specific agent or job duty could be identified as the causative factors for the bladder cancers (Cragle et al. 1992).

A more recent study followed a group of 10 individuals who had worked at a plant in Ontario, Canada, that manufactured an epoxy concrete surfacing material using 4,4’-methylenedianiline as a hardener (Liss and Guirguis 1994). Between 1967 and 1976 these subjects suffered acute episodes of jaundice. The study followed the group from the date of intoxication through the end of 1991 for cancer incidence by matching with the Ontario Cancer Registry. At the time of the intoxication the length of employment ranged from 7 days to 2.5 months. The results of the follow-up revealed that one case of bladder cancer developed among the group; it was diagnosed in 1990, 23 years after the intoxication. The standardized incidence ratio for the bladder cancer was 19.3 (95% confidence interval 0.5-107; p [one or more cases]=0.051). Liss and Guirguis (1994) suggested that their results should be interpreted with caution, given the small number of events, the absence of smoking histories, and the presence of other exposures.

The potential carcinogenicity of 4,4’-methylenedianiline by the dermal route was examined in mice (Holland et al. 1987). A solution of the test material in ethanol was applied to the clipped skin of
male and female C3Hf/Bd mice 3 times per week for 24 months. Positive controls were treated with benzo[a]pyrene and negative controls with vehicle alone. Estimated doses were 5.3, 10.7, and 21.3 mg 4,4’-methylenedianiline/kg/day. Treatment with 4,4’-methylenedianiline did not produce tumors at the application site, but increased the incidence of hepatic tumors in females in a dose-related manner (11% vehicle control, 22% low-dose, 25% mid-dose, 85% high-dose); however, a statistical analysis of the results was not provided. The incidence of tumors in the spleen, lungs, kidneys, and ovaries/testes did not increase relative to negative controls. According to the investigators, the C3Hf/Bd strain of mice is unusually susceptible to liver tumors and, therefore, the significance of the findings requires further study. Nevertheless, the dose of 5.3 mg 4,4’-methylenedianiline/kg/day is listed as a Cancer Effect Level in Table 2-3.

In summary, there is insufficient information to assess the potential carcinogenicity of 4,4’-methylenedianiline by the dermal route of exposure.

2. 3 TOXICOKINETICS

Data regarding toxicokinetics of 4,4’-methylenedianiline in humans are limited to information from cases of accidental ingestion of food contaminated with the chemical and cases of occupational exposure in the workplace, where dermal contact with 4,4’-methylenedianiline is considered the predominant route of exposure. Humans can absorb 4,4’-methylenedianiline by the inhalation, oral, and dermal routes of exposure. Limited data suggest that in humans the rate of absorption of 4,4’-methylenedianiline through the respiratory tract is faster than dermal absorption. There are no data regarding quantitative oral absorption in humans or animals. There are no inhalation data in animals. Limited dermal data in animals showed a higher absorption rate in rats than in guinea pigs. Furthermore, the absorption rate was a saturable process. No remarkable pattern of accumulation of 4,4’-methylenedianiline occurred in tissues of either species, although the liver seemed to have a higher concentration of 4,4’-methylenedianiline than other tissues. 4,4’-Methylenedianiline may be metabolized by cytochrome P-450 to polar metabolites that can undergo conjugation with glutathione; this is inferred from data on structurally similar chemicals. 4,4’-Methylenedianiline can also be acetylated and, in humans and animals, such metabolites have been detected in the urine. Because of the limited data available, a physiologically based pharmacokinetic (PBPK) model for 4,4’-methylenedianiline has not been developed.
2.3.1 Absorption

2.3.1.1 Inhalation Exposure

Only qualitative information is available regarding absorption of 4,4′-methyleneedianiline by the inhalation route in humans. Biological monitoring was used to assess exposure to 4,4′-methyleneedianiline in a cohort composed of 411 men employed in industries that manufactured or used 4,4′-methyleneedianiline in the United Kingdom (Cocker et al. 1994). The results showed that when exposure to 4,4′-methyleneedianiline was through inhalation, postshift urine samples had higher concentration of 4,4′-methyleneedianiline than samples taken preshift the next day. This suggested that 4,4′-methyleneedianiline is rapidly absorbed through the inhalation route and that peak excretion is reached after the end of the shift. In a recent study (Schiitze et al. 1995), 4,4′-methyleneedianiline was found in the urine of 33 workers exposed to low levels of 4,4′-methyleneedianiline. By using personal air samplers to monitor exposure, the authors found that most workers were exposed to <20 µg 4,4′-methyleneedianiline/m3, the detection limit. The metabolite of 4,4′-methyleneedianiline, N′-acetylmethyleneedianiline, was found in the urine of all but 4 workers. In addition, hemoglobin adducts of 4,4′-methyleneedianiline were found in all blood samples from the exposed workers. The possibility of dermal absorption was not discussed.

Very limited, and only indirect evidence of pulmonary absorption of 4,4′-methyleneedianiline exists in animals. Guinea pigs exposed nose-only to an aerosol of 4,4′-methyleneedianiline intermittently for 2 weeks showed retinal lesions, which were attributed to exposure to the test compound (Leong et al. 1987). This indicates that absorption through the respiratory tract had occurred. No further information was located.

2.3.1.2 Oral Exposure

In 1965, a group of 84 subjects in Epping, England, developed clinical signs consistent with toxic hepatitis shortly after eating bread prepared with flour that was later found to have been contaminated with 4,4′-methyleneedianiline (Kopelman et al. 1966). This was the first documented case of human poisoning with 4,4′-methyleneedianiline and provided clear evidence that the chemical can be absorbed from the gastrointestinal tract. No quantitative estimates of absorption were made. A later case report described a variety of toxic effects in a man who accidentally ingested a solution containing
4,4ʹ-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). Although in this case there was simultaneous ingestion of other chemicals, some clinical signs were consistent with 4,4ʹ-methylenedianiline toxicity and provide further evidence of oral absorption in humans.

Oral absorption of 4,4ʹ-methylenedianiline in animals can be inferred from the numerous reports of toxic effects after oral administration of 4,4ʹ-methylenedianiline summarized in Section 2.2.2. Gastrointestinal absorption occurred when 4,4ʹ-methylenedianiline was administered by gavage, mixed with food, or in the drinking water. However, no quantitative data exist.

Further evidence of oral absorption is provided in a study in which metabolites of 4,4ʹ-methylenedianiline were detected in the urine from rats administered a single dose of the test material (Tanaka et al. 1985). The extent of absorption was not determined.

2.3.1.3 Dermal Exposure

Indirect evidence of dermal absorption of 4,4ʹ-methylenedianiline in humans is provided by the various reports of adverse health effects observed in individuals exposed to the chemical in the workplace; these studies are summarized in Section 2.2.3. It should be mentioned, however, that in some of these cases inhalation of dusts of 4,4ʹ-methylenedianiline cannot be ruled out. Evidence of dermal absorption was also presented by Cocker et al. (1986a, 1994) who detected 4,4ʹ-methylenedianiline and/or metabolites in the urine of workers exposed primarily by the dermal route. Quantitative data were not provided in these studies. A recent study by Bnmmark et al. (1995) showed that approximately 28% of a dose of 4,4ʹ-methylenedianiline in isopropanol applied in a patch for 1 hour to the ventral skin of the forearm of male volunteers was absorbed. In that study, 4,4ʹ-methylenedianiline reached a peak in hydrolyzed plasma 3-4 hours after initiation of exposure and declined thereafter, with a time course consistent with first-order, one compartment kinetics. Additional information can be drawn from a study that used human skin in vitro (Hotchkiss et al. 1993). When the skin was unoccluded after application of 4,4ʹ-methylenedianiline in ethanol, 13% of the applied dose was detected in a receptor fluid at 72 hours. When the skin was occluded, 33% of the applied dose appeared in the receptor fluid at 72 hours. It was also observed that a considerable amount of 4,4ʹ-methylenedianiline (23-58% of the applied dose) remained within the skin at the end of the experiment. The authors noted that this finding may be of concern in terms of exposure in the
workplace, where gloves or clothing may be worn on top of the chemicals after they come in contact with the skin.

Quantitative aspects of dermal absorption have been examined in rats, guinea pigs and monkeys. A single dose of $^{14}$C-ring labeled 4,4’-methylenedianiline (2 or 20 mg/kg) in ethanol/water was applied to the back of rats and the area was covered with a cup (El-Hawari et al. 1986). After an exposure of 6 hours, a total of approximately 12% of the applied radioactivity was recovered in the urine, feces, gastrointestinal tract and tissues; 62% was recovered in a wash with soap and water, and 30% remained in the application site (total recoveries from radiotracer studies often exceed 100%). A 24-hour exposure period resulted in a combined 27% of the dose in urine, feces, gastrointestinal tract and tissues, 52% in the wash, and 25% in the application site. After a 96-hour exposure period, 55% of the dose was accounted for by the combined urine, feces, gastrointestinal tract, and tissues, 25% was in the wash and 26% in the application site. The results also showed that after washing, test material which remained within the skin continued to be absorbed. Moreover, occlusion facilitated absorption. When two dose levels were tested, the amount of radioactivity in tissues was higher after the high dose, but the proportion of the applied dose was lower than with the low dose, which suggested a dose-dependent absorption rate. Finally, a greater percentage of the dose was absorbed when the application site was washed with acetone and water 5 minutes after dosing than when washed with soap and water.

In guinea pigs subjected to the same protocol (El-Hawaii et al. 1986), approximately 3.5% of the applied dose (2 or 20 mg/kg) was recovered in the urine, feces, gastrointestinal tract and tissues immediately after a 6-hour exposure period; 81% was recovered in the wash and 11% remained in the application site. After a 96-hour exposure period, recovery in urine, feces, gastrointestinal tract and tissues amounted to 30% of the dose. Occlusion of the exposure site did not significantly affect absorption rate. In contrast with the results in rats, washing the area with soap and water prevented further absorption. On the other hand, as seen in rats, absorption rate was dose-dependent and a wash with acetone and water facilitated absorption relative to washing with soap and water.

A less comprehensive study was conducted in monkeys (El-Hawari et al. 1986). The treatment protocol was similar to that used in rats and guinea pigs, but the dose was kept in place for only 24 hours, after which time the site was washed with soap and water. As a percentage of the applied dose, the cumulative excretion of radioactivity over a 168-hour period was 18.8% in the urine and
1.9% in feces. Forty-seven percent was recovered in the wash for a total recovery of about 68%. Since tissues from monkeys were not analyzed, a 21% absorption rate represents only a lower limit. When the application site was washed with soap and water 5-10 minutes after the 24-hour exposure period, 63% was recovered in the wash; when acetone and water were used, recovery was 53%.

A study with rat skin in vitro showed that after application of 4,4´-methylenedianiline in ethanol to an unoccluded piece of skin, about 6% of the applied dose reached a receptor compartment at 72 hours (Hotchkiss et al. 1993). When the application site was occluded, absorption was enhanced, reaching 13.3% of the applied dose in 72 hours. These values are lower than those observed in experiments with human skin in vitro (Hotchkiss et al. 1993).

Dermal absorption has not been quantitated in mice, but toxic effects observed after dermal exposure, reported in studies summarized in Section 2.2.3, indicate that 4,4´-methylenedianiline is absorbed in this species.

2.3.2 Distribution

2.3.2.1 Inhalation Exposure

No studies were located regarding distribution of 4,4´-methylenedianiline in humans following inhalation exposure.

From the study by Leong et al. (1987) (Section 2.2.1) in which retinal toxicity was reported in guinea pigs exposed nose-only to an aerosol of 4,4´-methylenedianiline it could be inferred that 4,4´-methylenedianiline (or metabolites) reached the eye after pulmonary absorption. No further relevant information was located.

2.3.2.2 Oral Exposure

The fact that subjects who ate bread made with flour contaminated with 4,4´-methylenedianiline developed toxic hepatitis (Kopelman et al. 1966) provides indirect evidence that in humans, 4,4´-methylenedianiline distributes to the liver after oral exposure. In another study (Roy et al. 1985), a subject who accidentally drank a solution containing 4,4´-methylenedianiline, potassium carbonate,
and gamma-butyrolactone showed adverse effects on the liver and kidneys, and also had severe loss of visual acuity (Section 2.2.2). This could indicate that 4,4’-methylenedianiline (or metabolites) distribute to the liver, kidney and eye. However, because other chemicals were involved, the evidence is inconclusive.

Studies regarding quantitative distribution of 4,4’-methylenedianiline in tissues and organs from animals after oral exposure were not located. Nevertheless, the numerous reports that showed toxic responses in various organs after oral administration of 4,4’-methylenedianiline (summarized in Section 2.2.2) indicate that 4,4’-methylenedianiline (or metabolites) can distribute to these organs. The organs involved are the liver, kidneys, spleen, thymus, uterus, adrenal glands, and thyroid.

2.3.2.3 Dermal Exposure

Case reports of humans who developed hepatitis after dermal contact with 4,4’-methylenedianiline in the workplace provide indirect evidence that 4,4’-methylenedianiline distributes to the liver. No further information was located regarding distribution in humans after dermal exposure.

Quantitative distribution studies have been conducted in rats and guinea pigs after skin application of 14C-ring-labeled 4,4’-methylenedianiline (El-Hawari et al. 1986). Rats were applied a single dose (2 and 20 mg/kg) of the test material in ethanol/water and the dose area was covered with a cup. The dose was kept on site for 6, 24, or 96 hours. After these times, the skin was washed with soap and water and the animals were either sacrificed or returned to the cages for later sacrifice. Organs and tissues were prepared for radiochemical analysis. The results showed that the gastrointestinal tract contained the highest amounts of radioactivity at 6 (3.8%) and 24 (3%) hours, followed by the liver (2% and 1.2%, respectively). After 96 hours, both the gastrointestinal tract and liver had about 0.5% of the applied radioactivity. On a per gram basis, the liver had the highest amount of radioactivity at all times, followed by the adrenals and kidneys; this was also the case with the high dose. With the exception of the liver, preferential accumulation of 4,4’-methylenedianiline (or metabolites) was not apparent.

The experimental procedure and dose levels in guinea pigs were the same as in the rat study (El-Hawari et al. 1986). The gastrointestinal tract had the highest amount of radioactivity at 6 (1.5%) and 24 (2.8%) hours followed by the liver (0.4% and 0.5 %, respectively). After 96 hours, both the
gastrointestinal tract and liver contained about 0.5% of the applied dose. On a per gram basis, the adrenals had the highest amount of radioactivity at all times (about 3 times the liver). The distribution pattern did not appear to be dose-dependent and preferential accumulation in organs and tissues was not apparent.

2.3.2.4 Other Routes of Exposure

The distribution of 4,4´-methylenedianiline (or metabolites) has also been studied in rats and guinea pigs after a single intravenous injection (El-Hawari et al. 1986). Rats were injected 14C-ring-labeled 4,4´-methylenedianiline (2 mg/kg) in ethanol water and sacrifices were conducted 6, 24, or 96 after dosing. After 6 hours, the gastrointestinal tract had the highest amount of radioactivity (24% of the dose); this was followed by the liver (9.5%), skin (3.2%), and blood (2.8%). Twenty-four hours after the injection, the amount of radioactivity had decreased considerably in all tissues, and the liver and gastrointestinal tract had about 4% of the administered dose. Ninety-six hours after dosing, the liver had 4 or more times higher radioactivity (0.9%) than any other tissue. On a per gram basis, the liver had the highest concentration of radioactivity at all times, followed by the lungs at 6 and 24 hours and the spleen at 96 hours. Except for the liver, no preferential accumulation was apparent.

The guinea pigs were treated the same as the rats except that sacrifices were conducted only 96 hours after the injection (El-Hawari et al. 1986). As a percentage of the applied dose, the liver had the most radioactivity, about 3 times that found in blood. On a per gram basis, radioactivity was most concentrated in the spleen, followed by the liver, and preferential accumulation in these organs was suggested. Total recovery as a percentage of the dose, in blood, tissues, and gastrointestinal tract was 0.55%, 2.4% and 0.61%, respectively.

2.3.3 Metabolism

Limited information exists regarding the metabolism of 4,4´-methylenedianiline. By inference from structurally similar compounds, it has been assumed that 4,4´-methylenedianiline is oxidized to N-hydroxymethylenedianiline by the monooxygenase system (Cocker et al. 1986a; Farmer and Bailey 1989). This reaction leads to the formation of potentially toxic derivatives that may bind to cell macromolecule. The aryl hydroxylamine can be further oxidized to nitrosomethylenedianiline which can then be conjugated with glutathione and excreted in the urine. A different type of reaction is
acetylation of 4,4´-methylenedianiline to form N-acetylmethylenedianiline and N,N´ diacetylmethyl-
enedianiline. Both the mono- and di-acetylated metabolites have been identified in the urine of exposed workers (Robert et al. 1995). N-acetylmethylenedianiline has been identified in the urine from rats treated with 4,4´-methylenedianiline orally (Tanaka et al. 1985) and in the urine of humans exposed to the chemical in the workplace (Cocker et al. 1986a, 1994; Schtitze et al. 1995). This apparently represents a detoxification pathway since acetylated metabolites are not mutagenic (Cocker et al. 1986b; Tanaka et al. 1985). Results from an in vitro study in which 4,4´-methylenedianiline was incubated with rabbit liver microsomes showed that three metabolites were formed: azodiphenylmethane, azoxydiphenylmethane, and 4-nitroso-4´-aminodiphenylmethane (Kajbaf et al. 1992). According to the investigators, the latter may have been formed via a nonenzymatic reaction, whereas the former two were produced enzymatically. A schematic diagram of the metabolism of 4,4´-methylenedianiline is presented in Figure 2-3.

2.3.4 Elimination and Excretion

2.3.4.1 Inhalation Exposure

Data in humans are provided by a study which detected 4,4´-methylenedianiline (cl00 nmol/mmol creatinine) in the urine of workers exposed through inhalation of solid material or contaminated dust (Cocker et al. 1994). A recent study found levels of 4,4´-methylenedianiline from 0.013 to 2.76 nmol/L in the urine of workers exposed presumably by inhalation to low levels of 4,4´-methylenedianiline (Schtitze et al. 1995). The urinary concentration of the metabolite N´-acetylmethylenedianiline ranged from 0.045 to 23.4 nmol/L. No further information was located.

No studies were located regarding elimination and excretion of 4,4´-methylenedianiline or metabolites in animals after inhalation exposure.

2.3.4.2 Oral Exposure

No studies were located regarding elimination and excretion of 4,4´-methylenedianiline or metabolites in humans after oral exposure.

Information regarding animals is limited to a report by Tanaka et al. (1985), who demonstrated the presence of the N-acetyl and N,N´-diacetyl derivatives of 4,4´-methylenedianiline in the urine of rats
Figure 2-3. Proposed Metabolic Pathway for 4,4'-Methylenedianiline

Source: Adapted from Kajbaf et al. 1992; Cocker et al. 1986a, 1994
treated with a single gavage dose of 50 mg/kg of the test material in gum arabic. No quantitative information was provided.

2.3.4.3 Dermal Exposure

4,4′-Methylenedianiline and an N-acetyl conjugate of methylenedianiline have been identified in the urine of individuals who had dermal contact with 4,4′-methylenedianiline in the workplace (Cocker et al. 1986a, 1994). Over 300 urine samples were analyzed and in 81% the concentration of 4,4′-methylenedianiline was below the detection limit for the method (gas chromatography-mass spectrometry) (Cocker et al. 1986a). In those found to contain 4,4′-methylenedianiline, the concentration ranged from 6 to 175 nmol/mmol creatinine; the average was 26 nmol/mmol creatinine. 

N,N′-diacetylmethylenedianiline has also been found in the urine of exposed workers, although at much lower concentrations than the monoacetyl derivative (Robert et al. 1995). A recent study examined the excretion of 4,4′-methylenedianiline in seven male workers from a work site where the chemical was used as curing agent for an epoxy resin (Dalene et al. 1995). The study was conducted during 4 workdays and one weekend. Exposure was considered to be mainly dermal in spite of extensive protection measures. The excretion rate in urine ranged from 0 to 90 µmol/hour and was lower after the weekend than after workdays. On workdays, the cumulative excretion ranged from 0.04 to 1.2 µmol/day; during the weekend, the range was 0.005-0.51 µmol/day. The excretion of 4,4′-methylenedianiline in the urine correlated linearly with its concentration in blood. 4,4′-Methylenedianiline was applied to 5 volunteers in a patch to the ventral forearm for 1 hour; excretion of the compound in hydrolyzed urine reached a peak 6-11 hours after exposure and most of the 4,4′-methylenedianiline was eliminated within 24 hours (Brunmark et al. 1995). Of the dose absorbed, a median of 16% was excreted in the urine. Half-life elimination ranged from 9.2 to 19 hours in plasma and from 4.6 to 11 hours in urine. In two volunteers who received three dose levels, elimination half-lives were not dose-dependent (Brunmark et al. 1995).

Quantitative information regarding excretion of 4,4′-methylenedianiline is available from studies in rats, guinea pigs, and monkeys after skin application of 14C-ring-labeled 4,4′-methylenedianiline (El-Hawari et al. 1986). Rats were applied a single dose (2 or 20 mg/kg) of the test material in ethanol/water and the dose area was covered with a cup. The dose was kept on site for 6, 24, or 96 hours. After these times, the skin was washed with soap and water and the animals were either sacrificed or returned to the cages for later sacrifice. Urine and feces were collected at various
intervals for up to 96 hours. After a 6-hour exposure, 2.5% and 0.04% of the administered radioactivity were recovered in urine and feces, respectively. After a 24-hour exposure, urine and feces had 20% and 2.3% of the dose, respectively. The corresponding values for a 96-hour exposure were 43% and 10%. Rats in which the application site was not covered had lower amounts of radioactivity in the excreta. The distribution of radioactivity between urine and feces was not dose dependent. The results showed that the main excretory route in the rat is the urine.

In guinea pigs subjected to the same protocol (El-Hawari et al. 1986), 0.35% and 0.1% of the administered dose were recovered in urine and feces, respectively. After a 24-hour exposure, the recoveries in urine and in feces were 7.8% and 5.7%, respectively. The corresponding values for a 96-hour exposure were 10.5% and 17.6%. Treatment with the high dose also resulted in similar percentages of radioactivity in urine (2.8%) and feces (3.6%). In contrast with results in the rat, both urine and feces appear to be main excretory routes in guinea pigs.

In monkeys, the treatment protocol was similar to that used in rats and guinea pigs, but the dose was kept in place for only 24 hours, after which time the site was washed with soap and water (El-Hawari et al. 1986). Following dosing, urine and feces were collected for up to 168 hours. As a percentage of the applied dose, the cumulative excretion of radioactivity in the urine and feces over the 168-hour period was 18.8% and 1.9%, respectively, indicating that in monkeys, the urine is the main excretory route for 4,4’-methylenedianiline or metabolite(s).

2.3.4.4 Other Routes of Exposure

The pattern of excretion of 4,4’-methylenedianiline or metabolites was also examined in rats, guinea pigs, and monkeys after a single intravenous dose of 2 mg/kg of 14C-ring-labeled 4,4’-methylenedianiline (El-Hawari et al. 1986). In rats and guinea pigs, urine and feces were collected for up to 96 hours after dosing; in monkeys, excreta were collected for 168 hours. In rats, the amount of radioactivity recovered in the urine and feces 6 hours after the injection was (as a percentage of the dose) 55% and 0.3%, respectively. After 24 hours, the respective percentages were 67.4% and 21.8% and, after 96 hours, 67% and 31%. This indicates that the urine is the main excretory route and that excretion was almost complete 24 hours after the injection.
In guinea pigs, urinary and fecal excretion reached maximums of 34% and 51% of the dose, respectively, at about 48 hours after dosing, thus indicating that the feces is the main route of elimination of 4,4’-methyleneedianiline in guinea pigs (El-Hawari et al. 1986).

In monkeys, most of the dose (79%) was excreted in the urine within the first 48 hours; at this time 6.5% of the dose appeared in the feces (El-Hawari et al. 1986). Total recovery of radioactivity over a the 168-hour collection period amounted to 94% of the injected dose. As seen in rats, urine was the main route of excretion in monkeys.

2.3.5 Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models

Physiologically based pharmacokinetic (PBPK) models use mathematical descriptions of the uptake and disposition of chemical substances to quantitatively describe the relationships among critical biological processes (Krishnan et al. 1994). PBPK models are also called biologically based tissue dosimetry models. PBPK models are increasingly used in risk assessments, primarily to predict the concentration of potentially toxic moieties of a chemical that will be delivered to any given target tissue following various combinations of route, dose level, and test species (Clewell and Andersen 1985). Physiologically based pharmacodynamic (PBPD) models use mathematical descriptions of the dose-response function to quantitatively describe the relationship between target tissue dose and toxic end points.

PBPK/PD models refine our understanding of complex quantitative dose behaviors by helping to delineate and characterize the relationships between: (1) the external/exposure concentration and target tissue dose of the toxic moiety, and (2) the target tissue dose and observed responses (Andersen et al. 1987; Andersen and Krishnan 1994). These models are biologically and mechanistically based and can be used to extrapolate the pharmacokinetic behavior of chemical substances from high to low dose, from route to route, between species, and between subpopulations within a species. The biological basis of PBPK models results in more meaningful extrapolations than those generated with the more conventional use of uncertainty factors.

The PBPK model for a chemical substance is developed in four interconnected steps: (1) model representation, (2) model parametrization, (3) model simulation, and (4) model validation (Krishnan and Andersen 1994). In the early 1990s validated PBPK models were developed for a number of
toxicologically important chemical substances, both volatile and nonvolatile (Krishnan and Andersen 1994; Leung 1993). PBPK models for a particular substance require estimates of the chemical substance-specific physicochemical parameters, and species-specific physiological and biological parameters. The numerical estimates of these model parameters are incorporated within a set of differential and algebraic equations that describe the pharmacokinetic processes. Solving these differential and algebraic equations provides the predictions of tissue dose. Computers then provide process simulations based on these solutions.

The structure and mathematical expressions used in PBPK models significantly simplify the true complexities of biological systems. If the uptake and disposition of the chemical substance(s) is adequately described, however, this simplification is desirable because data are often unavailable for many biological processes. A simplified scheme reduces the magnitude of cumulative uncertainty. The adequacy of the model is, therefore, of great importance, and model validation is essential to the use of PBPK models in risk assessment.

PBPK models improve the pharmacokinetic extrapolations used in risk assessments that identify the maximal (i.e., the safe) levels for human exposure to chemical substances (Andersen and Krishnan 1994). PBPK models provide a scientifically-sound means to predict the target tissue dose of chemicals in humans who are exposed to environmental levels (for example, levels that might occur at hazardous waste sites) based on the results of studies where doses were higher or were administered in different species. Figure 2-4 shows a conceptualized representation of a PBPK model.

If PBPK models for 4,4’-methyleneedianiline exist, the overall results and individual models are discussed in this section in terms of their use in risk assessment, tissue dosimetry, and dose, route, and species extrapolations.

Very limited information exists regarding toxicokinetics of methylenedianiline; consequently, a PBPK model for 4,4’-methyleneedianiline has not yet been developed.

2.4 MECHANISMS OF ACTION

The mechanism of 4,4’-methyleneedianiline absorption in the gastrointestinal tract, lungs, or skin is not known. Also, no information is available on how 4,4’-methyleneedianiline is transported in the blood
Figure 2-4. Conceptual Representation of a Physiologically Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance

Source: adapted from Krishnan et al. 1994

Note: This is a conceptual representation of a physiologically based pharmacokinetic (PBPK) model for a hypothetical chemical substance. The chemical substance is shown to be absorbed via the skin, by inhalation, or by ingestion, metabolized in the liver, and excreted in the urine or by exhalation.
or stored in tissues. 4,4’-Methylenedianiline can undergo N-hydroxylation leading to the formation of intermediates suspected of being mutagenic and possibly carcinogenic. In contrast, N-acetylation, a reaction shown to occur in humans and animals, leads to less toxic derivatives. The liver and thyroid are targets for 4,4’-methylenedianiline toxicity in animals. Limited data indicate that the liver is also a target in humans. The existing information does not suggest route-specific toxicity.

### 2.4.1 Pharmacokinetic Mechanisms

The mechanism of absorption of 4,4’-methylenedianiline by the inhalation, oral, and dermal routes is not known. Based on the chemical properties of 4,4’-methylenedianiline (poorly soluble in water, soluble in lipids), a passive transfer process from the aqueous environment of the intestine across cell membranes can be anticipated. Limited dermal data in animals suggest that absorption by this route is a saturable process (El-Hawari et al. 1986). It was also shown in dermal studies that the administration vehicle plays a role in the extent of absorption. For example, 4,4’-methylenedianiline was more toxic to rabbits when applied in ethanol (DuPont 1976a) than when applied as an aqueous paste (DuPont 1975). The results from a study in humans exposed to 4,4’-methylenedianiline in the workplace suggested that pulmonary absorption is faster than dermal absorption (Cocker et al. 1994).

No information was located regarding the distribution of 4,4’-methylenedianiline in plasma, but it would be reasonable to assume that distribution is determined by partition among the various proteins according to lipid solubility and concentration. A first-pass effect in the liver can be expected. The limited toxicokinetics information available (El-Hawari et al. 1986) suggests that, with the exception of the liver, there is no preferential accumulation of 4,4’-methylenedianiline or metabolites in tissues or organs. No specific mechanism for storage of 4,4’-methylenedianiline or metabolites is apparent. Based on data from structurally-related compounds and from a study with 4,4’-methylenedianiline in vitro (Kajbaf et al. 1992), it seems that metabolism of 4,4’-methylenedianiline occurs in the liver. N-hydroxylation leads to N-hydroxylamine, a potentially toxic intermediate that can bind to macromolecule, or can be deactivated by conjugation with glutathione.

Humans and animals eliminate 4,4’-methylenedianiline in the urine (Cocker et al. 1986a, 1994; El-Hawaii et al. 1986; Tanaka et al. 1985). Results from a dermal study showed that rats and monkeys excrete 4,4’-methylenedianiline preferentially in the urine, whereas in guinea pigs both excretion routes are of equal importance (El-Hawari et al. 1986). The existing information on
4,4’-methyleneedianiline does not suggest route-dependent toxicity, except for dermal effects which may occur after topical application of high concentrations of 4,4’-methyleneedianiline.

2.4.2 Mechanisms of Toxicity

The mechanism of 4,4’-methyleneedianiline toxicity is not completely understood. Based on information on structurally similar compounds, many of the toxic properties of 4,4’-methyleneedianiline have been attributed to a reactive metabolic intermediate, N-hydroxymethyleneedianiline, which results from the enzymatic oxidation of 4,4’-methyleneedianiline (Cocker et al. 1986a; Lamb et al. 1986). In support of this view is the lack of genotoxicity of 4,4’-methyleneedianiline in the absence of metabolic activation (Section 2.5). On the other hand, metabolic formation of N-acetylmethyleneedianiline or N,N’-diacetylmethyleneedianiline appears to represent a detoxification pathway since these metabolites were not mutagenic (Cocker et al. 1986b; Tanaka et al. 1985).

Results from studies summarized in Section 2.2 strongly suggest that the liver is a target for 4,4’-methyleneedianiline toxicity in humans and animals (Bailie et al. 1993, 1994; Hagiwara et al. 1993; Kanz et al. 1992; Kopelman et al. 1966; Schmidt et al. 1980) and that the thyroid may also be a target for toxicity in animals (Pukushima et al. 1981; Hiasa et al. 1984; NTP 1983; Tsuda et al. 1987). Liver toxicity may be caused by a reactive electrophile formed during metabolism (Lamb et al. 1986) since the liver has the enzymes necessary for activation. Results from single-dose oral studies have shown that biliary epithelial cells are earlier toxicity targets than liver parenchymal cells (Bailie et al. 1994), but the exact mechanism involved is unknown. A recent study showed that bile is a major route of biliary epithelial cell exposure to proximate toxicants of 4,4’-methyleneedianiline (Kanz et al. 1995). This conclusion was based on the fact that rats infused through their common bile duct with bile from rats treated with 4,4’-methyleneedianiline exhibited a much greater percentage of necrosis in the common bile duct than rats infused with control bile. The mechanism of thyroid toxicity has not yet been resolved. 4,4’-Methyleneedianiline induced a slight decrease in serum T₃ and T₄ in rats (Hiasa et al. 1984). This decrease, according to the investigators (Hiasa et al. 1984), may have triggered secretion of thyroid stimulating hormone (TSH), which in turn induced thyroid hyperplasia.

4,4’-Methyleneedianiline was carcinogenic in rats and mice administered via drinking water (Lamb et al. 1986; NTP 1983). Although the mechanism of carcinogenicity is not known, some investigators
speculated that, at least for the liver, carcinogenicity may be related to the formation of a reactive metabolic intermediate which could bind to DNA (Lamb et al. 1986). However, a nongenetic mechanism of liver cancer is supported by recent data showing a relatively low potential of 4,4´-methylenedianiline to bind to rat liver DNA in vivo (Schiitze et al. 1996; Vock et al. 1996). As for thyroid cancer, some investigators believe that the goitrogenic activity of 4,4´-methylenedianiline supports a nongenetic mechanism (Lamb et al. 1986). Therefore, the appearance of liver and thyroid tumors may be the consequence of chronic tissue-damaging or tissue-stimulating effects, respectively, of 4,4´-methylenedianiline. 4,4´-Methylenedianiline inhibited neoplastic responses in the liver, kidney, and urinary bladder in initiated rats (Fukushima et al. 1981; Masui et al. 1986). The mechanism is not known but, according to the investigators, it may be related to a reduction in food consumption and, consequently, reduced growth. 4,4´-Methylenedianiline promoted the development of thyroid tumors in rats (Hiasa et al. 1984); the authors speculated that hypersecretion of TSH may have contributed to tumor formation in initiated cells.

2.4.3 Animal-to-Human Extrapolations

The limited information available shows that N-acetylation of 4,4´-methylenedianiline is a metabolic reaction shared by humans and animals. N-acetylmethylenedianiline has been detected in the urine of rats exposed by the oral route (Tanaka et al. 1985) and in humans exposed to 4,4´-methylenedianiline in the workplace (Cocker et al. 1986a, 1994). However, an attempt to discuss potential interspecies differences or similarities in 4,4´-methylenedianiline toxicity based solely on this information seems inappropriate at this time.

2.5 RELEVANCE TO PUBLIC HEALTH

Overview.

Most of the information on human health effects of 4,4´-methylenedianiline emanates from a case of ingestion of contaminated food and several reports of exposure in the workplace. The inhalation and dermal routes represent the most likely routes of exposure to 4,4´-methylenedianiline in occupational settings. Exposure to 4,4´-methylenedianiline for the general population seems unlikely. Case reports generally have many limitations, including lack of precise exposure data and presence of other compounds to which individuals may have been exposed, as well as confounding factors. The
presence of liver effects appears to be well established in workers exposed to \(4,4'\)-methylenedianiline and in subjects who ingested food contaminated with \(4,4'\)-methylenedianiline. Dermal effects also have been reported in occupationally exposed humans. There are also reports of respiratory, cardiovascular, hematological, and renal effects in exposed humans, but the evidence is not strong enough to conclusively establish cause-effect relationships. Results from one study were also inconclusive with respect to \(4,4'\)-methylenedianiline and cancer in humans. \(4,4'\)-Methylenedianiline was carcinogenic in rats and mice. Studies in animals are consistent and supportive of the human data with regard to liver effects. In addition to liver effects, the thyroid also appears to be a target for \(4,4'\)-methylenedianiline toxicity in animals. No thyroid effects have been reported in humans, but this end point has not been appropriately evaluated. Little is known about toxicokinetics of \(4,4'\)-methylenedianiline in humans. \(4,4'\)-Methylenedianiline has been detected in the urine of workers exposed to the chemical. Animals can absorb \(4,4'\)-methylenedianiline through inhalation, ingestion, or skin contact with the chemical. Methods are currently being developed to assess exposure by quantitatively determining \(4,4'\)-methylenedianiline adducts to hemoglobin. Because \(4,4'\)-methylenedianiline is a liver toxicant, individuals with compromised liver function may be at a greater risk. Based on limited data regarding environmental exposure, the most likely exposure route for populations living near hazardous waste sites is the dermal route. This route may be of concern to humans since animal studies have shown that \(4,4'\)-methylenedianiline which remains within the skin after washing with soap and water is a potential source for later absorption.

**Minimal Risk Levels for \(4,4'\)-Methylenedianiline.**

**Inhalation MRLs.**

No inhalation MRLs were derived for \(4,4'\)-methylenedianiline due to lack of human and animal data.

**Oral MRLs.**

- An MRL of 0.2 mg/kg/day has been derived for acute oral exposure (14 days or less) to \(4,4'\)-methylenedianiline.

The acute oral MRL is based on a minimal LOAEL of 25 mg/kg for liver toxicity in rats administered a single dose of 0 (controls), 25, 50, 75, 100, 125, or 225 mg \(4,4'\)-methylenedianiline/kg by gavage in corn
oil (Bailie et al. 1993). The minimal effective dose was between 25 and 75 mg/kg. The severity of the effects was dose-related. Effects observed included increased serum alanine aminotransferase and gamma-glutamyl transferase, increased serum bilirubin, decreased bile flow, and increased relative liver weight. All these effects were indicative of hepatic parenchymal injury. A dose of 100 mg/kg caused hepatocellular necrosis, bile ductular necrosis, portal edema, and neutrophil infiltration. Bile ductular necrosis was observed 4 hours after dosing, and this was the earliest change identified. These findings are supported by those of Bailie et al. (1994) and Schmidt et al. (1980), who also observed liver damage in rats after a single dose of 50 mg 4,4´-methyleneedianiline. It has also been shown that the liver is a target for 4,4´-methyleneedianiline toxicity in humans by the oral and dermal routes, and in animals by the dermal route.

- An MRL of 0.08 mg/kg/day has been derived for intermediate oral exposure (15-364 days) to 4,4´-methyleneedianiline.

The intermediate oral MRL is based on a NOAEL of 8.3 mg/kg/day for liver effects in rats administered daily doses of 4,4´-methyleneedianiline by gavage in propylene glycol for 12 weeks (Pludro et al. 1969). Effects observed at a higher dose of 83 mg/kg/day included increased relative weight of the liver and kidney, atrophy of the liver parenchyma with hyperplasia of the stroma, particularly at portal areas, and increased serum beta-globulin and decreased albumin. Many studies have identified LOAELs in the range of 67-100 mg/kg/day, which is consistent with the results of Pludro et al. (1969). However, most of these studies tested only one dose level. A study conducted by NTP (1983) defined hepatic NOAELs of 35 mg/kg/day and 58 mg/kg/day for rats and mice, respectively.

No chronic oral MRL was derived for 4,4´-methyleneedianiline. The lowest LOAEL for a number of endpoints was 2.7 mg/kg/day in a study in which dogs were treated for 54-84 months with 4,4´-methyleneedianiline in a capsule 3 days per week (Deichmann et al. 1978). However, this study was not well-designed and was poorly reported, which greatly diminished the potential significance of the results. No other study in dogs that could support the results of Deichmann et al. (1978) was located.

**Death.** No studies were located that reported deaths in humans attributable to exposure to 4,4´-methyleneedianiline. In a population that had consumed bread made with flour contaminated with
4,4’-methyleneedianiline in 1965 (Kopelman et al. 1966) the causes of death over the next 20 years were unremarkable (Hall et al. 1992). Observed/expected ratios for specific illnesses were well below 1.0. Data from acute, intermediate, and chronic-duration oral exposure in animals suggest that mice are more sensitive than rats (Griswold et al. 1968; Lamb et al. 1986; NTP 1983). Decreased survival rates were observed in mice treated with 36 mg 4,4’-methyleneedianiline kg/day for 30 days (Griswold et al. 1968) and in mice treated with 57 mg/kg/day for 103 weeks (Lamb et al. 1986; NTP 1983). Mice were also more sensitive to acute dermal doses of 4,4’-methyleneedianiline than rabbits (DuPont 1976a; Holland et al. 1987). Although environmental data are limited, it is unlikely that 4,4’-methyleneedianiline levels near hazardous waste sites are sufficient to cause death in exposed populations after single or few exposures. The limited data in animals are inconclusive in determining whether prolonged low-level exposures in humans may represent a health hazard.

**Systemic Effects.**

**Respiratory Effects.** No respiratory effects were observed in a population that became ill after ingesting bread contaminated with 4,4’-methyleneedianiline (Kopelman et al. 1966), and no studies were located that described respiratory effects in subjects exposed to 4,4’-methyleneedianiline in the workplace. The respiratory system was not a target for 4,4’-methyleneedianiline toxicity in animals by the inhalation (Leong et al. 1987), oral (NTP 1983) or dermal (DuPont 1975) route. For populations near Superfund sites, the potential for developing adverse respiratory effects from exposure to 4,4’-methyleneedianiline seems unlikely.

**Cardiovascular Effects.** Lingering myocardial abnormalities were reported in a subject that had dermal contact with 4,4’-methyleneedianiline in the workplace during a period of 2 weeks (Brooks et al. 1979). However, no information was provided regarding possible simultaneous exposure to other chemicals or past medical history. Furthermore, no acute or chronic signs of cardiovascular disease were observed in a population that was exposed to 4,4’-methyleneedianiline by eating contaminated bread (Kopelman et al. 1966; Roy et al. 1985). The cardiovascular system was not a target for 4,4’-methyleneedianiline toxicity in animals exposed orally in intermediate- or chronic-duration studies (Lamb et al. 1986; NTP 1983). There is insufficient evidence to assess the risk of cardiovascular disease in populations exposed to low levels of 4,4’-methyleneedianiline for a prolonged period of time.
**Gastrointestinal Effects.** No acute or chronic signs of gastrointestinal disease were observed in a population that ate bread contaminated with 4,4´-methylenedianiline (Kopelman et al. 1966; Roy et al. 1985); however, nausea, abdominal pain, and vomiting were experienced by a subject who ingested 4,4´-methylenedianiline in an alcoholic beverage (Tillmann et al. 1997). Gross abnormalities were observed in the stomach of female rats treated with $\geq 261$ mg 4,4´-methylenedianiline in the drinking water for 14 days (NTP 1983). However, the significance of this finding is unclear since this effect was not seen in males treated in a similar manner with doses $\geq 235$ mg/kg/day, although lesions were evident at 469 mg/kg/day (NTP 1983). Oral intermediate or chronic-duration studies in animals showed no adverse effects in the gastrointestinal tract after administration of 4,4´-methylenedianiline. The evidence in animals suggest that the gastrointestinal tract is not a target for 4,4´-methylenedianiline toxicity, but the human data are inconclusive as to the potential effects of prolonged oral exposure to 4,4´-methylenedianiline.

**Hematological Effects.** Eosinophilia was described in a subject who accidentally ingested a solution containing 4,4´-methylenedianiline (Roy et al. 1985). Other components in the solution were potassium carbonate and gamma-butyrolactone. Mild leucocyte elevation was reported in a young man who ingested an undetermined amount of 4,4´-methylenedianiline mixed in an alcoholic beverage (Tillmann et al. 1997). Eosinophilia was also observed in four of six men who became in contact with 4,4´-methylenedianiline at work (Williams et al. 1974). Eosinophilia is a characteristic response observed in allergic reactions; therefore, in these cases it was most likely caused by exposure to 4,4´-methylenedianiline, even though other chemicals may have been involved. No significant hematological effects have been reported in animals after exposure to 4,4´-methylenedianiline; however, this system has not been appropriately evaluated. The exact role of eosinophils in the allergic immunological reaction is not known. The existing information suggests that eosinophilia may develop after exposure to 4,4´-methylenedianiline as may occur in populations living near hazardous waste sites.

**Musculoskeletal Effects.** The only information regarding musculoskeletal effects in humans after exposure to 4,4´-methylenedianiline is from a case report of a young man who complained of joint and muscle pain on admission to the hospital one day after drinking an alcoholic beverage spiked with an unknown amount of 4,4´-methylenedianiline (Tillmann et al. 1997). Long-term oral studies in both rats and mice found no evidence of gross or histopathological alterations in the musculoskeletal system.
(Lamb et al. 1986; NTP 1983). Based on this limited evidence, no predictions can be made regarding musculoskeletal effects in populations exposed to 4,4´-methyleneedianiline.

**Hepatic Effects.** It is well established that 4,4´-methyleneedianiline is a liver toxicant in humans (Bastian 1984; Brooks et al. 1979; Kopelman et al. 1966; McGill and Motto 1974; Tillmann et al. 1997; Williams et al. 1974) and animals (Bailie et al. 1993, 1994; Fukushima et al. 1981; Kanz et al. 1992; Lamb et al. 1986; NTP 1983) regardless of the route of exposure. In humans that developed toxic hepatitis as a result of accidentally ingesting 4,4´-methyleneedianiline (Kopelman et al. 1966), there was no sign of progressive hepatic disease upon examination two years after the poisoning episode (Kopelman 1968). No deaths attributed to liver disease were reported among this same population over a period of 20 years after the accident (Ball et al. 1992). In animals, the severity of the effects was generally dose-related, and results from single dose studies showed that biliary epithelial cells are affected earlier than parenchymal liver cells (Bailie et al. 1994) and that the toxicant responsible for the damage is present in the bile (Kanz et al. 1995), although its chemical nature has not been established. It was also shown that leukocyte infiltration observed in the liver after an oral dose of 4,4´-methyleneedianiline does not contribute to liver damage. Acute-duration studies, mainly in rats, identified minimal effective doses around 25 mg/kg/day (Bailie et al. 1993). A NOAEL of 8.3 mg/kg/day was established in intermediate-duration study in rats (Pludro et al. 1969); LOAELs ranged between 67 and 100 mg/kg/day. Chronic LOAELs were 9 and 25 mg/kg/day in rats and mice, respectively (Lamb et al. 1986; NTP 1983). A LOAEL of 2.7 mg/kg/day was reported in dogs, but this study had severe design limitations (Deichmann et al. 1978). An acute oral MRL of 0.2 mg/kg/day was derived based on the results of a study by Bailie et al. (1993). In deriving this MRL, a modifying actor of 0.5 was used to account for the possibility that the corn oil vehicle might have facilitated the absorption of 4,4´-methyleneedianiline in the gastrointestinal tract. An intermediate MRL of 0.08 mg/kg/day was based on results from Pludro et al. (1969). The exact mechanism involved in liver toxicity is unknown, but some have suggested that it may be related to the formation of a reactive electrophile as a result of metabolic activation of 4,4´-methyleneedianiline (Lamb et al. 1986). The existing evidence indicates that humans exposed to elevated levels of 4,4´-methyleneedianiline, which may be found in the workplace, may be at risk of developing liver disease.

**Endocrine Effects.** No information was located regarding endocrine effects in humans exposed to 4,4´-methyleneedianiline by any route. Results from oral studies in animals suggest that the thyroid is a target for 4,4´-methyleneedianiline toxicity. Gross and histopathological alterations have been described
in rats and mice in acute- (Tullner 1960), intermediate- (Fukushima et al. 1981; Hagiwara et al. 1993; Hiasa et al. 1984; NTP 1983; Tsuda et al. 1987), and chronic-duration (Lamb et al. 1986; NTP 1983) oral studies. The mechanism of thyroid toxicity is not known, but some have speculated that it may be related to induced hypersecretion of thyroid stimulating hormone (TSH) which in turn induces thyroid hyperplasia (Hiasa et al. 1984). The relevance of the thyroid effects observed in animals to human health is unknown. Thyroid function has not been examined in populations with known exposure to 4,4´-methylenedianiline.

**Renal Effects.** Results from clinical tests in a subject who accidentally drank a solution containing 4,4´-methylenedianiline, potassium carbonate, and gamma-butyrolactone were consistent with renal tubular dysfunction (Roy et al. 1985), but the role of 4,4´-methylenedianiline, if any, is unknown. The only other relevant information in humans is a case in which proteinuria and erythrocyturia were observed in a young man who drank an alcoholic beverage spiked with 4,4´-methylenedianiline (Tillmann et al. 1997). No further data in humans were located. Intermediate-duration oral studies in animals established NOAELs of 83-141 mg/kg/day (Fukushima et al. 1979, 1981; NTP 1983; Pludro et al. 1969). However, in chronic-duration oral studies, high incidence of nephropathy was reported in mice at 19-25 mg 4,4´-methylenedianiline/kg/day and a high incidence of kidney mineralization was seen in male rats at 16 mg/kg/day (Lamb et al. 1986; NTP 1983). A much lower LOAEL of 2.7 mg/kg/day was reported in dogs, but this study suffered from severe limitations (Deichmann et al. 1978). The results from these long-term studies in animals suggest that the possibility exists for similar effects to occur in humans exposed to low 4,4´-methylenedianiline levels for a prolonged period of time, such as can occur near Superfund waste sites.

**Dermal Effects.** Erythema multiform was observed in a subject who accidentally ingested a liquid containing 4,4´-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). Although more than one chemical was involved in this case, the presence of erythema multiform is consistent with an immunological allergic reaction. A recent study reported that a subject who drank an alcoholic beverage spiked with 4,4´-methylenedianiline developed a skin rash (Tillmann et al. 1997). Skin rashes have commonly been described in individuals who had dermal contact with 4,4´-methylenedianiline (Brooks et al. 1979; Emmett 1976; McGill and Motto 1974; Van Joost et al. 1987). Also, a case of photosensitivity to 4,4´-methylenedianiline was described (Levine 1983). Exposure concentrations were not available. In some cases (Brooks et al. 1979; McGill and Motto 1974), skin rashes and pruritus could have been a manifestation of the underlying liver disease.
However, in other cases (Emmett 1976; Van Joost et al. 1987), proof of dermal sensitization was obtained when patch testing with 4,4’-methylenedianiline gave positive results. In the case described by Van Joost et al. (1987), the patient had no known previous exposure to 4,4’-methylenedianiline, but had been exposed to other chemicals. This led the investigators to suggest that cross sensitization may have occurred, specifically, to agents with substitution in the para position. The possibility of dermal sensitization across routes of exposure was examined in guinea pigs and the results were negative (Leong et al. 1987). The existing evidence indicates that 4,4’-methylenedianiline can cause dermal sensitization in humans.

**Ocular Effects.** Prolonged and severe loss of visual acuity was reported in a subject who accidentally ingested a liquid containing 4,4’-methylenedianiline, potassium carbonate, and gamma-butyrolactone (Roy et al. 1985). These effects could not, however, be conclusively attributed to 4,4’-methylenedianiline. No other reports were found regarding ocular effects in humans after exposure to 4,4’-methylenedianiline. Supporting the findings of Roy et al. (1985) is a report in which guinea pigs exposed nose-only to an aerosol of 4,4’-methylenedianiline for 2 weeks had serious morphological alterations in the retina (Leong et al. 1987). It is assumed that nose-only exposure prevented direct contact of the aerosol with the eye. On the other hand, no specific complaints of ocular effects were observed among a group of individuals who ate bread contaminated with 4,4’-methylenedianiline and developed toxic hepatitis (Kopelman 1968; Kopelman et al. 1966). Deposition of solid 4,4’-methylenedianiline into the eye of rabbits induced acute reversible effects in the cornea, iris, and conjunctiva (DuPont 1976b). The evidence available suggests that populations living near Superfund waste sites are not at a greater risk of developing adverse ocular effects due to exposure to 4,4’-methylenedianiline. However, direct contact with concentrated material, as may occur occupationally, can be hazardous.

**Body Weight Effects.** No studies were located regarding body weight effects in humans exposed to 4,4’-methylenedianiline by any route. Acute (NTP 1983), intermediate (Fukushima et al. 1979, 1981; Hagiwara et al. 1993; Hiasa et al. 1984; Miyamoto et al. 1977; NTP 1983; Tsuda et al. 1987), or chronic (Lamb et al. 1986; NTP 1983) oral exposure to 4,4’-methylenedianiline in animals usually resulted in a reduction in final body weight. Similar results were reported in rabbits exposed dermally for 10 days (DuPont 1976a). Food consumption data were not provided in these studies. The relevance of these findings to human health is unknown.
**Immunological and Lymphoreticular Effects.** Several cases of dermal sensitization have been described in subjects who became in contact with 4,4’-methylenedianiline in the workplace (Emmett 1976; Levine 1983; Van Joost et al. 1987), but no information exists regarding possible effects of 4,4’-methylenedianiline on human immunocompetence. Studies in animals have described histopathological alterations in the thymus of rats after single oral doses of 250 mg 4,4’-methylenedianiline/kg (Kanz et al. 1992). No histopathological alterations were seen in organs of the lymphoreticular system from rats or mice in intermediate or chronic oral studies (Lamb et al. 1986; NTP 1983). Immunocompetence has not been evaluated in animals. There is insufficient information to assess the potential immunological effects of prolonged exposure to low levels of 4,4’-methylenedianiline in humans.

**Neurological Effects.** No studies were located regarding neurological effects in humans after exposure to 4,4’-methylenedianiline by any route. Studies in animals provided no evidence of neurotoxicity after intermediate or chronic exposure (Lamb et al. 1986; NTP 1983), although a complete neurological examination including neurobehavioral testing has not been performed. Because 4,4’-methylenedianiline is a solid with low volatility and because of what is known about structurally similar compounds, it is not anticipated that exposures around Superfund sites will cause neurological damage in humans.

**Reproductive Effects.** No studies were located regarding reproductive effects in humans after exposure to 4,4’-methylenedianiline by any route. Hypertrophy of the uterus was observed in rats treated orally for 5-14 days with 110 mg 4,4’-methylenedianiline/kg/day (Tullner 1960). However, no such effects were seen after exposure to similar daily doses for 13 weeks (NTP 1983) or to lower doses for up to 103 weeks (Lamb et al. 1986; NTP 1983). Endometrial hyperplasia was reported in a dogs treated with doses of about 2.7 mg 4,4’-methylenedianiline/kg/day for over 54 months, but this study had severe design limitations (Deichmann et al. 1978). None of the animal studies examined reproductive function. There is not enough information to predict whether reproductive function in humans might be affected after exposure to 4,4’-methylenedianiline.

**Developmental Effects.** No studies were located regarding developmental effects in humans after exposure to 4,4’-methylenedianiline by any route. A single oral study in rats administered 4,4’-methylenedianiline during gestation showed that doses $\geq$37 mg/kg/day may cause developmental abnormalities in the offspring (Bourdelat et al. 1983). It should be noted, however, that these dose
levels also induced maternal toxicity. Furthermore, only one rat was used as control and the results were poorly reported. Based on the evidence available, no predictions can be made regarding developmental effects in populations exposed to 4,4'-methylenedianiline.

**Genotoxic Effects.** As seen in Table 2-4, limited information exists regarding genotoxic effects of 4,4'-methylenedianiline *in vivo*. In rats, a single intraperitoneal injection of approximately 73 mg 4,4'-methylenedianiline/kg significantly increased the incidence of DNA fragmentation in liver cells (Parodi et al. 1981). In a study in which only two rats were used, intraperitoneal administration of approximately 80 mg 4,4'-methylenedianiline/kg to one rat increased the amount of DNA adducts in the liver relative to the control rat treated with the vehicle only (Endo and Hara 1991). In more recent studies, it was shown that 4,4'-methylenedianiline has a relatively low DNA binding potency (Schitze et al. 1996). In that study, the total radioactivity bound to liver DNA from rats treated with a single intraperitoneal dose of 0.2 or 23 mg of 4,4'-methylenedianiline/kg corresponded to 0.06 and 2.7 adducts per 10.' nucleotides, and this put 4,4'-methylenedianiline in the range of weakly genotoxic compounds. The structure of the adducts was not determined, but it was determined that adduct formation was covalent in nature. Similar results were reported in rats after acute oral administration of 4,4'-methylenedianiline (Vock et al. 1996). In the latter case, no adducts could be detected in the bladder and peripheral lymphocytes. Contradictory results were obtained regarding sex-linked recessive lethal mutations in *Drosophila melanogaster*. Fourman et al. (1994) reported a significant increase in the incidence of recessive lethal mutations while Ho et al. (1979) reported no significant increase.

Many studies have examined the genotoxic properties of 4,4'-methylenedianiline *in vitro* in *Salmonella typhimurium* (Andersen et al. 1980; Cocker et al. 1986a; Ho et al. 1979; LaVoie et al. 1979; McCarthy et al. 1982; Messerly et al. 1987; Parodi et al. 1981; Rannug et al. 1984; Rao et al. 1982; Takemura and Shimizu 1978; Tanaka et al. 1985; Tsuchiya 1995). With few exceptions (see Table 2-5), the results showed that 4,4'-methylenedianiline is genotoxic after metabolic activation with S-9 systems. Although not conclusively established, the genotoxic properties have been attributed to a reactive electrophile formed as a result of N-hydroxylation. On the other hand, N-acetylated metabolites have been shown to be nonmutagenic (Cocker et al. 1986b; Tanaka et al. 1985). 4,4'-Methylenedianiline also was shown to be mutagenic in the eukaryotic organism *Saccharomyces cerevisiae* with and without metabolic activation (Ho et al. 1979). 4,4'-Methylenedianiline did not increase the incidence of sister chromatid exchanges in human leukocytes *in vitro* when tested with or without activation (Ho...
Table 2-4. Genotoxicity of 4,4'-Methylenedianiline *In Vivo*

<table>
<thead>
<tr>
<th>Species (test system)</th>
<th>End point</th>
<th>Results</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat liver cells</td>
<td>DNA fragmentation</td>
<td>+</td>
<td>Parodi et al. 1981</td>
</tr>
<tr>
<td>Mouse bone marrow cells</td>
<td>Sister chromatid exchange</td>
<td>+</td>
<td>Parodi et al. 1981</td>
</tr>
<tr>
<td><em>Drosophila melanogaster</em></td>
<td>Sex linked recessive lethal mutations</td>
<td>+</td>
<td>Foureman et al. 1994</td>
</tr>
<tr>
<td><em>D. melanogaster</em></td>
<td>Sex linked recessive lethal mutations</td>
<td>–</td>
<td>Ho et al. 1979</td>
</tr>
</tbody>
</table>

+ = Positive result; – = negative result; DNA = deoxyribonucleic acid
Table 2-5. Genotoxicity of 4,4'-Methylenedianiline *In Vitro*

<table>
<thead>
<tr>
<th>Species (test system)</th>
<th>End point</th>
<th>Result With activation</th>
<th>Result Without activation</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Prokaryotic organisms:</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Salmonella typhimurium</em></td>
<td>Reverse mutation</td>
<td>+</td>
<td>ND</td>
<td>Andersen et al. 1980</td>
</tr>
<tr>
<td>TA 100 on plates</td>
<td></td>
<td>+</td>
<td>ND</td>
<td>Parodi et al. 1981</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>ND</td>
<td>McCarthy et al. 1982</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>ND</td>
<td>Cocker et al. 1986b</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Takemora and Shimizu 1978</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>LaVoi et al. 1979</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Tanaka et al. 1985</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Rao et al. 1982</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Messerly et al. 1987</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Tsuchiya 1995</td>
</tr>
<tr>
<td><em>S. typhimurium</em></td>
<td>Reverse mutation</td>
<td>-</td>
<td>ND</td>
<td>Parodi et al. 1981</td>
</tr>
<tr>
<td>TA 98 on plates</td>
<td></td>
<td></td>
<td>-</td>
<td>Takemora and Shimizu 1978</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Tanaka et al. 1985</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Rao et al. 1982</td>
</tr>
<tr>
<td></td>
<td></td>
<td>+</td>
<td>-</td>
<td>Rannug et al. 1984</td>
</tr>
<tr>
<td></td>
<td>(+)</td>
<td></td>
<td>-</td>
<td>Messerly et al. 1987</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>-</td>
<td>Tsuchiya 1995</td>
</tr>
<tr>
<td><em>S. typhimurium</em></td>
<td>Reverse mutation</td>
<td>+</td>
<td>-</td>
<td>Rannug et al. 1984</td>
</tr>
<tr>
<td>TA 1538, TA 1535 on plates</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eukaryotic organisms:</td>
<td>DNA repair</td>
<td>+</td>
<td>+</td>
<td>Ho et al. 1979</td>
</tr>
<tr>
<td><em>Saccharomyces cerevisiae</em></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mammalian cells</td>
<td>Unscheduled DNA synthesis</td>
<td>ND</td>
<td>+</td>
<td>Mori et al. 1988</td>
</tr>
<tr>
<td>Rat hepatocytes</td>
<td>Unscheduled DNA synthesis</td>
<td>-</td>
<td>+</td>
<td>Mirsalis et al. 1983</td>
</tr>
<tr>
<td>Rat hepatocytes</td>
<td>Unscheduled DNA synthesis</td>
<td>-</td>
<td>+</td>
<td>McGregor et al. 1988</td>
</tr>
<tr>
<td>L5178Y mouse lymphoma cells</td>
<td>Forward mutation</td>
<td>ND</td>
<td>+</td>
<td>Ho et al. 1979</td>
</tr>
<tr>
<td>Human leukocytes</td>
<td>Sister chromatid exchange</td>
<td>-</td>
<td>-</td>
<td>Ho et al. 1979</td>
</tr>
</tbody>
</table>

- = negative results; + = positive results; (+) = weakly positive results; DNA = deoxyribonucleic acid; ND = no data
et al. 1979), but increased unscheduled DNA synthesis in rat hepatocytes (Mirsalis et al. 1983; Mori et al. 1988) and the incidence of forward mutations in L5178Y mouse lymphoma cells (McGregor et al. 1988). The overall evidence is insufficient to indicate that exposure to 4,4´-methylenedianiline may cause genetic damage in humans.

Cancer. Limited information exists regarding potential carcinogenicity of 4,4´-methylenedianiline in humans. No association was found between deaths from cancers and 4,4´-methylenedianiline in a group of 84 subjects who ate bread contaminated with 4,4´-methylenedianiline (Hall et al. 1992). The evaluation was conducted over a period of 20 years following the accident. However, the cohort is quite small for drawing any firm conclusions. Similar lack of association was reported for a cohort of 550 male and 45 female Swedish workers exposed to 4,4´-methylenedianiline (predominantly by dermal contact) (Selden et al. 1992). The exposure period was not clearly established. It was noted, however, that the subjects were quite young and had not reached cancer-prone age, and that the follow-up period was short and may not have covered the latency period for some cancers, in particular bladder cancer, which has a latency of about 20 years. Two additional studies also provided inconclusive evidence of increased incidence of bladder cancer among workers exposed to 4,4´-methylenedianiline; studies were inconclusive mainly because of simultaneous exposure to other chemicals (Cragle et al. 1992; Liss and Guirguis 1994).

Results from a 2-year drinking water bioassay provided clear evidence of carcinogenicity in rats and mice (Lamb et al. 1986; NTP 1983). Under the conditions of the study, 4,4´-methylenedianiline caused significantly increased incidences of thyroid follicular cell carcinomas in male rats, thyroid follicular cell adenomas in female rats and both male and female mice, C-cell adenomas of the thyroid gland in female rats, neoplastic nodules in the liver of male rats, hepatocellular carcinomas in male and female mice, adenomas of the liver and malignant lymphomas in female mice, and adrenal pheochromocytomas in male mice. It should be mentioned, however, that considerable debate exists on whether adenomas result in, or are a precursor to, malignant neoplasms.

4,4´-Methylenedianiline was not a promoter of liver, kidney, or urinary bladder tumors in rats (Fukushima et al. 1981; Masui et al. 1986); in fact, it inhibited the neoplastic response induced by the carcinogens alone. It was postulated that the mechanism may be related to a decreased food consumption and reduced growth (Fukushima et al. 1981; Masui et al. 1986). In contrast,
4,4′-methyleneedianiline promoted the formation of thyroid tumors in rats initiated with N-bis(2-hydroxypropyl)nitrosamine (Hiasa et al. 1984).

The mechanism of 4,4′-methyleneedianiline carcinogenicity is not known. It has been suggested that a reactive intermediate resulting from metabolic activation may be responsible for the liver carcinogenicity by binding to cell macromolecules, and that a nongenetic mechanism may be involved in thyroid carcinogenicity (Lamb et al. 1986). Based on recent data regarding the low potential of 4,4′-methyleneedianiline to bind to rat liver DNA (Schiitze et al. 1996), a nongenotoxic mechanism of liver carcinogenicity would also appear likely. Based on inadequate evidence of carcinogenicity in humans and sufficient evidence in experimental animals, the International Agency for Research on Cancer (IARC 1987) has designated 4,4′-methyleneedianiline a Group 2B carcinogen and possibly carcinogen in humans.

2.6 BIOMARKERS OF EXPOSURE AND EFFECT

Biomarkers are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility (NAS/NRC 1989).

Due to a nascent understanding of the use and interpretation of biomarkers, implementation of biomarkers as tools of exposure in the general population is very limited. A biomarker of exposure is a xenobiotic substance or its metabolite(s), or the product of an interaction between a xenobiotic agent and some target molecule(s) or cell(s) that is measured within a compartment of an organism (NRC 1989). The preferred biomarkers of exposure are generally the substance itself or substance-specific metabolites in readily obtainable body fluid(s) or excreta. However, several factors can confound the use and interpretation of biomarkers of exposure. The body burden of a substance may be the result of exposures from more than one source. The substance being measured may be a metabolite of another xenobiotic substance (e.g., high urinary levels of phenol can result from exposure to several different aromatic compounds). Depending on the properties of the substance (e.g., biologic half-life) and environmental conditions (e.g., duration and route of exposure), the substance and all of its metabolites may have left the body by the time samples can be taken. It may be difficult to identify individuals exposed to hazardous substances that are commonly found in body tissues and fluids (e.g.,
essential mineral nutrients such as copper, zinc, and selenium). Biomarkers of exposure to 4,4′-methyleneedianilne are discussed in Section 2.6.1.

Biomarkers of effect are defined as any measurable biochemical, physiologic, or other alteration within an organism that, depending on magnitude, can be recognized as an established or potential health impairment or disease (NAS/NRC 1989). This definition encompasses biochemical or cellular signals of tissue dysfunction (e.g., increased liver enzyme activity or pathologic changes in female genital epithelial cells), as well as physiologic signs of dysfunction such as increased blood pressure or decreased lung capacity. Note that these markers are not often substance specific. They also may not be directly adverse, but can indicate potential health impairment (e.g., DNA adducts). Biomarkers of effects caused by 4,4′-methyleneedianilne are discussed in Section 2.6.2.

A biomarker of susceptibility is an indicator of an inherent or acquired limitation of an organism’s ability to respond to the challenge of exposure to a specific xenobiotic substance. It can be an intrinsic genetic or other characteristic or a preexisting disease that results in an increase in absorbed dose, a decrease in the biologically effective dose, or a target tissue response. If biomarkers of susceptibility exist, they are discussed in Section 2.8, Populations That Are Unusually Susceptible.

### 2.6.1 Biomarkers Used to Identify or Quantify Exposure to 4,4′-Methyleneedianilne

Exposure to 4,4′-methyleneedianilne in manufacturing and use has been assessed by monitoring 4,4′-methyleneedianilne in the urine (Cocker et al. 1986a, 1994; Dalene et al. 1995; Schtitze et al. 1995). A metabolite of 4,4′-methyleneedianilne, N-acetylmethyleneedianilne, also has been quantified in urine (Cocker et al. 1986a; Schtitze et al. 1995). Quantification of these substances is performed by gas chromatography-mass spectrophotometry. Cocker et al. (1994) was able to show that when exposure to 4,4′-methyleneedianilne occurred through inhalation, postshift urine samples contained higher concentrations of 4,4′-methyleneedianilne than samples taken preshift next day. When exposure was thought to be through the dermal route, urine collected preshift next day had higher-concentration of 4,4′-methyleneedianilne than samples taken immediately postshift on the day of exposure. These results suggested that pulmonary absorption is relatively fast such that peak excretion is reached at the end of the shift. In contrast, absorption through the skin was slower and maximum excretion occurred the next morning. N,N′-diacetylmethyleneedianilne has also been found in the urine of exposed workers, but at a much lower concentration than the monoacetylated metabolite (Robert et al. 1995).
Little is known about the pharmacokinetics of 4,4´-methyleneedianiline in humans, but data in animals suggest that it is eliminated fairly rapidly and does not accumulate. This is consistent with what has been found in occupationally exposed humans. In a study involving 411 exposed workers, 42% of the urine samples had no detectable 4,4´-methyleneedianiline (Cocker et al. 1994). In another study of 111 exposed workers, the concentration of 4,4´-methyleneedianiline in 81% of over 300 urine samples was below the detection limit for the method (Cocker et al. 1986a). This suggests that monitoring urinary 4,4´-methyleneedianiline may not be sensitive enough in cases of brief or single exposures to low levels of the chemical. Furthermore, only an indication of recent exposure would be obtained. In contrast with the results from Cocker et al. (1986a), a recent study by Schütze et al. (1995) found 4,4´-methyleneedianiline in the urine from workers exposed to low levels of the chemical (<20 pg/m³). Thirty-three workers were examined and 4,4´-methyleneedianiline was detected in the urine (0.013-2.76 nmol/L) from all of them. Urinary levels of the metabolite N-acetylmethyleneedianiline ranged from 0.045 to 23.4 nmol/L.

Recent studies have examined the formation of adducts of 4,4´-methyleneedianiline with hemoglobin as a means for monitoring exposure (Bailey et al. 1990; Farmer and Bailey 1989; Schütze et al. 1995). In the recent study by Schütze et al. (1995), the authors showed that there was a good correlation (r=0.77) between adduct levels in hemoglobin and 4,4´-methyleneedianiline released after acid treatment of urine. The following mechanism of adduct formation has been postulated (Farmer and Bailey 1989). N-hydroxymethyleneedianiline resulting from the enzymatic oxidation of 4,4´-methyleneedianiline is further oxidized to nitroso compounds, which may react with sulfhydryl groups in hemoglobin. This yields N-hydroxy sulphenamides, which then rearrange to more stable sulphinamides. Hydrolysis of the sulphinamides under mild acidic or basic conditions releases the parent amine, which may be extracted and quantitated. This approach has a clear advantage over directly monitoring 4,4´-methyleneedianiline because the adduct is stable in vivo and its elimination is related to the lifespan of the erythrocytes, which in humans is 120 days (Farmer and Bailey 1989). In addition, there is an advantage over DNA adducts which are subjected to DNA repair reactions. Although no data exist yet in humans, studies in rats showed that 4,4´-methyleneedianiline and N-acetylmethyleneedianiline formed adducts with hemoglobin and that a dose-response relationship could be established over the range of oral doses used (1-12 mg/kg) (Bailey et al. 1990).
2.6.2 Biomarkers Used to Characterize Effects Caused by 4,4´-Methylenedianiline

There are no specific biomarkers of effects for 4,4´-methylenedianiline. Studies summarized in Section 2.2 indicate that exposure to 4,4´-methylenedianiline causes adverse liver effects in humans (Kopelman et al. 1966; McGill and Motto 1974; Williams et al. 1974). However, exposure to many other chemicals, not even necessarily of a structure similar to 4,4´-methylenedianiline, can produce a similar clinical picture. Also, the properties of dermal sensitizer are not unique to 4,4´-methylenedianiline, and a positive patch testing response to 4,4´-methylenedianiline may also be obtained in cases of previous contact with chemicals substituted in the para-position (cross sensitization) (Van Joost et al. 1987).

For more information on biomarkers for renal and hepatic effects of chemicals see the ATSDR/CDC Subcommittee Report on Biological Indicators of Organ Damage (1990); for information on biomarkers for neurological effects, see OTA (1990).

2.7 INTERACTIONS WITH OTHER CHEMICALS

No information was located regarding the influence of other chemicals on the toxicity of 4,4´-methylenedianiline in humans. Limited data in animals showed that pretreatment of rats with the monooxygenase function inhibitor aminobenzotriazol, ameliorated the hepatic effects of 4,4´-methylenedianiline (Bailie et al. 1993). However, pretreatment with the inhibitor SKF-525A had no effect. Based on these results, the investigators proposed that 4,4´-methylenedianiline requires metabolic activation for its toxicity and that activation is carried out by an isozyme of cytochrome P-450 that is inhibited by aminobenzotriazol, but not SKF-525A (Bailie et al. 1993).

4,4´-Methylenedianiline inhibited the formation of liver, kidney, and urinary bladder tumors when administered orally to rats following initiation with various carcinogens (Fukushima et al. 1981; Masui et al. 1986). The mechanism for this interaction is not known. Some have suggested that a reduction in food consumption and, consequently, in growth, induced by 4,4´-methylenedianiline, may play a role (Fukushima et al. 1981). A different possibility that has been offered is that 4,4´-methylenedianiline may alter the activities of enzymes involved in carcinogen metabolism, resulting in reduced biotransformation of the carcinogens (Tsuda et al. 1987). 4,4´-Methylenedianiline has been shown to alter the activities of several biotransformation enzymes in rats (Wu et al. 1989).
2.8 POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE

A susceptible population will exhibit a different or enhanced response to 4,4′-methylenedianiline than will most persons exposed to the same level of 4,4′-methylenedianiline in the environment. Reasons may include genetic makeup, age, health and nutritional status, and exposure to other toxic substances (e.g., cigarette smoke). These parameters may result in reduced detoxification or excretion of 4,4′-methylenedianiline, or compromised function of target organs affected by 4,4′-methylenedianiline. Populations who are at greater risk due to their unusually high exposure to 4,4′-methylenedianiline are discussed in Section 5.6, Populations With Potentially High Exposure.

In the review of the literature regarding toxic effects of 4,4′-methylenedianiline, no information was located on any population that might be unusually sensitive to 4,4′-methylenedianiline. However, based on the knowledge of the primary toxic effect of 4,4′-methylenedianiline (liver toxicity), a number of groups may be proposed as being potentially highly sensitive to this chemical. Some of these groups include very young children who have an immature hepatic detoxification system and individuals with impaired liver function (i.e., liver cirrhosis). In addition, people who develop dermatitis following exposure to 4,4′-methylenedianiline in the workplace may become hypersensitive to subsequent exposure to the chemical or to structurally related chemicals (Van Joost et al. 1987). Also, since the enzyme N-acetyltransferase exhibits polymorphism, slow acetylators will be prone to more toxic insult of 4,4′-methylenedianiline than fast acetylators, assuming that acetylation represents a true detoxification pathway.

2.9 METHODS FOR REDUCING TOXIC EFFECTS

This section will describe clinical practice and research concerning methods for reducing toxic effects of exposure to 4,4′-methylenedianiline. However, because some of the treatments discussed may be experimental and unproven, this section should not be used as a guide for treatment of exposures to 4,4′-methylenedianiline. When specific exposures have occurred, poison control centers and medical toxicologists should be consulted for medical advice.

No texts were found that provided specific information about treatment following exposures to 4,4′-methylenedianiline.
2.9.1 Reducing Peak Absorption Following Exposure

4,4'-Methylenedianiline can be absorbed through the skin, by inhalation of fine dust or vapor, or by ingestion. The only recommendation found in the literature to reduce peak absorption after ingestion was induction of vomiting (Allied Chemical 1978). Removing the patient to fresh air is suggested if inhalation occurs, and washing with soap and water is recommended after contact with the skin (Allied Chemical 1978; NIOSH 1997). NIOSH (1997) further recommends immediately washing the eyes with large amounts of water, occasionally lifting the lower and upper lids. A recent study examined the efficacy of either 100% ethanol, 100% water, or 1% or 10% soap solution in removing 4,4'-methylenedianiline from human and rat skin \textit{in vitro} (Hewitt et al. 1995). The results showed that all solutions were equally effective in removing the chemical from the human skin surface, with 21-47% of the applied dose removed at 72 hours. In contrast, 100% ethanol or 10% soap solution were significantly more effective in removing 0 4,4'-methylenedianiline from the rat skin than 100% water or 1% soap solution. The results also showed that washing the skin surface 3 or 30 minutes after dosing significantly reduced absorption (2- to 3-fold) compared with control unwashed skin. Washing after 72 hours following application of the compound did not significantly reduce absorption. Data from an \textit{in vivo} study in a group of rats and guinea pigs and in a monkey showed that washing the application site with soap and water was more effective in removing 4,4'-methylenedianiline from the skin than washing with acetone and water (El-Hawaii et al. 1986).

2.9.2 Reducing Body Burden

No information was located regarding reducing the body burden after exposure to 4,4'-methylenedianiline.

2.9.3 Interfering with the Mechanism of Action for Toxic Effects

The mechanism of 4,4'-methylenedianiline toxicity is not known. Information on structurally related chemicals suggests that the toxicity of 4,4'-methylenedianiline is due to the formation of a reactive intermediate that can bind covalently to cell macromolecule. The reactive intermediate is produced by the enzymatic N-oxidation of 4,4'-methylenedianiline (Lamb et al. 1986). A different metabolic pathway, which has been demonstrated in humans, is N-acetylation (Cocker et al. 1986a, 1994). This pathway seems to represent a detoxification route since N-acetylmethylenedianiline and
N,N’-diacetylmethylenedianiline, in the presence of activating systems, were not genotoxic in mutagenicity assays (Cocker et al. 1986b; Tanaka et al. 1985). This was in contrast with a strong mutagenic effect of the parent compound with activation. Therefore, it would appear that if the acetylation pathway could be favored over the N-oxidation route, some toxic effects attributed to 4,4’-methylenedianiline might be diminished.

2.10 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of 4,4’-methylenedianiline is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 4,4’-methylenedianiline. The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

2.10.1 Existing information on Health Effects of 4,4’-Methylenedianiline

The existing data on health effects of inhalation, oral, and dermal exposure of humans and animals to 4,4’-methylenedianiline are summarized in Figure 2-5. The purpose of this figure is to illustrate the existing information concerning the health effects of 4,4’-methylenedianiline. Each dot in the figure indicates that one or more studies provide information associated with that particular effect. The dot does not necessarily imply anything about the quality of the study or studies, nor should missing information in this figure be interpreted as a “data need.” A data need, as defined in ATSDR’s Decision Guide for Identifying Substance-Specific Data Needs Related to Toxicological Profiles (ATSDR 1989), is substance-specific information necessary to conduct comprehensive public health assessments. Generally, ATSDR defines a data gap more broadly as any substance-specific information missing from the scientific literature.
2. HEALTH EFFECTS

Figure 2-5. Existing Information on Health Effects of 4,4'-Methylenedianiline

### Human

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### Animal

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● Existing Studies
Data regarding health effects of 4,4′-methylenedianiline in humans result from a few studies involving accidental acute oral exposure to 4,4′-methylenedianiline, and from cases of dermal exposure in the workplace. No information was located regarding inhalation exposure in humans. No information is available regarding neurological, reproductive, or developmental effects in humans by any route of exposure.

Most of the data regarding the health effects of 4,4′-methylenedianiline in animals were obtained from studies in which 4,4′-methylenedianiline was administered orally. A single study was located concerning health effects in animals following inhalation exposure, and few reports on dermal exposure to 4,4′-methylenedianiline were located. The available information from animal studies is insufficient to determine with certainty whether the effects of 4,4′-methylenedianiline are route- or species-specific, or age-dependent. Information on the dermal route of exposure would be useful because of the potential exposure via this route for workers in industry. Because of its low volatility, inhalation of 4,4′-methylenedianiline appears less likely than dermal contact, but cannot be completely ruled out. Although there are limited environmental data regarding 4,4′-methylenedianiline, the dermal route of exposure appears to be the most relevant for humans living near hazardous waste sites where 4,4′-methylenedianiline might be found.

2.10.2 Identification of Data Needs

**Acute-Duration Exposure** Populations living in the vicinity of hazardous waste sites may be exposed to 4,4′-methylenedianiline for a short time. Exposure would probably occur via the dermal route, but oral exposure, particularly by children ingesting contaminated soil, cannot be excluded.

The liver is the major target of 4,4′-methylenedianiline toxicity in humans following acute exposure by any route (Brooks et al. 1979; Kopelman et al. 1966; McGill and Motto 1974; Roy et al. 1985; Tillmann et al. 1997). The same was seen in rats (Bailie et al. 1993, 1994; Kanz et al. 1992; Schmidt et al. 1980), but other animal species have not been studied. An acute oral MRL was derived based on liver effects in rats identified in the Bailie et al. (1993) study. The mechanism of liver toxicity has not been elucidated, but it seems that the biliary tree epithelium is affected before the liver parenchyma (Bailie et al. 1994). Further studies are necessary to corroborate this finding and possibly identify the chemical entity responsible for the liver toxicity. In addition, studies providing quantitative estimates of excretion of 4,4′-methylenedianiline and/or metabolites in the bile would help
establish a possible relationship between this excretory route and bile duct toxicity. The existing information also showed that 4,4'-methylenedianiline can cause adverse dermal reactions in humans (Emmett 1976; Van Joost et al. 1987; Williams et al. 1974). Intermediate- and chronic-duration studies in animals suggest that the thyroid is also a target for 4,4'-methylenedianiline toxicity, but this end point has not been examined in acute-duration studies. Reproductive function has not been examined in acute-duration studies by any route; such studies could provide valuable information on possible effects of 4,4'-methylenedianiline on that system. Based on its chemical structure, 4,4'-methylenedianiline does not appear to be a neurotoxicant, but the nervous system has not been examined in acute-duration studies. A limited number of acute dermal studies in animals have suggested that the administration vehicle plays a role in the toxicity of 4,4'-methylenedianiline, possibly by influencing absorption (DuPont 1975, 1976a). Further dermal studies with different vehicles and also oral studies with 4,4'-methylenedianiline in different soils would provide valuable information for those living near hazardous waste sites where 4,4'-methylenedianiline may be found.

**Intermediate-Duration Exposure.** No studies were located on intermediate-duration exposure to 4,4'-methylenedianiline in humans by any route. The preponderance of data is available from rats exposed to 4,4'-methylenedianiline in the diet (Fukushima et al. 1979, 1981; Hagiwara et al. 1993; Hiasa et al. 1984; Miyamoto et al. 1977), by gavage (Pludro et al. 1969), or through the drinking water (NTP 1983). Results from these studies indicate that the liver and the thyroid are targets for 4,4'-methylenedianiline toxicity. An intermediate oral MRL was derived based on liver effects in rats reported by Pludro et al. (1969). Studies in other species would provide information on whether these end points are sensitive targets across species. The mechanism of thyroid toxicity is not known; therefore, research aimed to investigate this subject would be helpful. Neurological, reproductive, and developmental effects have not been appropriately studied in oral intermediate-duration studies, and no information is available regarding the inhalation and dermal routes of exposure. Pharmacokinetic data to help identify potential target organs after exposure by any route were not located.

**Chronic-Duration Exposure and Cancer.** No studies were located following chronic-duration inhalation or oral exposure to 4,4'-methylenedianiline in humans. Data were available concerning dermal exposure to 4,4'-methylenedianiline in the workplace. Results from one study confirmed that the liver is a target for 4,4'-methylenedianiline toxicity (Bastian 1984). A skin reaction triggered by exposure to sunlight was described in another study (Levine 1983). Research regarding the mechanism involved in photosensitivity to 4,4'-methylenedianiline would be helpful. Examination of
subjects occupationally exposed (presumably long-term exposure) revealed that 4,4´-methyleneedianiline can be absorbed through the lungs and the skin and that the parent compound and metabolites can be excreted in the urine (Cocker et al. 1986a, 1994; Schiitze et al. 1995). A limited number of oral studies in animals showed that the liver, thyroid, and perhaps the kidney are targets for 4,4´-methyleneedianiline toxicity (Deichmann et al. 1978; Lamb et al. 1986; NTP 1983). One of these studies, Deichmann et al. (1978), was conducted in dogs and the results suggested that dogs may be a particularly sensitive species for liver and kidney effects. However, this study was not well designed (only 9 dogs were tested and no concurrent controls were used), and therefore, the results are inconclusive. The inadequacies of this study precluded derivation of a chronic oral MRL. A well conducted chronic oral study in dogs could remove the uncertainty regarding dogs as very sensitive species for 4,4´-methyleneedianiline toxicity. A comprehensive clinical evaluation of individuals exposed to low levels of 4,4´-methyleneedianiline for many years in the workplace may provide evidence on less recognizable subtle effects of 4,4´-methyleneedianiline. Neither immunocompetence nor reproductive function has been examined in animals after chronic exposure to 4,4´-methyleneedianiline by any route of exposure. Such studies after low-level chronic exposure by the oral and dermal routes would be of value to determine whether exposures via these routes could cause toxicity in populations living near hazardous waste sites, or in those exposed in industries where this chemical is used.

Two-year cancer bioassays have been performed following oral and dermal exposure. In a well designed 2-year drinking water bioassay, there was clear evidence of carcinogenicity in rats and mice (Lamb et al. 1986; NTP 1983). An oral bioassay in dogs showed no evidence of carcinogenicity, but, as previously mentioned, the study was poorly designed and therefore, the results are inconclusive (Deichmann et al. 1978). A dermal bioassay in mice found 4,4´-methyleneedianiline to be a liver carcinogen, but it appears that the strain of mice used may have been particularly susceptible to liver tumors; therefore, the results are also inconclusive (Holland et al. 1987). A follow-up study on a group of individuals who had accidentally ingested 4,4´-methyleneedianiline and contracted toxic hepatitis did not find increased incidences of cancer (Hall et al. 1992). A similar conclusion was reached in a small group of power generation workers exposed to an epoxy resin containing 4,4´-methyleneedianiline (Selden et al. 1992). Two additional occupational studies found inconclusive evidence of bladder cancer in workers who were also exposed to other chemicals (Cragle et al. 1992; Liss and Guirguis 1994). Follow-up and other epidemiological studies are needed to adequately assess the carcinogenicity of 4,4´-methyleneedianiline in humans.
Genotoxicity. The only information in humans was provided by Ho et al. (1979), who reported that incubation of human leukocytes with 4,4´-methylenedianiline did not increase the incidence of sister chromatid exchanges. Existing genotoxicity studies indicate that 4,4´-methylenedianiline was mutagenic in *Salmonella* (Andersen et al. 1980; Cocker et al. 1986b; LaVoie et al. 1979; McCarthy et al. 1982; Parodi et al. 1981; Rao et al. 1982; Tanaka et al. 1985) with metabolic activation. 4,4´-Methylenedianiline also induced DNA damage in rat hepatocytes (Mirsalis et al. 1983; Mori et al. 1988) *in vitro* and formed adducts with rat liver DNA (Endo and Hara 1991; Schiitze et al. 1996; Vock et al. 1996). Cytogenic analysis of human populations exposed to 4,4´-methylenedianiline in occupational settings would provide an opportunity to assess the genotoxic potential of this chemical in humans.

Reproductive Toxicity. No studies were located regarding reproductive effects in humans after exposure to 4,4´-methylenedianiline by any route. A small number of studies provided information on the gross and histopathological appearance of reproductive organs in animals after acute (Tullner 1960), intermediate (NTP 1983), and chronic (Deichmann et al. 1978; NTP 1983) oral exposure to 4,4´-methylenedianiline. Hypertrophy of the uterus in rats was described in an acute study (Tullner 1960), and endometrial hyperplasia was reported in dogs after chronic treatment with relatively low 4,4´-methylenedianiline doses (Deichmann et al. 1978). However, the latter study has severe limitations (small number of animals and no concurrent controls) and the results should be interpreted with caution. A single acute dermal study found no alterations in the testes and epididymis from rabbits after treatment with 4,4´-methylenedianiline (DuPont 1975). Since none of these studies examined reproductive function, additional studies are necessary to adequately evaluate this parameter. A multi-generational study would provide valuable information. Examination of clinical records from women exposed for prolonged periods of time to low levels of 4,4´-methylenedianiline at work may provide information on reproductive outcome for this population. Oral and dermal studies in animals would provide information on susceptibility by these routes. The dermal route is of particular concern in individuals exposed to 4,4´-methylenedianiline in the workplace.

Developmental Toxicity. No studies were located regarding developmental effects in humans after exposure to 4,4´-methylenedianiline by any route. A single study in rats showed that exposure to 4,4´-methylenedianiline during gestation may result in adverse developmental effects (Bourdelat et al. 1983). The study, however, was of limited scope and was not well conducted. Further well-designed studies in animals exposed during pregnancy would provide information on the potential effects of
4,4’-methyleneedianiline on the developing organism. Studies by the dermal route are of particular interest since this route is of concern in occupational settings and to people living near hazardous waste sites where 4,4’-methyleneedianiline might be found.

**Immunotoxicity.** Information in humans indicates that dermal contact with 4,4’-methyleneedianiline may trigger allergic dermal reactions in some individuals (Emmett 1976; McGill and Motto 1974; Van Joost et al. 1987). In one case, a reaction was triggered by exposure to sunlight (Levine 1983). Animal studies have provided limited information. An acute oral study found histopathological alterations in the thymus from rats (Kanz et al. 1992). Intermediate oral studies found no histopathological alterations in organs of the lymphoreticular system from rats and mice (NTP 1983). Similar findings were reported in chronic oral studies in rats and mice (NTP 1983). The NTP (1983) study did not evaluate blood components. A chronic oral study in dogs reported spleen effects at a relatively low dose (Deichmann et al. 1978); however, because of study limitations, the results must be interpreted with caution. Two acute dermal studies provided information on spleen weight in mice (Holland et al. 1987) and on histopathology of spleen and thymus in rabbits (DuPont 1975). None of the studies in animals examined immunocompetence. Studies that examine antibody levels and responses to bacterial and viral infections after exposure to 4,4’-methyleneedianiline would provide valuable information on the immune system. Some immune parameters (e.g., serum immunoglobulin levels, response to mitogen stimulation) have been found to be quite sensitive to chemical insult. Also, evaluation of morbidity among individuals exposed to 4,4’-methyleneedianiline in the workplace may provide important indirect evidence regarding their immune status.

**Neurotoxicity.** No studies were located regarding neurotoxicity in humans after exposure to 4,4’-methyleneedianiline by any route. An acute dermal study in rabbits (DuPont 1975) and intermediate and chronic oral studies in rats and mice (Lamb et al. 1986; NTP 1983) found no gross or histopathological lesions in tissues from the peripheral and central nervous system. However, more sensitive neurological end points have not been examined. Laboratory animal studies that focus on subtle neurological effects following acute, intermediate, or chronic exposure via oral and dermal routes would help estimate potential neurotoxic effects in humans living near hazardous waste sites and in workers who might be exposed in certain occupational settings. Furthermore, evaluation of neurological end points in offspring from animals exposed during gestation would provide information that may be relevant to children of pregnant women exposed to 4,4’-methyleneedianiline in the workplace.
**Epidemiological and Human Dosimetry Studies.** A retrospective study investigated the relationship between deaths caused by neoplastic and non-neoplastic diseases and accidental ingestion of 4,4′-methylenedianiline in a cohort originally composed of 84 subjects (Hall et al. 1992). The study period covered approximately 20 years after the accident occurred. The results showed no obvious link between ingestion of 4,4′-methylenedianiline and any particular cancer or non-neoplastic disease. Another study conducted a retrospective assessment of exposure to 4,4′-methylenedianiline and cancer morbidity in power generation workers in Sweden (Selden et al. 1992). An overall standardized cancer incidence ratio of 0.52 indicated no increased incidence of cancer due to exposure to 4,4′-methylenedianiline, but several study limitations were identified (small size and young age of the cohort, and short follow-up period). These limitations should be addressed in additional well-conducted epidemiology studies in occupational settings. This information would provide valuable data on possible adverse effects in humans. There is evidence that 4,4′-methylenedianiline may be released from certain medical devices made of polyurethane such as plasma separators and artificial dialyzers (Shintani 1991). This suggests a potential health risk for uremic patients or patients who receive frequent blood transfusions. Because of the known hepatic effects of 4,4′-methylenedianiline, liver tests should be periodically conducted in these patients as well as in occupationally exposed individuals for early detection of liver toxicity.

**Biomarkers of Exposure and Effect.**

**Exposure.** Exposure to 4,4′-methylenedianiline may be assessed by determining levels of 4,4′-methylenedianiline or N-acetylated metabolites in urine (Cocker et al. 1986a, 1994; Dalene et al. 1995; Robert et al. 1995; Schfitze et al. 1995). However, no quantitative estimates of exposure have been provided. In a study of 411 individuals occupationally exposed, 42% of urine samples examined had no detectable 4,4′-methylenedianiline (Cocker et al. 1994). In another study of 111 workers exposed to 4,4′-methylenedianiline, 81% of over 300 urine samples had concentrations of 4,4′-methylenedianiline below 5 nmol/mmol creatinine, which was below the detection limit of 50 nmol/L (Cocker et al. 1986a). A preferred method for assessing exposure may be measuring hemoglobin adducts. Studies in rats have shown that 4,4′-methylenedianiline can form adducts with hemoglobin which are stable in vivo and persist for the life span of the erythrocyte-120 days in humans (Bailey et al. 1990; Farmer and Bailey 1989). Similar findings have recently been reported in humans (Schiitze et al. 1995). 4,4′-Methylenedianiline can be released from the adduct and quantitated. The advantage of this method is that it provides information that is not restricted to recent
exposure. Preliminary results in rats showed that a dose-response can be established. Further studies on this subject would provide valuable information that could lead to early detection of 4,4’-methyleneedianiline exposure.

**Effect.** There are no specific biomarkers of effects for 4,4’-methyleneedianiline. Many different classes of chemicals are liver toxicants or may cause dermal sensitization. Further studies to identify specific biomarkers of effects of 4,4’-methyleneedianiline would facilitate medical surveillance leading to early detection of potentially adverse health effects and possible treatment.

**Absorption, Distribution, Metabolism, and Excretion.** There is no quantitative information on the rates of absorption of 4,4’-methyleneedianiline in humans following inhalation or oral exposure. A study in volunteers showed that a maximum of approximately 28% of a dose of 4,4’-methyleneedianiline applied dermally for one hour was absorbed (Brunmark et al. 1995). Limited data from workers exposed to 4,4’-methyleneedianiline su,, Ooested that absorption by inhalation is faster than dermal absorption (Cocker et al. 1994). There is also no information on absorption rates in animals following inhalation or oral exposure. Dermal data are available in rats, guinea pigs, and monkeys (El-Hawari et al. 1986). Obtaining additional quantitative data in animals via inhalation and oral routes, and using different vehicles, would be helpful for estimating absorption in humans.

There are no distribution data in humans. Data on distribution via the inhalation and dermal routes for animals were not located. There is limited information describing distribution following acute dermal exposure to 4,4´-methyleneedianiline in animals. Studies indicate that it is distributed among all tissues (El-Hawari et al. 1986). Additional studies regarding repeated dermal exposure would help elucidate the distribution pattern of 4,4’-methyleneedianiline. The dermal route appears to be the most likely route of exposure near hazardous waste sites. Dermal contact is also the most likely route of exposure in manufacturing and process industries.

No information was located regarding metabolism of 4,4´-methyleneedianiline in humans-following inhalation or oral exposure. Data from humans exposed primarily by dermal contact showed that 4,4´-methyleneedianiline can be acetylated *in vivo* to form N-acetylmethyleneedianiline (Cocker et al. 1986a; Robert et al. 1995) and to a small extent, N,N´-diacetylmethyleneedianiline (Robert et al. 1995). Acetylation has been demonstrated in rats exposed orally (Tanaka et al. 1985). No information was located regarding metabolism in animals exposed by inhalation or dermally. Information on
metabolism after dermal exposure in animals would be useful because the potential exists for exposure to occur in humans via this route.

Workers and volunteers exposed to 4,4′-methyleneedianiline by inhalation or by skin contact excreted 4,4′-methyleneedianiline and/or acetylated metabolites in the urine (Brunmark et al. 1995; Cocker et al. 1986a, 1994; Dalene et al. 1995). There is no information regarding excretion in humans after oral exposure. Acute dermal studies in animals showed that urinary excretion is a major excretory pathway, but fecal excretion also occurs (El-Hawari et al. 1986). Only one study was located that provided excretion data in animals following oral exposure (Tanaka et al. 1985); no inhalation data were located. Further examination of excretion patterns in animals following repeated dermal exposures would provide valuable information relevant to humans exposed continuously in the workplace.

**Comparative Toxicokinetics.** There is insufficient information to determine possible differences or similarities in toxicokinetic patterns for 4,4′-methyleneedianiline between humans and animals. The limited data suggest that 4,4′-methyleneedianiline can be acetylated by humans and animals, and that the resulting metabolite is excreted in the urine. Studies have also demonstrated that the liver is a target for 4,4′-methyleneedianiline toxicity in humans and the animal species tested. Acute dermal studies in animals showed some qualitative differences in excretory patterns between species. The urine was the main excretory route in rats and monkeys, whereas guinea pigs excreted similar amounts of 4,4′-methyleneedianiline (or metabolites) in urine and feces (El-Hawari et al. 1986). Physiologically based pharmacokinetic models for 4,4′-methyleneedianiline have not been developed. Further studies are necessary to determine which species might be a suitable animal model.

**Methods for Reducing Toxic Effects.** The mechanism by which 4,4′-methyleneedianiline enters the bloodstream in humans is not known, and there are no established methods for reducing absorption after inhalation and oral exposure other than minimizing exposure. A study of dermal absorption through human skin *in vitro* provided information on decontamination procedures that could potentially be used *in vivo* (Hewitt et al. 1995). Further studies of decontamination strategies by using different washing solutions would be valuable. Animal studies using gastrointestinal sorbents such as activated carbon and resins, which can bind amines, might give insight into additional methods for reducing systemic absorption of this chemical. Suggested methods for treating the effects of acute exposure to 4,4′-methyleneedianiline are generally supportive. Toxic hepatitis that developed after exposure to
4,4′-methylenedianiline has been treated conventionally. There are no established methods for reducing body burdens in humans. It is assumed that metabolism of 4,4′-methylenedianiline leads to the formation of highly reactive and potentially toxic derivatives. Thus, additional studies examining the feasibility of favoring metabolic pathways leading to the formation of nontoxic metabolites (N-acetylation pathway) would be valuable.

2.10.3 Ongoing Studies

Dr. Mary Kanz from the Department of Pathology, University of Texas at Galveston, is conducting research aimed to understand how and why 4,4′-methylenedianiline causes early, selective injury to bile duct cells. She is focusing the research on 4,4′-methylenedianiline metabolites excreted in bile and their capacity to damage biliary epithelial cells (FEDRIP 1997).
3. CHEMICAL AND PHYSICAL INFORMATION

3.1 CHEMICAL IDENTITY

Methylenedianilines are diphenylmethane compounds with an amino group added to each benzene ring. Depending on the position of the amino group, methylenedianilines can exist in six isomeric forms: 2,2′-methylenedianiline; 2,3′-methylenedianiline; 2,4′-methylenedianiline; 3,3′-methylenedianiline; 3,4′-methylenedianiline; and 4,4′-methylenedianiline. Of the six isomers, 2,2′-methylenedianiline and 2,4′-methylenedianiline are produced on a small scale as research chemicals (HSDB 1996). The isomer 4,4′-methylenedianiline is produced in the United States for industrial use. Therefore, this profile will limit its discussion to 4,4′-methylenedianiline. Information regarding the chemical identity of 4,4′-methylenedianiline is presented in Table 3-1.

3.2 PHYSICAL AND CHEMICAL PROPERTIES

Information regarding the physical and chemical properties of 4,4′-methylenedianiline is presented in Table 3-2. Exposure of 4,4′-methylenedianiline to air and light results in polymerization and oxidation of the compound, as evidenced by darkening of color and polymerization leading to the formation of polymeric amines (IARC 1986; Moore 1978). When heated to decomposition, 4,4′-methylenedianiline emits toxic fumes of aniline and nitrogen oxides (NIEHS 1994). It is a weak base (pH 7.7, Allied Chemical Corp. 1978), but its experimentally determined pKₐ value was not located in the literature. The estimated pKₐ for this compound is 4.88 (EPA 1995). Since 4,4′-methylenedianiline is a base, it forms dihydrochloride salts with hydrochloric acid (IARC 1986). The dihydrochloride of 4,4′-methylenedianiline or other salts of the compound with strong inorganic acids will be more water soluble than the free base. The dihydrochloride salt is soluble in water (NIEHS 1994).
Table 3-1. Chemical Identity of 4,4’-Methylenedianiline

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Information</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chemical name</td>
<td>4,4’-Methylenedianiline</td>
<td></td>
</tr>
<tr>
<td>Synonym(s)</td>
<td>4,4’-Methylene-bis(benzeneamine); 4,4’-diaminodiphenylmethane; 4,4’-methylenebisaniline; p,p’-methylenedianiline; MDA; Methyleneedianiline</td>
<td>RTECS 1996</td>
</tr>
<tr>
<td>Registered trade name(s)</td>
<td>Ancamine TL; Epicure DDM; Tonox Jeffamine AP-20</td>
<td>IARC 1986; RTECS 1996</td>
</tr>
<tr>
<td>Chemical formula</td>
<td>C_{13}H_{14}N_{2}</td>
<td>Merck 1989</td>
</tr>
<tr>
<td>Chemical structure</td>
<td>![Chemical Structure Image]</td>
<td>Merck 1989</td>
</tr>
</tbody>
</table>

Identification numbers:
- CAS Registry: 101-77-9
- NIOSH RTECS: BY5425000
- EPA Hazardous Waste: No data
- OHM/TADS: No data
- DOT/UN/NA/IMCO: UN2651
- HSDB: 2541
- NCI: C54604

CAS = Chemical Abstracts Services; DOT/UN/NA/IMCO = Department of Transportation/United Nations/North America/International Maritime Dangerous Goods Code; EPA = Environmental Protection Agency; HSDB = Hazardous Substances Data Bank; IARC = International Agency for Research on Cancer; NCI = Hazardous Substances Data Bank; NCI = National Institute for Occupational Safety and Health; OHM/TADS = Oil and Hazardous Materials/Technical Assistance Data System; RTECS = Registry of Toxic Effects of Chemical Substances.
<table>
<thead>
<tr>
<th>Property</th>
<th>Information</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Molecular weight</td>
<td>198.27</td>
<td>Lide 1994</td>
</tr>
<tr>
<td>Color</td>
<td>Colorless to pale yellow or light brown</td>
<td>IARC 1986; Lewis 1993</td>
</tr>
<tr>
<td>Physical state</td>
<td>Crystalline solid</td>
<td>Moore 1978</td>
</tr>
<tr>
<td>Melting point</td>
<td>92–3 °C</td>
<td>Lide 1994</td>
</tr>
<tr>
<td>Boiling point</td>
<td>398–9 °C</td>
<td>Lide 1994</td>
</tr>
<tr>
<td>Specific gravity</td>
<td>1.1</td>
<td>IARC 1986</td>
</tr>
<tr>
<td>Odor</td>
<td>Faint amine-like</td>
<td>Moore 1978</td>
</tr>
<tr>
<td>Odor threshold: Water</td>
<td>No data</td>
<td></td>
</tr>
<tr>
<td>Odor threshold: Air</td>
<td>No data</td>
<td></td>
</tr>
<tr>
<td>Solubility: Water at 25 °C</td>
<td>1,000 mg/L; slightly soluble</td>
<td>Moore 1978; Merck 1989</td>
</tr>
<tr>
<td>Solubility: Organic solvent(s)</td>
<td>Very soluble in ethanol, benzene and diethyl ether</td>
<td>Merck 1989; Lide 1994</td>
</tr>
<tr>
<td>Partition coefficients:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Log $K_{ow}$</td>
<td>1.59</td>
<td>HSDB 1996</td>
</tr>
<tr>
<td>Log $K_{oc}$</td>
<td>2.24 (estimated)</td>
<td>Lyman 1990</td>
</tr>
<tr>
<td>Vapor pressure: at 25 °C</td>
<td>2.15x10$^7$ mm Hg (estimated from H and water solubility)</td>
<td>Thomas 1990</td>
</tr>
<tr>
<td>at 197 °C</td>
<td>1 mm Hg</td>
<td>IARC 1986</td>
</tr>
<tr>
<td>Henry's law constant (H) at 25 °C</td>
<td>5.6x10$^{11}$ atm-m$^3$/mole (estimated)</td>
<td>HSDB 1996</td>
</tr>
<tr>
<td>Autoignition temperature</td>
<td>No data</td>
<td></td>
</tr>
<tr>
<td>Flashpoint</td>
<td>221.1 °C (closed cup); 220 °C (closed cup)</td>
<td>Moore 1978; NFPA 1994</td>
</tr>
<tr>
<td>Flammability limits at 25 °C</td>
<td>No data</td>
<td></td>
</tr>
<tr>
<td>NFPA Hazard Class:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Health</td>
<td>3 (material extremely hazardous to health)</td>
<td>NFPA 1994</td>
</tr>
<tr>
<td>Flammability</td>
<td>1 (material must be preheated before ignition)</td>
<td></td>
</tr>
<tr>
<td>Reactivity</td>
<td>0 (material normally stable even under fire conditions and is not reactive with water)</td>
<td></td>
</tr>
<tr>
<td>Conversion factors (25 °C)</td>
<td>1 mg/m$^3$ = 8.11 ppm</td>
<td>IARC 1986</td>
</tr>
<tr>
<td></td>
<td>1 ppm = 0.123 mg/m$^3$</td>
<td></td>
</tr>
<tr>
<td>Explosive limits</td>
<td>No data</td>
<td></td>
</tr>
</tbody>
</table>

NFPA = National Fire Protection Association
4. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

4.1 PRODUCTION

4,4´-Methylenedianiline is produced by the condensation of formaldehyde with aniline in the presence of an acid catalyst. The reaction produces a mixture of di-, tri-, and poly-methyleneanilines. Aniline is removed from the reaction mixture by distillation. The percentage of 4,4´-methylenedianiline in the mixtures manufactured varies from producer to producer. 4,4´-Methylenedianiline can be isolated from the residual mixture by crystallization with a suitable solvent (IARC 1986; Merck 1989; Moore 1978). 4,4´-Methylenedianiline commercially available in bulk quantities contains approximately 96% 4,4´-methylenedianiline, 3% other isomeric amines, and traces of aniline (IARC 1986).

The quantity of 4,4´-methylenedianiline produced by six manufacturers was approximately 230 million pounds (104 million kg) in 1981 (CMA 1982). In 1982, approximately 90-180 million kg (200-400 million pounds) of 4,4´-methylenedianiline was produced in the United States by a total of seven manufacturers (NIEHS 1994; NIOSH 1986). According to data in Toxics Release Inventory (TRI), four companies produced 4,4´-methylenedianiline in the United States in 1994 for distribution and sale, as well as their own use, captively or otherwise (TR194 1996): Dow Chemicals, U.S.A., La Porte, Texas; an unidentified company in New Martinsville, West Virginia; Rubicon, Inc., Geismar, Louisiana; and Uniroyal Chemical Co., Inc., Naugatuck, Connecticut. Another report indicates three companies producing 4,4´-methylenedianiline in the United States (SRI 1994): Dow Chemicals, U.S.A., La Porte, Texas; Bayer Corporation, Polymers Division, New Martinsville, West Virginia; and Uniroyal Chemical Co., Inc., Naugatuck, Connecticut.

The data on production capacity or the amount of 4,4´-methylenedianiline produced in the United States in recent years are not available. Table 4-1 lists the facilities in each state that manufacture or process 4,4´-methylenedianiline, the intended use, and the range of maximum amounts of 4,4´-methylenedianiline that are stored on site. The data listed in Table 4-1 are derived from the Toxics Release Inventory (TR194 1996). Only certain types of facilities were required to report; therefore, this is not an exhaustive list.
Table 4-1. Facilities That Manufacture or Process 4,4′-Methylenedianiline

<table>
<thead>
<tr>
<th>Facility</th>
<th>Location*</th>
<th>Range of maximum amounts on site in pounds</th>
<th>Activities and uses</th>
</tr>
</thead>
<tbody>
<tr>
<td>CIBA GEIGY CORP.</td>
<td>MC INTOSH, AL</td>
<td>100,000-999,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>A. O. SMITH CORP.</td>
<td>LITTLE ROCK, AR</td>
<td>10,000-99,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>A. O. SMITH CORP.</td>
<td>LITTLE ROCK, AR</td>
<td>1,000-9,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>NA</td>
<td>MAGNOLIA, AR</td>
<td>10,000-99,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>HEXCEL CORP.</td>
<td>CHATSWORTH, CA</td>
<td>10,000-99,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>AIR PRODS. &amp; CHEMICALS INC.</td>
<td>LOS ANGELES, CA</td>
<td>10,000-99,999</td>
<td>As a reactant; Produce; For on-site use/processing; For sale/distribution; As a formulation component</td>
</tr>
<tr>
<td>UNIROYAL CHEMICAL CO. INC.</td>
<td>CT</td>
<td>100,000-999,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>UOP</td>
<td>MC COOK, IL</td>
<td>10,000-99,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>ALLCO ACQUISITIONS</td>
<td>ELK GROVE VILLAGE, IL</td>
<td>10,000-99,999</td>
<td>As a reactant</td>
</tr>
<tr>
<td>AIR PRODS. &amp; CHEMICALS INC.</td>
<td>WICHITA, KS</td>
<td>1,000,000-9,999,999</td>
<td>As a reactant; Produce; For on-site use/processing; As a reactant</td>
</tr>
<tr>
<td>A. O. SMITH CORP.</td>
<td>WICHITA, KS</td>
<td>10,000-99,999</td>
<td>As a reactant; Produce; For on-site use/processing; For sale/distribution; As a reactant</td>
</tr>
<tr>
<td>BASF CORP.</td>
<td>GEISMAR, LA</td>
<td>100,000-999,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>RUBICON INC.</td>
<td>GEISMAR, LA</td>
<td>1,000,000-999,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>NA</td>
<td>SAINT LOUIS, MO</td>
<td>100,000-999,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>COOK COMPOSITES &amp; POLYMERS CO.</td>
<td>NORTH KANSAS CITY, MO</td>
<td>1,000-9,999</td>
<td>As a reactant; Import; For on-site use/processing; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>UNIROYAL CHEMICAL CO. INC.</td>
<td>GASTONIA, NC</td>
<td>10,000-99,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>COOKSON AMERICA</td>
<td>GRANITE FALLS, NC</td>
<td>1,000-9,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>AMOCO CORP.</td>
<td>PIEDMONT, SC</td>
<td>10,000-99,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>AMERON INC.</td>
<td>SPARTANBURG, SC</td>
<td>1,000-9,999</td>
<td>As a reactant; Produce; For on-site use/processing; As a reactant</td>
</tr>
<tr>
<td>DOW CHEMICAL CO.</td>
<td>LA PORTE, TX</td>
<td>1,000,000-9,999,999</td>
<td>As a reactant; Produce; For on-site use/processing; For sale/distributi</td>
</tr>
<tr>
<td>NA</td>
<td>BAYTOWN, TX</td>
<td>1,000,000-9,999,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>AMERON INC.</td>
<td>BURKBNURNETT, TX</td>
<td>10,000-99,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>HERCULES INC.</td>
<td>CLEARFIELD, UT</td>
<td>10,000-99,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>3M CO.</td>
<td>PRAIRIE DU CHIEN, WI</td>
<td>10,000-99,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
<tr>
<td>RPM INC.</td>
<td>GREEN BAY, WI</td>
<td>1,000-9,999</td>
<td>As a reactant; As a reactant; As a formulation component</td>
</tr>
</tbody>
</table>
Table 4-1. Facilities That Manufacture or Process 4,4'-Methylenedianiline (continued)

<table>
<thead>
<tr>
<th>Facility</th>
<th>Location*</th>
<th>Range of maximum amounts on site in pounds</th>
<th>Activities and uses</th>
</tr>
</thead>
<tbody>
<tr>
<td>NA</td>
<td>NEW MARTINSVILLE, WV</td>
<td>1,000,000-9,999,999</td>
<td>Produce; For on-site use/processing; For sale/distribution; As a reactant</td>
</tr>
<tr>
<td>BASF CORP.</td>
<td>HUNTINGTON, WV</td>
<td>1,000-9,999</td>
<td>Produce; For on-site use/processing; As a reactant</td>
</tr>
</tbody>
</table>

Source: TRI94 1996

* Post office state abbreviations used

NA = not available
4.2 IMPORT/EXPORT

The exports of 4,4'-methylenedianiline from the United States to other countries in 1989, 1990, 1991, 1992 and 1993 were 13.1 million kg (28.9 million pounds), 13.5 million kg (29.8 million pounds), 5.8 million kg (12.8 million pounds), 7.1 million kg (15.7 million pounds), and 4.5 million kg (9.9 million pounds), respectively (NTDB 1994). There was a marked decrease in amounts of 4,4'-methylenedianiline exported to other countries during the period 1990-1993. The imports of 4,4'-methylenedianiline from other countries to the United States in 1989, 1990, 1991, 1992, and 1993 were 1.5 million kg (3.3 million pounds), 1.3 million kg (2.9 million pounds), 1.1 million kg (2.4 million pounds), 0.9 million kg (2.0 million pounds), and 0.5 million kg (1.1 million pounds), respectively (NTDB 1994). These figures indicate a continual and gradual decrease in the amounts of 4,4'-methylenedianiline imported into the United States from other countries from 1989 to 1993.

4.3 USE

Over 90% of 4,4'-methylenedianiline produced in the United States is used captively for the production of 4,4'-methylenedianiline diisocyanate and other polymeric isocyanates (IARC 1986). These di- or poly-isocyanates are used in a variety of polymer and resin production, including polyurethane foam, isocyanate resins and elastomer (e.g., Spandex® fiber). Small amounts of 4,4'-methylenedianiline are used as an azo dye intermediate; as a chemical reagent for the determination of tungsten and sulfates; as a corrosion inhibitor; as an antioxidant and curative agent in rubber; as a raw material in the production of resins; and as an epoxy-resin hardening agent in adhesives, encapsulants, coatings, filament windings and binders (CMA 1982; IARC 1986; Lewis 1993; Tucker et al. 1993).

4.4 DISPOSAL

Incineration is one of the feasible methods for disposal of wastes containing 4,4'-methylenedianiline. Gas-fired incinerators in which first-stage combustion takes place with a less than stoichiometric airfuel ratio, followed by a second-stage combustion with excess air, are suitable for disposal of 4,4'-methylenedianiline wastes (HSDB 1996). The temperature and the residence time inside the combustion zone of the incinerators should be such that they ensure complete destruction (>99.99%) of the compound (HSDB 1996).
5. POTENTIAL FOR HUMAN EXPOSURE

5.1 OVERVIEW

4,4’-Methylenedianiline is released to air and water during industrial production and use of this compound (IARC 1986). 4,4’-Methylenedianiline may also be released in water as a result of hydrolysis of treated and untreated 4,4’-methylenediphenyl diisocyanate wastewater discharged into surface water or public sewers. However, there are no monitoring data that confirm the presence of 4,4’-methylenedianiline in waste effluents from the plants that manufacture and use 4,4’-methylenedianiline, or in natural waters where the treated or untreated wastewaters from these plants are discharged. 4,4’-Methylenedianiline may be found in soils of controlled or uncontrolled hazardous waste sites as a result of disposal of wastes containing either 4,4’-methylenedianiline or products such as 4,4’-methylenediphenyl diisocyanate. 4,4’-Methylenedianiline may also be released to soil as a result of accidental spillage during storage or transport. Smaller amounts of 4,4’-methylenedianiline may be released to water and soil as a result of terrestrial and hydrospheric precipitation and scavenging of atmospheric 4,4’-methylenedianiline. However, very few monitoring data are available that confirm the presence of 4,4’-methylenedianiline in soil.

Experimental data on the environmental fate of 4,4’-methylenedianiline are scarce. Most conclusions about the environmental fate and transport of 4,4’-methylenedianiline are estimated, based on its physical and chemical properties. In air, the compound will primarily exist as particulate matter and will be removed from the atmosphere by dry deposition and scavenging by rain and snow. The small amount of 4,4’-methylenedianiline present in the vapor phase in the atmosphere will be transformed by reaction with hydroxyl radicals with an estimated half-life of 1.6 hours. In water and soil, the material will be degraded primarily via biodegradation. The estimated half-lives of biodegradation in surface water, groundwater and soil are 1-7 days, 2-14 days, and 1-7 days, respectively. 4,4’-Methylenedianiline will covalently bind with humic materials in soil and water. As a result, it will be present predominantly in the bound form in sediment and suspended solids in water, and its leaching from most soils will not be important. Three factors indicate that 4,4’-methylenedianiline will neither bioconcentrate in aquatic organisms nor biomagnify in terrestrial or aquatic food chains: (1) the experimental bioconcentration factor in carp (Caprinus carpio); (2) the estimated bioconcentration factor and octanol/water partition coefficient (indicator of lipid solubility), and (3) the fact that it is metabolized quickly in higher trophic level animals.
No monitoring data exist on the levels of 4,4’-methylenedianiline in ambient air, surface water, industrial effluents, soil or any fruits and vegetables. Levels of the compound in occupational air have been measured; and the levels rarely exceed the American Conference of Governmental Industrial Hygienists (ACGIH) threshold limit value of 0.8 mg/m³ for an 8-hour weighted average concentration. 4,4’-Methylenedianiline and its metabolites have also been measured in urine of workers. The maximum urinary 4,4’-methylenedianiline concentration of 6,871 nmol/mmol creatinine was detected in a worker at a manufacturing plant in Britain (Cocker et al. 1994).

4,4’-Methylenedianiline has been found in none of the 1,445 current or former EPA National Priorities List (NPL) hazardous wastes sites (HazDat 1997). However, the number of sites evaluated for methylenedianiline is not known.

5.2 RELEASES TO THE ENVIRONMENT

Anthropogenic releases of 4,4’-methylenedianiline are primarily to the soil via underground injection. According to the Toxics Chemical Release Inventory, in 1994, a total of 36,531 pounds of 4,4’-methylenedianiline were released to the environment (air, water, soil) from 27 large processing facilities (TR194 1996). Table 5-1 lists the amounts released from these facilities. In addition, an estimated 1,889 pounds of 4,4’-methylenedianiline were released by these manufacturing and processing facilities to publicly owned treatment works (POTWs), and an estimated 184,458 pounds were transferred off-site (TR194 1996). The data listed in the TRI should be used with caution because only certain types of facilities are required to report (EPA 1996). This is not an exhaustive list. Manufacturing and processing facilities are required to report information to the Toxics Release Inventory if they employ 10 or more full-time employees; if their facility is classified under Standard Industrial Classification (SIC) codes 20 through 39; and if their facility produces, imports, or processes 25,000 or more pounds of any TRI chemical or otherwise uses more than 10,000 pounds of a TRI chemical in a calendar year (EPA 1996).

5.2.1 Air

4,4’-Methylenedianiline is released to the air during the industrial production of this compound (IARC 1986). It is also released into the air during industrial production processes that use 4,4’-methylenedianiline as a component. For example, it is released to the air during the production of reinforced
Table 5-1. Releases to the Environment from Facilities That Manufacture or Process 4,4'-Methylenedianiline

<table>
<thead>
<tr>
<th>State</th>
<th>City</th>
<th>Facility</th>
<th>Air</th>
<th>Water</th>
<th>Land</th>
<th>Underground injection</th>
<th>Total environment[^a]</th>
<th>POTW transfer</th>
<th>Off-site waste transfer</th>
</tr>
</thead>
<tbody>
<tr>
<td>AL</td>
<td>MC INTOSH</td>
<td>CIBA GEIGY CORP.</td>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td>4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AR</td>
<td>LITTLE ROCK</td>
<td>A. O. SMITH CORP.</td>
<td>1</td>
<td></td>
<td>1</td>
<td></td>
<td>1</td>
<td>2,700</td>
<td></td>
</tr>
<tr>
<td>AR</td>
<td>LITTLE ROCK</td>
<td>A. O. SMITH CORP.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>180</td>
<td></td>
</tr>
<tr>
<td>AR</td>
<td>MAGNOLIA</td>
<td>NA</td>
<td>1,000</td>
<td></td>
<td></td>
<td></td>
<td>1,000</td>
<td>750</td>
<td></td>
</tr>
<tr>
<td>CA</td>
<td>CHATSWORTH</td>
<td>HEXCEL CORP.</td>
<td>255</td>
<td></td>
<td></td>
<td></td>
<td>255</td>
<td>2,708</td>
<td></td>
</tr>
<tr>
<td>CA</td>
<td>LOS ANGELES</td>
<td>AIR PRODS. &amp; CHEMICALS INC.</td>
<td>28</td>
<td></td>
<td></td>
<td></td>
<td>28</td>
<td>72</td>
<td>34,680</td>
</tr>
<tr>
<td>CT</td>
<td>NAUGATUCK</td>
<td>UNIROYAL CHEMICAL CO. INC.</td>
<td>500</td>
<td></td>
<td></td>
<td></td>
<td>500</td>
<td>402</td>
<td>10,650</td>
</tr>
<tr>
<td>IL</td>
<td>ELK GROVE VILLAGE</td>
<td>ALLCO ACQUISITIONS</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IL</td>
<td>MC COOK</td>
<td>UOP</td>
<td>250</td>
<td></td>
<td></td>
<td></td>
<td>250</td>
<td>250</td>
<td>250</td>
</tr>
<tr>
<td>KS</td>
<td>WICHITA</td>
<td>AIR PRODS. &amp; CHEMICALS INC.</td>
<td>2,400</td>
<td></td>
<td></td>
<td></td>
<td>2,464</td>
<td>56</td>
<td></td>
</tr>
<tr>
<td>KS</td>
<td>WICHITA</td>
<td>A. O. SMITH CORP.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>LA</td>
<td>GEISMAR</td>
<td>BASF CORP.</td>
<td>11</td>
<td></td>
<td></td>
<td></td>
<td>11</td>
<td>6,900</td>
<td></td>
</tr>
<tr>
<td>LA</td>
<td>GEISMAR</td>
<td>RUBICON INC.</td>
<td>20</td>
<td></td>
<td></td>
<td></td>
<td>26,000</td>
<td>26,020</td>
<td>27,910</td>
</tr>
<tr>
<td>MO</td>
<td>NORTH KANSAS CITY</td>
<td>COOK COMPOSITES &amp; POLYMERS CO.</td>
<td>10</td>
<td></td>
<td></td>
<td></td>
<td>10</td>
<td>5</td>
<td></td>
</tr>
<tr>
<td>MO</td>
<td>SAINT LOUIS</td>
<td>NA</td>
<td>255</td>
<td></td>
<td></td>
<td></td>
<td>255</td>
<td>750</td>
<td></td>
</tr>
<tr>
<td>NC</td>
<td>GASTONIA</td>
<td>UNIROYAL CHEMICAL CO. INC.</td>
<td>250</td>
<td></td>
<td></td>
<td></td>
<td>250</td>
<td>250</td>
<td></td>
</tr>
<tr>
<td>NC</td>
<td>GRANITE FALLS</td>
<td>COOKSON AMERICA</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SC</td>
<td>PIEDMONT</td>
<td>AMOCO CORP.</td>
<td>18</td>
<td></td>
<td></td>
<td></td>
<td>18</td>
<td>5</td>
<td>118</td>
</tr>
<tr>
<td>SC</td>
<td>SPARTANBURG</td>
<td>AMERON INC.</td>
<td>296</td>
<td></td>
<td></td>
<td></td>
<td>296</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
### Table 5-1. Releases to the Environment from Facilities That Manufacture or Process 4,4'-Methylenedianiline

<table>
<thead>
<tr>
<th>State</th>
<th>City</th>
<th>Facility</th>
<th>Air</th>
<th>Water</th>
<th>Land</th>
<th>Underground injection</th>
<th>Total environment&lt;sup&gt;b&lt;/sup&gt;</th>
<th>POTW transfer</th>
<th>Off-site waste transfer</th>
</tr>
</thead>
<tbody>
<tr>
<td>TX</td>
<td>BAYTOWN</td>
<td>NA</td>
<td>1,505</td>
<td>250</td>
<td></td>
<td></td>
<td>1,755</td>
<td></td>
<td>8,600</td>
</tr>
<tr>
<td>TX</td>
<td>BURKBURNETT</td>
<td>AMERON INC.</td>
<td>120</td>
<td></td>
<td></td>
<td></td>
<td>120</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TX</td>
<td>LA PORTE</td>
<td>DOW CHEMICAL CO.</td>
<td>95</td>
<td></td>
<td></td>
<td></td>
<td>95</td>
<td></td>
<td>9,732</td>
</tr>
<tr>
<td>UT</td>
<td>CLEARFIELD</td>
<td>HERCULES INC.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>57,910</td>
</tr>
<tr>
<td>WI</td>
<td>GREEN BAY</td>
<td>RPM INC.</td>
<td>250</td>
<td></td>
<td></td>
<td></td>
<td>250</td>
<td></td>
<td>500</td>
</tr>
<tr>
<td>WI</td>
<td>PRAIRIE DU CHIEN</td>
<td>3M CO.</td>
<td>5</td>
<td></td>
<td></td>
<td></td>
<td>5</td>
<td></td>
<td>18,000</td>
</tr>
<tr>
<td>WV</td>
<td>HUNTINGTON</td>
<td>BASF CORP.</td>
<td>310</td>
<td></td>
<td></td>
<td></td>
<td>310</td>
<td>1,160</td>
<td>1,240</td>
</tr>
<tr>
<td>WV</td>
<td>NEW MARTINSVILLE</td>
<td>NA</td>
<td>2,159</td>
<td>475</td>
<td></td>
<td></td>
<td>2,634</td>
<td></td>
<td>250</td>
</tr>
<tr>
<td></td>
<td><strong>Totals</strong></td>
<td></td>
<td>9,742</td>
<td>725</td>
<td>26,064</td>
<td>36,531</td>
<td>1,889</td>
<td>184,458</td>
<td></td>
</tr>
</tbody>
</table>

Source: TRI94 1996

<sup>a</sup> Post office state abbreviations used

<sup>b</sup> The sum of all releases of the chemical to air, land, water, and underground injection wells by a given facility

NA = not available; POTW = publicly owned treatment works
plastics, during encapsulation of instruments with polyurethane mixture, and during coremaking of iron and steel foundries where 4,4´-methyleneedianiline polymers are used as binders in molding (IARC 1986).

According to the Toxics Chemical Release Inventory (TRI) in 1994, releases of 4,4´-methylenedianiline to the air from 27 large processing facilities were 9,742 pounds (4,419 kg) (TR194 1996). Release to the air constituted 26.7% of the total environmental release. Table 5-l lists amounts released from these facilities. The TRI data should be used with caution because only certain types of facilities are required to report. This is not an exhaustive list (EPA 1996).

5.2.2 Water

4,4´-Methylenedianiline is released to water as aqueous effluents from industries that produce or use it, for example, from methylenediphenyl diisocyanate manufacturing plants. The source of 4,4´-methyleneedianiline in treated and untreated waste water from methylenediphenyl diisocyanate plants may be due to release of the starting material (4,4´-methylenedianiline) in waste water or from the hydrolysis of methylenediphenyl diisocyanate. Small amounts of 4,4´-methylenedianiline may also be released to surface water via storm water runoff from waste sites containing this compound. However, there is no monitoring data that confirm the presence of the compound in waste effluents from the plants that manufacture and use it, or in natural waters where the treated or untreated wastewaters from these plants are discharged.

According to the TRI, in 1994 releases of 4,4´-methylenedianiline to water from the 27 large processing facilities totaled 725 pounds (329 kg) (TR194 1996). Release to water constituted 2% of the total environmental release. An additional 1,889 pounds (857 kg) were released indirectly to POTWs and some of this volume ultimately may have been released to surface waters. Table 5-l lists amounts released from these facilities. The TRI data should be used with caution because only certain types of facilities are required to report. This is not an exhaustive list (EPA 1996).III

5.2.3 Soil

4,4´-Methylenedianiline may be found in soils of controlled or uncontrolled hazardous waste sites as a result of disposal of wastes containing either 4,4´-methylenedianiline or reaction products, such as
methylenediphenyl diisocyanate. Products such as polyurethane foams which contain the reactant diphenylisocyanate moiety (NIOSH 1976) can break down into 4,4´-methylenedianiline either chemically or biologically in soil. 4,4´-Methylenedianiline may also be released to soil as a result of accidental spillage during storage or transport. However, very few monitoring data are available that confirm the presence of 4,4´-methylenedianiline in soil.

According to the TRI, in 1994 there were no releases of 4,4´-methylenedianiline to soil from the 27 large processing facilities (TR194 1996). However, an additional 26,064 pounds (11,818 kg) was released by underground injection. These releases represent 71% of all environmental releases. Table 5-l lists amounts released from these facilities. The TRI data should be used with caution because only certain types of facilities are required to report. This is not an exhaustive list (EPA 1996).

5.3 ENVIRONMENTAL FATE

5.3.1 Transport and Partitioning

The vapor pressure of 4,4´-methylenedianiline at 25 °C has been estimated to be $2.82 \times 10^{-10}$ atm (2.15x$10^{-7}$ mm Hg) by multiplying its water solubility of 5.04 moles/m$^3$ by its Henry’s law constant (H) of $5.6 \times 10^{-11}$ atm-m$^3$/mole (H = vapor pressure/water solubility) (Thomas 1990a). This is comparable to an estimated vapor pressure of $1.5 \times 10^{-7}$ mm Hg at 25 °C (NIOSH 1986). Organic compounds with a vapor pressure of $2.15 \times 10^{-7}$ mm Hg will exist in the air mostly as an aerosol and partly in the vapor phase (Eisenreich et al. 1981). In a test atmosphere generated by vaporization of epoxy resin containing 4,4´-methylenedianiline hardener, no difference in the sample collection efficiency for 4,4´-methylenedianiline was observed when sulfuric acid-coated glass-fiber filters and simple Teflon filters were used (Gunderson and Anderson 1988). This indicates that 4,4´-methylenedianiline will exist primarily in the aerosol phase in the atmosphere and, like other atmospheric aerosol, will be removed from the atmosphere by rain/snow scavenging and dry deposition (Bidleman 1988).

4,4´-Methylenedianiline has an estimated $K_{oc}$ value of 174 (see Table 3-2). It will be weakly to moderately adsorbed to suspended solids and sediment in water (Swarm et al. 1983), and a large percentage of 4,4´-methylenedianiline may exist in water in the dissolved state where it is susceptible
to degradation via chemical/biological processes. As the water solubility of amine salts is higher than the free base, the concentration of dissolved 4,4’-methyleneedianiline will increase in natural waters as the pH decreases below 7. However, aromatic amines, particularly primary aromatic amines, covalently and irreversibly bind to humic substances present in most natural waters (Parris 1980). Therefore, in deference to moderate/low physical adsorption, 4,4’-methyleneedianiline will become strongly bound (through covalent bonds) to humic materials in suspended solids and sediment present in most waters. Therefore, the percentage of 4,4’-methyleneedianiline present in water may be much lower than is expected from simple physical adsorption of the compound. Organic compounds with Henry’s law constants <3.7x10^{-7} atm-m^3/mole are essentially non-volatile in water (Thomas 1990b). Therefore, 4,4’-methyleneedianiline, with an estimated H of 5.99x10^{-11} atm-m^3/mole (see Table 3-2), will remain essentially non-volatile in water.

Based on a value of 1.59 for log K\textsubscript{ow} and a regression equation (Bysshe 1990), the estimated bioconcentration factor for 4,4’-methyleneedianiline in fathead minnow, bluegill sunfish, rainbow trout, and mosquitofish is 9.5. Therefore, 4,4’-methyleneedianiline will not bioconcentrate in aquatic organisms. Carp (Cyprinus \textit{carpio}) were grown in a model river consisting of natural river water, 0.5% volume per volume (v/v) river bottom sludge, and 0.1 mg/L methylene-di-p-phenylene isocyanate (MDI) in an outdoor stainless steel tank for 8 weeks with water flowing in the tank at rates of 4-14 cm/sec (III 1981). Neither MD1 nor its decomposition product, 4,4’-methyleneedianiline, was detected (detection limit <0.1 mg/kg) in the whole body of fish. It was concluded that MD1 and 4,4’-methyleneedianiline do not bioaccumulate in carp (III 1981). No data were located in the literature that would suggest that 4,4’-methyleneedianiline will biomagnify in animals of higher trophic level via food chain biotransfer (e.g., bioaccumulation in algae < bioaccumulation in fish < bioaccumulation in human). This is not surprising, considering the low K\textsubscript{ow} value (indicative of low accumulation in lipids) and easy metabolism of the compound (Cocker et al. 1994) in higher trophic level animals.

The estimated K\textsubscript{oc} value of 174 indicates that the mobility of 4,4’-methyleneedianiline in soils having low organic carbon content will be moderate to high (Swarm et al. 1983). However, besides the physical adsorption to organic matter in soils, the compound will also become bound to organic matter (humates) by stronger covalent bonds (Parris 1980). In soils that exhibit this covalent bonding behavior, the mobility of 4,4’-methyleneedianiline will be low and the rate of leaching from soil to groundwater will not be important.
5.3.2 Transformation and Degradation

5.3.2.1 Air

Reactions with hydroxyl radicals, ozone and nitric acid may be important for 4,4´-methylenedianiline in the atmosphere (Atkinson 1985). The rate constant for the homogeneous vapor phase reaction of 4,4´-methylenedianiline with hydroxyl radicals at 25 °C is estimated to be 2.4lx 10^{-10} cm³/molecule-sec (HSDB 1996). The estimated value is in conformity with a value of 1.18x10^{-10} for reaction of aniline with hydroxyl radicals (Atkinson 1985). Assuming the atmospheric concentration of hydroxyl radicals to be constant at 5x10^{-5} molecules/cm³, the half-life of 4,4´-methylenedianiline for this pseudo first order reaction is 1.6 hours. However, since the compound will be present predominantly in the aerosol phase (see Section 5.3.1), the importance of the heterogeneous reaction of particulate 4,4´-methylenedianiline with gas phase hydroxyl radicals cannot be ascertained. Based on the rate constants for the reactions of primary amines with ozone (Atkinson and Carter 1984), the atmospheric reaction of 4,4´-methylenedianiline with ozone will not be important. On the other hand, based on the reaction of amines with atmospheric nitric acid (Atkinson 1985), the reaction of 4,4´-methylenedianiline with gas phase nitric acid may be an important loss process in urban atmospheres (where the concentration of atmospheric nitric acid is much higher than the concentration of nitric acid in rural atmospheres). But the importance of the heterogeneous reaction of particulate 4,4´-methylenedianiline with gas phase nitric acid in the atmosphere cannot be quantitatively ascertained.

5.3.2.2 Water

The ultraviolet absorption spectra of 4,4´-methylenedianiline solution in methanol shows an absorption maximum at 243 nm, and a much weaker maximum at 289 nm (Sadtler 1974). The weaker absorption extends well into wavelengths >290 nm. Therefore, solutions of 4,4´-methylenedianiline in water will absorb some sunlight (wavelength >290 nm) and may undergo photolysis in water. In water with pH <7, the absorption maximum shifts from 289 nm to a lower wavelength (261 nm), with little absorption at wavelengths 2290 nm (Sadtler 1974). Therefore, in water with pH <7, photolysis of 4,4´-methylenedianiline will not occur. Even in natural surface waters having pH ≥7, photolysis may not be important because 4,4´-methylenedianiline will be present predominantly in the adsorbed form in the sediment (see Section 5.3.1).
The hydrolysis of 4,4′-methylenedianiline in water will not be important because aromatic amines are generally resistant to hydrolysis (Harris 1990). Aromatic amines react with hydroxyl and peroxo radicals present in most waters found in the environment, and the pseudo-first-order half-lives of 4,4′-methylenedianiline due to these reactions are estimated to be upwards of 2.6 days (Howard et al. 1991). Based on aqueous screening test data for toluidines, the half-lives for biodegradation of 4,4′-methylenedianiline in surface waters under aerobic and anaerobic conditions have been estimated to be 1-7 days and 4-28 days, respectively (Howard et al. 1991). Similarly, the half-life for biodegradation of 4,4′-methylenedianiline in groundwater has been estimated to be 2-14 days (Howard et al. 1991). In an experimental biodegradation study in water, it was concluded that 4,4′-methylenedianiline (concentration 1.35 mg/L) was not biodegradable in 87 days with activated sludge alone (III 1986). Biodegradation occurred when the sludge was adapted with aniline and 4,4′-methylenedianiline and the rate of biodegradation was maximum when the adaptation period was >200 days. The biodegradation of 4,4′-methylenedianiline was concluded to be a co-metabolic process (III 1986) and the adapted sludge biodegraded 4,4′-methylenedianiline from its initial concentration of 12.7 mg/L to undetectable levels (detection limit <0.02 mg/L) in 8 days (III 1987). Under aerobic conditions, the nitrogen in 4,4′-methylenedianiline oxidized to ammonia and finally to nitrate (III 1987). There was some evidence of phenol formation, but this could not be confirmed. The degradation of 4, 4′-methylenedianiline in water was studied in a model river system (for a description of the model river, see Section 5.3.1). Methylene-di-p-phenylene isocyanate (MDI) was first added to the water; it degraded to form 4,4′-methylenedianiline at an initial concentration of 0.1 mg/L. After 4 days, the concentration of 4,4′-methylenedianiline was below the analytical methods detectable limit (<0.02 mg/L) (III 1980). It was concluded that 4,4′-methylenedianiline will biodegrade rapidly in most natural bodies of water. The main drawbacks of the International Isocyanate Institute (III 1980) study are inadequacy of the performed control tests, failure to identify the products of biodegradation, and failure to provide any mass balance of the original substrates with the products formed.

5.3.2.3 Sediment and Soil

No data, either experimental or estimated, were located for the rate of 4,4′-methylenedianiline loss from soil by reaction with oxidants and natural sunlight. The importance of photolysis beyond the surface layer of soil can be ruled out because of lack of light availability. The reaction of 4,4′-methylenedianiline with nitric acid (see Section 5.3.2.1) in soil, particularly in soil having pH <7, is an interesting subject that needs investigation. The pseudo-first-order aerobic biodegradation half-
life of 4,4´-methyleneedianiline in soil has been estimated to be 1-7 days (Howard et al. 1991). In deeper soil or sediment where there is a lack of oxygen, 4,4´-methyleneedianiline may be degraded by anaerobic biodegradation, but the rate of this reaction was not located in the literature.

5.4 LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT

5.4.1 Air

No monitoring data for the levels of 4,4´-methyleneedianiline in ambient air were located in the literature.

5.4.2 Water

No data regarding the level of 4,4´-methyleneedianiline in surface water, industrial effluents, groundwater, or drinking water were located in the literature.

5.4.3 Sediment and Soil

No data were located on the level of 4,4´-methyleneedianiline in contaminated soil or sediment.

5.4.4 Other Environmental Media

No data were located in the literature on the levels of 4,4´-methyleneedianiline in edible fish or aquatic organisms from contaminated water, fruits, or vegetables grown on contaminated land, and in other related environmental media. No residue of MD1 or 4,4´-methyleneedianiline was found at a detection limit of 0.01 mg/kg (fresh weight) in tomato, cucumber, melon, and lettuce grown on soilless reconstituted polyurethane foam with or without nutrient solutions (Rouchaud et al. 1992).

5.5 GENERAL POPULATION AND OCCUPATIONAL EXPOSURE

The consumption of bread accidentally contaminated with 4,4´-methyleneedianiline led to an outbreak of 84 cases of jaundice in Eppin, England, in the early 1960s (Kopelman et al. 1966). Six young people developed acute jaundice and may have suffered severe liver damage when they ingested an
alcoholic beverage spiked with 4,4´-methyleneedianiline, which they assumed was the psychometric
drug methylenedioxyamphetamine; both have the same abbreviation, MDA (Tillmann et al. 1997).
Consumer products that contain 4,4´-methyleneedianiline are polyurethane foam, Spandex® fiber, and
epoxy-containing products (see Section 4.3). Under normal use conditions, very little 4,4´-
methyleneedianiline is present in the free state in these consumer products. Potential consumer exposure
may occur via dermal contact with trace amounts of free 4,4´-methyleneedianiline found in automobile
cushioning or epoxy-containing products (NIEHS 1994). It has been reported that the levels of the
compound in food, food additives, and food packaging are so low that the potential daily intake via
food is virtually zero (NIEHS 1994). With the exception of certain medical devices, there is no
documentation of exposure to 4,4´-methyleneedianiline from any consumer products. Polyurethane is
widely used in such medical devices as potting materials for components of plasma separators and
artificial dialyzers (Shintani 1991). Although polyurethane in these materials contains methylene
diphenyldiisocyanate, polyurethane releases free 4,4´-methyleneedianiline during sterilization by gamma
irradiation or autoclaving (Shintani 1991). Therefore, there is a potential risk of exposure to
4,4´-methyleneedianiline for patients with kidney disease or patients who receive frequent blood
transfusions (Shintani 1991). The relative potential of sterilization methods to release 4,4´-
methyleneedianiline and mutagenic compounds from polyethylene foam potting material has been studied
(Shintani 1995a). Gamma-ray irradiation sterilization at 2.5 Mrad produced a few ppm of the
compound, but more importantly, produced unknown compounds which proved to be mutagenic in the
absence of metabolic activity. On the other hand, no formation of 4,4´-methyleneedianiline was
observed in autoclaved thermosetting polyurethane potting material (heated at 121°C for 60 minutes)
and smaller amounts of mutagenic compounds were determined. Thus, autoclave sterilization is more
appropriate, provided materials can withstand the process. Another sterilization technique, ethylene
oxide gas, produced the least amount of 4,4´-methyleneedianiline and other compounds, but the residue
of ethylene oxide gas is itself problematic (Shintani 1995b).

Occupational exposure to 4,4´-methyleneedianiline can occur during its production; during its use in the
manufacture of insulation materials (e.g., plastic insulating materials and polyurethane), coremaking in
iron and steel foundries, and encapsulation of instruments with polyurethane mixture; and during use
of epoxy resins in the construction of nuclear power plants (NIOSH 1976). The major routes for
occupational exposure to 4,4´-methyleneedianiline are dermal and inhalation (Cocker et al. 1994;
Peterson et al. 1991). The occupational exposure to 4,4´-methyleneedianiline can be assessed both by
air monitoring and biological monitoring of urinary 4,4´-methyleneedianiline and its metabolites in
workers. However, air monitoring alone would underestimate occupational exposure to the compound by failing to consider exposure from skin absorption (Peterson et al. 1991). There is some evidence which showed that when exposure to 4,4´-methylenedianiline was via inhalation, postshift urine samples of workers had higher concentrations of the compound than samples taken preshift next day (Cocker et al. 1994). When the exposure was mostly via the dermal route, urine samples taken preshift the next day had higher concentrations of 4,4´-methylenedianiline than urine samples collected immediately postshift (Cocker et al. 1994). Therefore, urine collection strategies may provide some clue about the route of entry into the body. Workers’ exposure to 4,4´-methylenedianiline used in an epoxy mixture to insulate electrical cables was monitored by analysis of the compound in blood and urine (Dalene 1995). At the end of the workday, workers removed material from their skin with a cleaning solution containing mainly butyl acetate. Some results suggested that using such cleaning solutions may enhance the absorption of 4,4´-methylenedianiline (compared to washing with soap and water).

Occupational exposure to 4,4´-methylenedianiline has been assessed by monitoring workplace air, analysis of the substance collected on dermal pads, and analysis of urine. The concentrations of 4,4´-methylenedianiline in the air of working areas for some of these industries are given in Table 5-2. A concentration of \( \leq 29 \, \text{mg/m}^3 \) was detected in the occupational air of a facility where workers were blending and bagging a hardener containing 4,4´-methylenedianiline (Shuker et al. 1986). In a survey conducted in Britain, postshift urine samples were taken from 111 workers at 5 factories that used 4,4´-methylenedianiline (Cocker et al. 1986a). The urinary concentrations (both free and conjugated 4,4´-methylenedianiline) in 77.2% of the samples were below the detection limit (<5 nmol 4,4´-methylenedianiline/ mmol creatinine) and the concentration range was 6- 175 nmol 4,4´-methylenedianiline/ mmol creatinine in the rest of the samples. In a more recent survey (1989-1990) by the same group, 960 preshift and postshift urine samples from 411 workers from 45 factories that manufactured or used 4,4´-methylenedianiline were analyzed for total 4,4´-methylenedianiline (Cocker et al. 1994). The compound was not detected in 57% of postshift and 42% of the preshift urine samples. The concentrations of 4,4´-methylenedianiline in about 95% of urine samples were below 100 nmol/mmol creatinine. However, the concentration of 4,4´-methylenedianiline in 3 postshift urine samples exceeded 1,300 nmol/mmol creatinine and the concentration in one sample was 687 1 nmol/mmol creatinine. The maximum urinary 4,4´-methylenedianiline levels were found among manufacturers and formulators. In a Swedish study of 8 epoxy resin workers, the urinary total 4,4´-methylenedianiline concentrations (not standardized with creatinine level in urine) were in the
Table 5-2. Concentrations of 4,4'-Methylenedianiline in Air/Dermal Pad in Some Working Areas

<table>
<thead>
<tr>
<th>Industry</th>
<th>Sample type</th>
<th>Concentration range (µg/m³)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>4,4'-Methylenedianiline</td>
<td>Personal area</td>
<td>5–74</td>
<td>Boeniger 1984a</td>
</tr>
<tr>
<td>production</td>
<td>Dermal pad</td>
<td>13–651</td>
<td>Boeniger 1984a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>4.2–54&lt;sup&gt;a&lt;/sup&gt;</td>
<td>Boeniger 1984a</td>
</tr>
<tr>
<td>Plastic insulating materials</td>
<td>Personal area</td>
<td>1–690</td>
<td>IARC 1986</td>
</tr>
<tr>
<td></td>
<td></td>
<td>&lt;0.3–99</td>
<td>IARC 1986</td>
</tr>
<tr>
<td>Coremaking in iron and steel</td>
<td>Area</td>
<td>Up to 1,600</td>
<td>Toeniskoetter 1981</td>
</tr>
<tr>
<td>foundries</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Coremaking in aluminum</td>
<td>Area</td>
<td>&gt;1,600</td>
<td>Toeniskoetter 1981</td>
</tr>
<tr>
<td>foundries</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Encapsulation of instruments</td>
<td>Area</td>
<td>10</td>
<td>IARC 1986</td>
</tr>
<tr>
<td>with polyurethane</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<sup>a</sup>This value is in µg/cm³
range of 10-3,026 nmol/L (2-600 µg/L) (Tiljander et al. 1989). 4,4´-Methylenedianiline was detected (detection limit < 1.7 nmol/mmol creatinine) in <20% of urine samples from more than 160 workers at 10 industrial sites in the United States at a concentration range of 1.7-119.8 nmol 4,4´-methyleneedianiline/ mmol creatinine (2.9-210 µg/g creatinine) (Peterson et al. 1991). The National Occupational Exposure Survey (NOES) conducted by NIOSH during 1981-83 estimated that a total of 15,178 workers in different industries are potentially exposed to 4,4´-methylenedianiline (NIOSH 1989). However, the NOES database does not contain data on the frequency, duration, concentration, or route of exposure of workers to 4,4´-methylenedianiline (or any other chemical). German researchers (Sepai 1995) found that a hemoglobin adduct of an acetylated metabolite of 4,4´-methylenedianiline can be detected after occupational exposure to MDI. The authors consider this evidence for the biological availability of 4,4´-methylenedianiline from in vivo hydrolysis of MDI.

5.6 POPULATIONS WITH POTENTIALLY HIGH EXPOSURES

Within the general population, there are a few groups that are potentially exposed to 4,4´-methylenedianiline at higher levels than the background population. In addition to individuals exposed to the compound in the workplace, these groups include: patients with kidney disease, patients who receive frequent blood transfusions (Shintani 1991), and people who live in the vicinity of 4,4´-methylenedianiline disposal facilities. With the exception of occupational groups, the levels of exposure in other potentially high exposure groups have not been documented either by measuring levels in the contaminated media (e.g., air, transfused plasma) or the levels in tissues or body fluids of the exposed persons.

5.7 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of 4,4´-methylenedianiline is available. Where adequate information is not available, ATSDR, in conjunction with the NTP, is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 4,4´-methylenedianiline.
The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

5.7.1 Identification of Data Needs

**Physical and Chemical Properties.** Data on physical and chemical properties are available for 4,4′-methyleneedianiline (HSDB 1996; IARC 1986; Moore 1978), although some of these data (vapor pressure, \( p_b \), and Henry’s law constant) are estimated by reliable methods. The available data will permit the prediction of environmental fate processes for 4,4′-methyleneedianiline. Therefore, a data need has not been identified.

**Production, Import/Export, Use, Release, and Disposal.** Five companies that manufacture 4,4′-methyleneedianiline for sale and/or distribution also utilize the predominant amount of manufactured material captively for the production of other products (IARC 1986; TR194 1996). Therefore, data on recent production volumes or the trend in yearly production volume for 4,4′-methyleneedianiline remain unavailable. The availability of this data could be important, because it provides indirect evidence of environmental release. Information regarding the recent trend in import/export volumes and its use is well documented in the literature (IARC 1986; NTDB 1994). Since medical devices such as plasma separators or artificial dialyzers are often fabricated with polyurethane as a potting material, they release 4,4′-methyleneedianiline during sterilization (Shintani 1991). This is one of the documented sources of consumer exposure to the compound. Although some information regarding the method of disposal is available (HSDB 1996), it would be helpful to obtain more information on the methods currently used by industries for the disposal of 4,4′-methyleneedianiline wastes that are transferred off-site. 4,4′-Methyleneedianiline itself is used to prepare other chemicals or products; it is not used in the home or the general environment and thus, would not contaminate any home or environmental media. No literature data were found on 4,4′-methyleneedianiline as a food contaminant. Levels of the compound in food, food additives, and food packaging are so low that potential daily intake via these routes is virtually zero (NIEHS 1994). No residue of the compound was found on vegetables grown on soilless reconstituted polyurethane foam with or without nutrient solutions (Rouchaud et al. 1992). Consumer products that contain
4,4’-methyleneedianiline are polyurethane foam, Spandex® fiber, and epoxy-containing products (see Section 4.3). Under normal use conditions, very little 4,4’-methyleneedianiline is present in the free state in such products.

According to the Emergency Planning and Community Right-to-Know Act of 1986, 42 U.S.C. Section 11023, industries are required to submit chemical release and off-site transfer information to the EPA. The Toxics Release Inventory (TRI), which contains this information for 1992, became available in May of 1994. This database will be updated yearly and should provide a list of industrial production facilities and emissions.

**Environmental Fate.** Experimental data on the fate of 4,4’-methyleneedianiline in air, water, and soil are scarce. Homogeneous reaction of vapor-phase 4,4’-methyleneedianiline with the hydroxyl radical in the atmosphere is known to be an important removal mechanism (half-life estimated at 1.6 hours). However, 4,4’-methyleneedianiline exists in the atmosphere predominantly as either a solid aerosol or adsorbed on other particles. Very little is known about the reaction mechanisms, rate of removal, or degradation products formed when 4,4’-methyleneedianiline reacts heterogeneously with hydroxyl radical or gas-phase nitric acid. Based on its low estimated vapor pressure of 2.15×10^{-7} mm Hg at 25 °C, and its ability of form covalent bonds with humic substances present in soil and sediment (Parris 1980), 4,4’-methyleneedianiline will partition into sediment of surface water and in soil. The strong binding of the compound with humic substances in soil will retard its mobility, resulting in low leachability in most soils. However, it would be important to verify experimentally the sorption behavior of 4,4’-methyleneedianiline in sediment and soil. The half-lives of 4,4’-methyleneedianiline due to biodegradation in aerobic soil and anaerobic sediment/soil are predicted to be 1-7 days and 4-28 days, respectively (Howard et al. 1991). Therefore, the compound will persist in deeper soil and sediment where there is a lack of oxygen. However, these estimated halflives and the prediction that biodegradation will be the most important fate of 4,4’-methyleneedianiline in water and soil need further experimental verification.

**Bioavailability from Environmental Media.** People who live in the vicinity of disposal facilities may be exposed to 4,4’-methyleneedianiline by inhalation of airborne dust, ingestion of soil (children), and dermal contact with soil (children). However, no monitoring data on levels of the compound in air, water, or sediment and soil were located in the literature search. Analyses of fish and vegetables have been conducted, but no residues were found at the detection limits attained (see
Section 5.4). Thus, no information on the absorption of 4,4′-methyleneedianiline from contaminated air, water, soil, or plant material is available. In order to estimate the uptake of the substance as a result of exposure, it is important that the bioavailability of 4,4′-methyleneedianiline from each route of entry be known. The bioavailability may not depend entirely on the sorption coefficient of 4,4′-methyleneedianiline on soil or air particles, but may also depend on the pH of the media of contact. Because 4,4′-methyleneedianiline is a weak base, its bioavailability may increase as the medium becomes more and more acidic. Since no data on the bioavailability of 4,4′-methyleneedianiline from contaminated air and soil are available, it would be helpful to develop this information.

Reliable monitoring data for the levels of 4,4′-methyleneedianiline in contaminated media at hazardous waste sites are needed so that the information obtained on levels of 4,4′-methyleneedianiline in the environment can be used in combination with the known body burdens of 4,4′-methyleneedianiline to assess the potential risk of adverse health effects in populations living in the vicinity of hazardous waste sites.

**Food Chain Bioaccumulation.** Both the experimentally determined $K_w$ value (HSDB 1996) and the estimated bioconcentration value indicate that 4,4′-methyleneedianiline will not bioconcentrate in lipids and will not bioconcentrate in aquatic organisms or animals. Experimental study also supports this conclusion (III 1981). No data located in the literature suggest that 4,4′-methyleneedianiline will biomagnify in animals of higher trophic levels through aquatic or terrestrial food chains. Since experimental data on the bioaccumulation and biomagnification potential of 4,4′-methyleneedianiline are lacking, it would be helpful to develop these data in both aquatic and terrestrial food chains.

**Exposure Levels in Environmental Media.** Other than in occupational air, measured values for the levels of 4,4′-methyleneedianiline in ambient air, water, soil, plant materials, and foodstuffs have not been reported. Consequently, no estimate of human intake of the compound from various environmental media was located in the literature. Reliable monitoring data for the levels of 4,4′-methyleneedianiline in contaminated media at hazardous waste sites are needed so that the information obtained on levels in the environment can be used in combination with the known body burden of the substance to assess the potential risk of adverse health effects in populations living in the vicinity of hazardous waste sites.
Reliable monitoring data for the levels of methylenedianiline in contaminated media at hazardous waste sites are needed so that the information obtained on levels of 1,4-dichlorobenzene in the environment can be used in combination with the known body burdens of 1,4-dichlorobenzene to assess the potential risk of adverse health effects in populations living in the vicinity of hazardous waste sites.

**Exposure Levels in Humans.** Data on the levels of 4,4´-methylenedianiline in urine of occupationally exposed workers are available (Cocker et al. 1994; Peterson et al. 1991; Tiljander et al. 1989). No data were located on the levels in other tissues or body fluids of exposed or unexposed people. Similarly, no biological monitoring study has been conducted for populations in the vicinity of hazardous waste sites. This information is necessary for assessing the need to conduct health studies on these populations.

**Exposure Registries.** No exposure registries for 4,4´-methylenedianiline were located. This substance is not currently one of the compounds for which subregistry has been established in the National Exposure Registry. The substance will be considered in the future when chemical selection is made for subregistries to be established. The information that is amassed in the National Exposure Registry facilitates the epidemiological research needed to assess adverse health outcomes that may be related to exposure to this substance.

**5.7.2 Ongoing Studies**

A search of the Federal Research in Progress (FEDRIP) database did not identify any ongoing studies that could be useful in filling the data gaps discussed in Section 5.7.1.
6. ANALYTICAL METHODS

The purpose of this chapter is to describe the analytical methods that are available for detecting, and/or measuring, and/or monitoring 4,4’-methylenedianiline, its metabolites, and other biomarkers of exposure and effect to 4,4’-methylenedianiline. The intent is not to provide an exhaustive list of analytical methods. Rather, the intention is to identify well-established methods that are used as the standard methods of analysis. Many of the analytical methods used for environmental samples are the methods approved by federal agencies and organizations such as EPA and the National Institute for Occupational Safety and Health (NIOSH). Other methods presented in this chapter are those that are approved by groups such as the Association of Official Analytical Chemists (AOAC) and the American Public Health Association (APHA). Additionally, analytical methods are included that modify previously used methods to obtain lower detection limits, and/or to improve accuracy and precision.

6.1 BIOLOGICAL SAMPLES

Methods are available for the determination of 4,4’-methylenedianiline in blood and urine. Representative methods are summarized in Table 6-1. Most methods involve an extraction step, purification and fractionation procedures, and analysis, usually by gas chromatography (GC) or high performance liquid chromatography (HPLC).

The determination of 4,4’-methylenedianiline in blood, serum, or urine requires special precautions because the compound and some of its metabolites are heat labile (Cocker et al. 1988; Shintani 1992). During sample treatment, care should be taken to assure that no loss of 4,4’-methylenedianiline or its metabolites has occurred during evaporative steps used for the concentration in organic solutions and during hydrolysis of urine (Cocker et al. 1988; Shintani 1992).

Determination of 4,4’-methylenedianiline in blood/serum requires pretreatment steps involving deproteinization with a suitable reagent (e.g., perchloric acid), centrifugation for separation of 4,4’-methylenedianiline from the precipitate, and concentration (Shintani 1992). Determination of 4,4’-methylenedianiline in urine often uses solvent extraction and multiple clean-up steps (Avery 1989). An improved method uses solid phase extraction columns (octadecyl bonded silica) for the
<table>
<thead>
<tr>
<th>Sample matrix</th>
<th>Preparation method</th>
<th>Analytical method</th>
<th>Sample detection limit</th>
<th>Percent recovery</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood</td>
<td>Alkaline hydrolysis; solvent extraction; clean-up by acid-base extraction; re-extraction of basic sample in n-heptane-isooamylic alcohol; evaporation to dryness.</td>
<td>GC/ECD</td>
<td>10 ng/mL</td>
<td>72</td>
<td>Tortoreto et al. 1983</td>
</tr>
<tr>
<td>Serum</td>
<td>Solid-phase extraction (reverse phase C_{18}, phenyl, cyclohexyl columns)</td>
<td>HPLC/EICD</td>
<td>3 ng/mL</td>
<td>100</td>
<td>Shintani 1991, 1992</td>
</tr>
<tr>
<td>Serum and urine</td>
<td>Solid-phase extraction (C_{18}-bonded silica column); derivatization with PFPA.</td>
<td>Capillary GC/MS-NICI</td>
<td>0.1 ng/mL (water)</td>
<td>105</td>
<td>Avery 1989</td>
</tr>
<tr>
<td>Urine (MDA and N-acetyl and N-glucuronide metabolites)</td>
<td>Hydrolysis (10 M NaOH) at 80°C for 90 minutes; extraction of cooled sample with diethyl ether; concentration and derivatization with PFBA.</td>
<td>Capillary GC/MS</td>
<td>5 ng/mL</td>
<td>&gt;80</td>
<td>Cocker et al. 1988</td>
</tr>
<tr>
<td>Urine (total: free, acetylated and conjugated)</td>
<td>Hydrolysis; extraction on C_{18} solid phase columns; concentration.</td>
<td>HPLC/EICD; confirmation by GC/MS</td>
<td>2.5 ng/mL</td>
<td>68</td>
<td>Peterson et al. 1991</td>
</tr>
<tr>
<td>Urine (total: free, acetylated and conjugated)</td>
<td>Acid hydrolysis; extraction with toluene; derivatization with PFPA.</td>
<td>Capillary GC/MS-SIM</td>
<td>2 ng/mL</td>
<td>96</td>
<td>Tiljander et al. 1990</td>
</tr>
<tr>
<td>Urine (total: MDA, N-acetyl-MDA, N,N'-diacetyl-MDA)</td>
<td>Acid hydrolysis; extraction with toluene; derivatization with PFPA.</td>
<td>HPLC/UV (285 and 258 nm)</td>
<td>8 ng/mL</td>
<td>97</td>
<td>Tiljander and Skarping 1990</td>
</tr>
<tr>
<td>Urine (MDA and conjugates)</td>
<td>Alkaline hydrolysis; extraction with toluene; optional cleanup; derivatization with PFPA.</td>
<td>Micro LC/UV (258 nm)</td>
<td>0.4 ng/mL</td>
<td>89</td>
<td>Brunmark et al. 1992</td>
</tr>
<tr>
<td>Azo-, azoxy- and nitroso metabolites of MDA in microsomal incubation media</td>
<td>Protein precipitation followed by solid-phase column extraction; evaporation to dryness.</td>
<td>HPLC/PSMS; off-line HPLC tandem FAB/MS</td>
<td>No data</td>
<td>No data</td>
<td>Kajbaf et al. 1992</td>
</tr>
</tbody>
</table>

EICD = electrochemical detector; FAB = fast atom bombardment; GC = gas chromatography; ECD = electron capture detector; Hb = hemoglobin; HFBC = heptafluorobutryl chloride; HPLC = high performance liquid chromatography; LC = liquid chromatography; MDA = 4,4'-methylenedianiline; MS = mass spectrometry; NaOH = sodium hydroxide; NICI = negative ion chemical ionization; PFPA = pentafluoropropionic anhydride; PSMS = plasma spray mass spectrometry; SIM = selected ion monitoring; UV = ultraviolet detection.
separation of 4,4´-methyleneedianiline from interfering components in blood and hydrolyzed urine (Peterson et al. 1991; Shintani 1992).

Due to the polar nature of 4,4´-methyleneedianiline, derivatization of the amine groups is required to produce a species which can be analyzed by GC without peak tailing. This derivatization may not be necessary if HPLC is used (Peterson et al. 1991). The common derivatizing agents are trifluoroacetic anhydride, pentafluoropropionic anhydride (PFPA), and heptafluorobutyric anhydride (HFBA).

The commonly used HPLC techniques include ultraviolet absorbance detection (UV) (Brunmark et al. 1992; Tiljander et al. 1990; Tiljander and Skarping 1990) and electrochemical detection (Peterson et al. 1991; Shintani 1992). Generally, the detection limit of 4,4´-methyleneedianiline with electrochemical detection is in the range of 2-3 ng/mL (Peterson et al. 1991; Shintani 1992). This method is approximately two orders of magnitude better than the HPLC/UV method, which has a detection limit of 150 ng/mL (Shintani 1992). However, with special techniques, such as precolumn separation of derivatized 4,4´-methyleneedianiline and microliquid chromatography, it is possible to lower the detection limits with UV in the range 0.4 ng/mL-8.0 ng/mL (Brunmark et al. 1992; Tiljander and Skarping 1990; Tiljander et al. 1990). Interferences from biological materials may co-elute with 4,4´-methyleneedianiline, so confirmation is recommended. A second wavelength has been used with UV detection (Tiljander and Skarping 1990) and GC/mass spectrometry (Peterson et al. 1991).

GC may be used with electron capture detection (ECD) (Tortoreto et al. 1983), thermionic specific detection (Skarping et al. 1983), and mass spectrometry (MS) (Avery 1989; Cocker et al. 1988). Detection limits in the range of l-10 ng/mL are attainable by GC/MS (Avery 1989; Cocker et al. 1986a, 1988). For urine and serum, a detection limit of ≤1 ng/mL was obtained with solid phase extraction, derivatization, and GC/negative-ion chemical ionization MS (Avery 1989). The detection limit for GC/ECD is l-10 ng/mL (Skarping et al. 1983; Tortoreto et al. 1983). Although the detection limit with the thermionic specific detectors is about 10 times higher than electron capture detection (Skarping et al. 1983), both thermionic specific detector and MS detectors are more selective for 4,4´-methyleneedianiline determination (Cocker et al. 1986a; Skarping et al. 1983).

Besides the parent 4,4´-methyleneedianiline, N-acetyl-, N,N´-diacetyl-, and the N-glucuronide of 4,4´-methyleneedianiline may also be present in urine as metabolites (Cocker et al. 1986a, 1988;
Peterson et al. 1991). N-acetyl-4,4'-methylenedianiline has been identified in human urine (Cocker 1986a). Three metabolites have been identified in rabbit liver microsomal incubations: azodiphenylmethane (azo), azoxydiphenylmethane (azoxy), and 4-nitroso-4'-aminodiphenylmethane (nitroso) compounds (Kajbaf et al. 1992). The formation of metabolites is shown schematically in Figure 2-3. Methods that can be used for the determination of metabolites are included in Table 6-1.

It is possible to determine the free amine (4,4'-methylenedianiline) and the total 4,4'-methylenedianiline (free, acetylated, and conjugated) in urine. The treatment of urine with strong base or strong acid hydrolyzes acetylated and conjugated 4,4'-methylenedianiline to the free 4,4'-methylenedianiline (Cocker et al. 1988; Tiljander and Skarping 1990). Therefore, the determination of free and total 4,4'-methylenedianiline is possible by determining the amount of 4,4'-methylenedianiline in unhydrolyzed and hydrolyzed urine (Peterson et al. 1991). Methods for the determination of free and total 4,4'-methylenedianiline are included in Table 6-1.

Biological monitoring for 4,4'-methylenedianiline and its metabolites is used to assess exposure to 4,4'-methylenedianiline. Available methods are summarized in Table 6-2. The methods are similar to the methods described in Table 6-1, except that very sensitive and specific MS techniques are used for analysis. In addition, the monitoring strategy must take into account the route of exposure (Cocker et al. 1994). The corresponding hemoglobin adducts have also been measured. Adducts are analyzed by GC/MS after hydrolysis, extraction and derivatization of hemoglobin precipitated from lysated red blood cells (Bailey et al. 1990; Schutz et al. 1995). Very low detection limits have been reported; however, little other performance information is available (Bailey et al. 1990; Schutz et al. 1995).

6.2 ENVIRONMENTAL SAMPLES

Analytical methods available for the determination of 4,4'-methylenedianiline in environmental samples are listed in Table 6-3. As in the case of biological samples, 4,4'-methylenedianiline in environmental samples has been determined by HPLC/UV (Emes and Hanshumaker 1983; Gunderson and Anderson 1988; Mazzu and Smith 1984), HPLC/electrochemical detection (Concialini et al. 1983), and GC/thermionic detection (Audunsson and Mathiasson 1983). Although analysis of 4,4'-methylenedianiline by HPLC does not require derivatization, analysis of environmental samples is frequently performed after derivatization to reduce amine reactivity on HPLC column packing.
Table 6.2 Analytical Methods for Determining 4,4'-Methylenedianiline in Biological Materials

<table>
<thead>
<tr>
<th>Sample matrix</th>
<th>Sample preparation</th>
<th>Analytical method</th>
<th>Sample detection limit</th>
<th>Accuracy % recovery</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma and urine (MDA)</td>
<td>Alkaline hydrolysis; extraction with toluene; PFPA derivatization.</td>
<td>Capillary GC/MS-NICI</td>
<td>0.2 nmol/L</td>
<td>97 (urine) 96 (plasma)</td>
<td>Brunmark et al. 1995</td>
</tr>
<tr>
<td>Urine (AcMDA)</td>
<td>Hydrolysis at pH 13, 80°C; solvent extraction; HFBA derivitization.</td>
<td>Capillary GC/MS</td>
<td>No data</td>
<td>No data</td>
<td>Cocker et al. 1986a</td>
</tr>
<tr>
<td>Blood plasma and urine (MDA and AcMDA)</td>
<td>Alkaline hydrolysis; PFPA derivitization</td>
<td>Capillary GC/MS-NICI</td>
<td>0.2 nmol/L</td>
<td>97% (urine) 96% (plasma)</td>
<td>Brunmark et al. 1995</td>
</tr>
<tr>
<td>Blood and urine (MDA and conjugates)</td>
<td>Acid hydrolysis; solvent extraction; PFPA derivatization</td>
<td>capillary GC/MS-NICI</td>
<td>3 nmol/L</td>
<td>Not reported</td>
<td>Dalene et al. 1995</td>
</tr>
<tr>
<td>Serum and urine (MDA)</td>
<td>Acid hydrolysis; solvent extraction; PFPA derivatization</td>
<td>capillary GC/MS-NICI-SIM</td>
<td>0.05 μg/L</td>
<td>Not reported</td>
<td>Skarping et al. 1995</td>
</tr>
<tr>
<td>Blood (hemoglobin-MDA and -AcMDA adducts)</td>
<td>Precipitation of hemoglobin from lysated red blood; alkaline hydrolysis; extraction with ethyl acetate; PFPA derivatization.</td>
<td>Capillary GC/MS-SIM</td>
<td>&lt;10 pmol/g Hb</td>
<td>&gt;80</td>
<td>Bailey et al. 1990</td>
</tr>
<tr>
<td>Blood (hemoglobin-MDA and -AcMDA adducts)</td>
<td>Centrifugation; precipitation of hemoglobin from lysated red blood cells; SPE or solvent extraction; HFBA derivatization</td>
<td>capillary GC/MS-NICI-SIM</td>
<td>&lt;20 fmol/sample (MDA); 100 fmol/sample (AcMDA)</td>
<td>Not reported</td>
<td>Schutz et al. 1995</td>
</tr>
<tr>
<td>Plasma and urine (MDA and isomers 2,4'-MDA and 2,2'-MDA and methylated MDA)</td>
<td>Acid hydrolysis; solvent extraction; PFPA derivatization</td>
<td>capillary GC/MS-NICI</td>
<td>&lt;10 ng/L (instrumental)</td>
<td>Not reported</td>
<td>Skarping and Dalene 1995</td>
</tr>
</tbody>
</table>

AcMDA = N-acetyl-MDA; GC = Gas Chromatography; Hb = hemoglobin; HFBC = heptafluorobutryl chloride; HPLC = high performance liquid chromatography; MDA = 4,4'-methylenedianiline; MS = mass spectrometry; NICI = negative ion chemical ionization; PFPA = pentafluoropropanoic anhydride; SIM = selected ion monitoring; UV = ultraviolet (detection)
<table>
<thead>
<tr>
<th>Sample matrix</th>
<th>Preparation method</th>
<th>Analytical method</th>
<th>Sample detection limit</th>
<th>Percent recovery</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Occupational air</td>
<td>Collection on acid-coated glass-fiber filter; extraction with 0.26 N sodium hydroxide/5% acetonitrile; derivatization with acetic anhydride.</td>
<td>HPLC/UV</td>
<td>&lt;1.0 µg/m³</td>
<td>&gt;90</td>
<td>Gunderson and Anderson 1988</td>
</tr>
<tr>
<td>Occupational air</td>
<td>Collection in impingers containing 0.05 M sulfuric acid; addition of sodium hydroxide pellets to make basic; extraction with toluene.</td>
<td>GC/TSD</td>
<td>2 µg/m³</td>
<td>&gt;80</td>
<td>Audunsson and Mathiasson 1983</td>
</tr>
<tr>
<td>Occupational air (NIOSH Method 5029)</td>
<td>Collection on acid-coated glass-fiber filter; ultrasonic extraction with 0.1 N methanolic potassium hydroxide.</td>
<td>HPLC/UV and EICD</td>
<td>0.2 µg/m³ for 100 L sample</td>
<td>&gt;80</td>
<td>NIOSH 1994</td>
</tr>
<tr>
<td>Solution of neat compound</td>
<td>Acidification; derivatization to diazo-compound with N-(1-naphthyl)-ethylenediamine.</td>
<td>Spectrophoto-metric at 555 nm</td>
<td>No data</td>
<td>95–105</td>
<td>Norowitz and Kelber 1986</td>
</tr>
<tr>
<td>Polyurethane extract (autoclaved and soaked in water)</td>
<td>Extraction with diethyl ether; derivatization with triethylamine and benzoyl chloride; concentration.</td>
<td>HPLC/UV</td>
<td>&lt;70 ng/g in polyurethane and &lt;3.5 µg/L for extract</td>
<td>&gt;60</td>
<td>Mazzu and Smith 1984</td>
</tr>
<tr>
<td>Aqueous extract of polyurethane film</td>
<td>Extraction with diethyl ether; derivatization with triethylamine and benzoyl chloride; concentration.</td>
<td>HPLC/UV (280 and 254 nm)</td>
<td>0.05 µg/L for extract</td>
<td>&gt;89</td>
<td>Ernes and Hanshumaker 1983</td>
</tr>
<tr>
<td>Water</td>
<td>Extraction of sample spiked with tetradeterated analogue from C₁₈ cartridge; derivatization with PFPA.</td>
<td>Capillary GC/NICI-MS and EI/MS</td>
<td>2 ng/L</td>
<td>No data</td>
<td>Benfenati et al. 1992</td>
</tr>
</tbody>
</table>

EI/MS = electron impact mass spectrometry; GC = gas chromatography; EICD = electrochemical detector; HPLC = high performance liquid chromatography; MS = mass spectrometry; NICI = negative ion chemical ionization; PFPA = pentafluoropropion anhydride; TSD = thermionic specific detection; UV = ultraviolet detection
materials (Emes and Hanshumaker 1983; Gunderson and Anderson 1988; Mazzu and Smith 1984). Because of lower matrix interference environmental samples (air or water), the detection limits can be an order of magnitude lower than those achieved for biological samples (Avery 1989). The available analytical methods are capable of detecting 4,4´-methyleneedianiline in air at concentrations lower than the ACGIH threshold limit value of 0.08 mg/m³ (10 ppb) (for time-weighted average for 8 hours) and a short-term exposure limit of 0.8 mg/m³ (100 ppb) (Audunsson and Mathiasson 1983; Boeniger 1984a; Gunderson and Anderson 1988).

Special precautions should be taken during sampling of workplace air for 4,4´-methyleneedianiline. The compound can be present in workplace air, both in the vapor and particle phase. Therefore, air sampling methods must be able to collect 4,4´-methyleneedianiline present in both phases. Glass fiber filter/silica gel sampling tubes (Boeniger 1984a) or acid-impregnated glass fiber filters (Gunderson and Anderson 1988) are capable of collecting 4,4´-methyleneedianiline in both phases. In certain workplace environments, such as polyurethane industries that produce methylenediphenyl diisocyanate (MDI) or polymeric MDI, the air may contain both 4,4´-methyleneedianiline and MDI. Sampling methods that use impingers containing acids (Audunsson and Mathiasson 1983) or acid-coated glass fiber filters (Gunderson and Anderson 1988) would hydrolyze MDI to 4,4´-methyleneedianiline (Audunsson and Mathiasson 1983; Gunderson and Anderson 1988). Therefore, if a distinction is to be made between 4,4´-methyleneedianiline and MDI, either MDI or 4,4´-methyleneedianiline should be determined in the sample alone and could be subtracted from the total (following acid hydrolysis) for the determination of individual MDI and 4,4´-methyleneedianiline concentrations. Methods for such determinations are available (Audunsson and Mathiasson 1983; Boeniger 1984a).

6.3 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of 4,4´-methyleneedianiline is available. Where adequate information is not available, ATSDR, in conjunction with the NTP, is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 4,4´-methyleneedianiline.
The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

6.3.1 Identification of Data Needs

Methods for Determining Biomarkers of Exposure and Effect. The total amount of 4,4′-methylenedianiline (free and conjugated) in urine is used as an indicator of 4,4′-methylenedianiline exposure in workers (Bnmmark et al. 1995; Cocker et al. 1994). Total MDA in plasma is also an appropriate method for biological monitoring of 4,4′-methylenedianiline. Methods for the determination of total 4,4′-methylenedianiline in human urine and blood by alkaline hydrolysis of urine are available and shown in Table 6-1. However, no quantitative correlation has been established between level of workplace exposure and urinary level of free and/or conjugated 4,4′-methylenedianiline. Adducts of 4,4′-methylenedianiline with hemoglobin can also be used as an indicator of exposure (Bailey et al. 1990; Schutz et al. 1995). These methods are suitable for the monitoring of occupational exposure. It would be helpful to assess whether the existing analytical methods are capable of detecting the compound in biological samples at levels at which biological effects might occur.

There is no specific biomarker of effect that can be attributed to 4,4′-methylenedianiline exposure. Exposure to the compound is usually associated with occurrence of jaundice, bile duct inflammation, suppression of bile excretion, and clinical hepatitis (NIOSH 1986). However, none of these effects is specific to 4,4′-methylenedianiline exposure. It would be useful to establish that a certain effect could be specifically attributed to 4,4′-methylenedianiline exposure.

Methods for Determining Parent Compounds and Degradation Products in Environmental Media. Few methods are available for analysis of 4,4′-methylenedianiline in environmental media. Methods are available for determination in occupational air (Audunsson and Mathiasson 1983; Boeniger 1984a; Gunderson and Anderson 1988). Analytical methods of sufficient sensitivity, precision, and accuracy are available for the determination of 4,4′-methylenedianiline in occupational air at levels at least an order of magnitude lower than the ACGIH level of 0.8 mg/m³.
(Hoeniger 1984a; Gunderson and Anderson 1988). However, the compound has not been measured in ambient air. Methods are also available for the determination of extractable 4,4´-methylenedianiline in polyurethanes (Emes and Hanshumaker 1983; Mazzu and Smith 1984). Standardized analytical methods for the determination of 4,4´-methylenedianiline in water, soil, and other environmental media are not available. Therefore, it would be most useful to develop standardized analytical methods of sufficient sensitivity, precision, and accuracy for its determination, at least, in contaminated environmental media. Most important, analytical methods should be developed to determine the concentrations of 4,4´-methylenedianiline in soil and groundwater at Superfund waste disposal sites.

Experimental data on the environmental fate of 4,4´-methylenedianiline are very scarce. Consequently, the environmental degradation products of the compound have not been identified. Therefore, before the evaluation of this data need, it would be more important to conduct more studies to assess the environmental fate of 4,4´-methylenedianiline.

6.3.2 Ongoing Studies

No ongoing studies involving methods for determination of 4,4´-methylenedianiline were located.
7. REGULATIONS AND ADVISORIES

The international, national, and state regulations and guidelines regarding 4,4´-methylenedianiline in air, water, and other media are summarized in Table 7-1.

ATSDR has derived an acute oral MRL of 0.2 mg/kg/day based on a minimal LOAEL for liver effects observed in rats treated with a single dose of 4,4´-methylenedianiline (Bailie et al. 1993).

ATSDR has derived an intermediate oral MRL of 0.08 mg/kg/day based on a NOAEL for liver effects observed in rats treated daily with gavage doses of 4,4´-methylenedianiline for 12 weeks (Pludro et al. 1969).

Neither a reference dose nor a reference concentration for 4,4´-methylenedianiline is available at this time.

The International Agency for Research on Cancer (IARC) has classified 4,4´-methylenedianiline as 2B, probably carcinogenic to humans, based on limited evidence of carcinogenicity in humans but sufficient evidence of carcinogenicity in animals (IARC 1987). The EPA and the National Toxicology Program (NTP) have not classified the chemical for carcinogenicity. The National Institute for Occupational Safety and Health (NIOSH) has identified 4,4´-methylenedianiline as a potential occupational carcinogen and recommends that occupational exposures to the compound be limited to the lowest feasible concentration (NIOSH 1994).

4,4´-Methylenedianiline is on the list of chemicals appearing in The Emergency Planning and Community Right-to-Know Act of 1986 (EPCRA) (EPA 1987). Section 313 of Title III of EPCRA requires owners and operators of certain facilities that manufacture, import, process, or otherwise use the chemicals on this list to report annually their release of those chemicals to any environmental media.

OSHA requires employers of workers who are occupationally exposed to 4,4´-methylenedianiline to institute engineering controls and work practices to reduce and maintain employee exposure at or below permissible exposure limits (PEL). Except as provided by specific clauses within the standard given at 29 CFR 1910.1050 (e.g., products not capable of releasing MDA in excess of the action level;
conditions where no dermal exposure to MDA can occur, materials in any form containing less than 0.1% MDA by weight or volume, and construction work) for all occupational exposures to MDA, the employer must use engineering and work practice controls, if feasible, to reduce exposure to or below an 8-hour time-weighted average (TWA) of 10 ppb (OSHA 1992a, 1992b). The short term exposure limit (STEL), as determined by any 15-minute sampling period is 100 ppb (OSHA 1992a, 1992b). The OSHA standard for construction work is given in 29 CFR 1926.60; however, the PEL for these activities is the same as that for all other occupational exposures. The action level for occupational exposure to airborne 4,4’-methylenedianiline, including construction work, is 5 ppb, based on an 8-hour time-weighted average (OSHA 1992a, 1992b). The OSHA standard for construction work is applicable, but not limited, to activities such as alteration, repair, maintenance, or renovation of structures or substrates that contain 4,4’-methylenedianiline (OSHA 1992a, 1992b). Respirators must be provided and used during the time period necessary to install or implement feasible engineering and work practice controls (OSHA 1992a, 1992b).
# Table 7-1. Regulations and Guidelines Applicable to 4,4'-Methylenedianiline

<table>
<thead>
<tr>
<th>Agency</th>
<th>Description</th>
<th>Information</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>INTERNATIONAL Guidelines:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>WHO</td>
<td>Group (cancer ranking)</td>
<td>Sufficient Evidence of Carcinogenicity to Experimental Animals Group 2B</td>
<td>IARC 1987</td>
</tr>
<tr>
<td>IARC</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NATIONAL Regulations:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>a. Air:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Permissible Exposure Limit (Ceiling)</td>
<td>Dermal: Eye and skin contact (not permitted)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Short term exposure limit (STEL)</td>
<td>100 ppb</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Action Level for Occupational Exposures</td>
<td>5 ppb</td>
<td></td>
</tr>
<tr>
<td>EPA OAR</td>
<td>Hazardous Air Pollutants (HAPs)</td>
<td>Yes</td>
<td>Clean Air Act Amendment Title III, Section 112 (b) U.S. Congress 1990</td>
</tr>
<tr>
<td></td>
<td>Standards of Performance for New Stationary Sources equipment leaks of VOC in the SOCMILIST of chemicals produced by affected facilities</td>
<td>Yes</td>
<td>40 CFR 60.489 EPA 1981</td>
</tr>
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<td></td>
<td>National Emission Standards for Hazardous Air Pollutants (NESHAPs) from Source Categories-organic hazardous air pollutants from SOCMILIST</td>
<td>Yes</td>
<td>40 CFR 63.106 EPA 1994</td>
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<tr>
<td></td>
<td>National Emission Standards for Wood Furniture Manufacturing Operations Table 2- list of volatile hazardous air pollutants; Table 4-pollutants excluded from use in cleaning and washoff-solvents; Table 6-VHAP of potential concern</td>
<td>Yes</td>
<td>40 CFR 63, Subpart JJ EPA 1995b</td>
</tr>
<tr>
<td>b. Water</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>EPA OW</td>
<td>Effluent Guidelines and Standards organic chemicals, plastics, and synthetic fibers</td>
<td>Yes</td>
<td>40 CFR 414.70 EPA 1992</td>
</tr>
</tbody>
</table>
## 7. REGULATIONS AND ADVISORIES

### Table 7-1. Regulations and Guidelines Applicable to 4,4'-Methylenedianiline (cont.)

<table>
<thead>
<tr>
<th>Agency</th>
<th>Description</th>
<th>Information</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>NATIONAL (cont.)</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>c. Other:</td>
<td></td>
<td></td>
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</tr>
<tr>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Designation, Reportable Quantities, and Notification</td>
<td>1 pound (^a) (0.454 kg)</td>
<td>40 CFR 302.4 EPA 1995c</td>
</tr>
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<td></td>
<td>designation of hazardous substances</td>
<td>10 pounds (4.54 kg) (final RQ)</td>
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<tr>
<td></td>
<td>Community Right-to-Know -applicable chemicals and chemical categories</td>
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<td></td>
</tr>
<tr>
<td>FDA</td>
<td>Indirect Food Additives: Adhesives and Components of Coatings</td>
<td>Yes</td>
<td>21 CFR 175.300 FDA 1977a</td>
</tr>
<tr>
<td></td>
<td>substances for use as components of coatings-resinous and polymeric coatings</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Indirect Food Additives: Polymer</td>
<td>Yes</td>
<td>21 CFR 177.1680 FDA 1977b</td>
</tr>
<tr>
<td></td>
<td>substances for use as basic components of single and repeated use food contact surfaces-polyurethane resins</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>substances for use only as components of articles intended for repeated use-4,4'-isopropylidenephenolepichlorohydrin thermosetting epoxy resins</td>
<td>Yes</td>
<td>21 CFR 177.2280 FDA 1977c</td>
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<tr>
<td>Guidelines:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>ACGIH</td>
<td>Ceiling Limit for Occupational Exposure (TWA)</td>
<td>0.1 ppm (skin) 0.81 mg/m(^3) (skin)</td>
<td>ACGIH 1996</td>
</tr>
<tr>
<td>NIOSH</td>
<td>Recommended Exposure Limit for Occupational Exposure (TWA)</td>
<td>Potential occupational carcinogen; lowest feasible concentration (0.03 mg/m(^3)LOQ)</td>
<td>NIOSH 1994</td>
</tr>
</tbody>
</table>

### STATE

#### Regulations and Guidelines:

<table>
<thead>
<tr>
<th>Agency</th>
<th>Average Acceptable Ambient Air Concentrations Guidelines or Standards</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>CT</td>
<td>8 hours</td>
<td>8 (\mu)g/m(^3)</td>
</tr>
<tr>
<td>FL (Ft. L'dle)</td>
<td>8 hours</td>
<td>8x10(^{-5}) mg/m(^3)</td>
</tr>
<tr>
<td>FL (Pinellas Co.)</td>
<td>8 hours, 24 hours</td>
<td>8 (\mu)g/m(^3) 1.92 (\mu)g/m(^3)</td>
</tr>
<tr>
<td>KS</td>
<td>Annual</td>
<td>4x10(^{-5}) (\mu)g/m(^3)</td>
</tr>
<tr>
<td>KS-KC</td>
<td>Annual</td>
<td>4x10(^{-5}) (\mu)g/m(^3)</td>
</tr>
</tbody>
</table>
Table 7-1. Regulations and Guidelines Applicable to 4,4’-Methylenedianiline (cont.)

<table>
<thead>
<tr>
<th>Agency</th>
<th>Description</th>
<th>Information</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>STATE</td>
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<td></td>
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<tr>
<td>ND</td>
<td>NA</td>
<td>BACT</td>
<td></td>
</tr>
<tr>
<td>NV</td>
<td>8 hours</td>
<td>1.90x10^5 mg/m³</td>
<td></td>
</tr>
<tr>
<td>NY</td>
<td>Annual</td>
<td>2.70x10^-1 μg/m³</td>
<td></td>
</tr>
<tr>
<td>OK</td>
<td>24 hours</td>
<td>8 μg/m³</td>
<td></td>
</tr>
<tr>
<td>SC</td>
<td>24 hours</td>
<td>4 μg/m³</td>
<td></td>
</tr>
<tr>
<td>TX</td>
<td>30 minutes</td>
<td>8.10 μg/m³</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Annual</td>
<td>8.10x10^(-1) μg/m³</td>
<td></td>
</tr>
<tr>
<td>VA</td>
<td>24 hours</td>
<td>8.10 μg/m³</td>
<td></td>
</tr>
<tr>
<td>WA-SWEST</td>
<td>24 hours</td>
<td>2.60 μg/m³</td>
<td></td>
</tr>
</tbody>
</table>

a OSHA 1992 denotes the effective date (September 9, 1992) for the standard.
b Indicates that the statutory source for designation of this hazardous substance under CERCLA is CWA Section 3(b)(4).

ACGIH = American Conference of Governmental Industrial Hygienists; BACT = Best Available Control Technology; CERCLA = Comprehensive Environmental Response, Compensation, and Liability Act; CFR = Code of Federal Regulations; CWA = Clean Water Act; DOT = Department of Transportation; EPA = Environmental Protection Agency; IARC = International Agency for Research on Cancer; LOQ = Limit of Quantitation; NAAQS = National Ambient Air Quality Standard; NATICH = National Air Toxics Information Clearinghouse; NIOSH = National Institute for Occupational Safety and Health; OERR = Office of Emergency and Remedial Response; OSHA = Occupational Health and Safety Administration; OWRS = Office of Water Regulations and Standards; PAH = Polycyclic Aromatic Hydrocarbons; PEL = Permissible Exposure Limit; RCRA = Resource Conservation and Recovery Act; REL = Recommended Exposure Limit; RID = Reference Dose; STEL = Short Term Exposure Level; TLV = Threshold Limit Value; TWA = Time Weighted Average; u.f. = Uncertainty Factor; VHAP = Volatile Hazardous Air Pollutants; WHO = World Health Organization.
8. REFERENCES


*ACGIH. 1996. Threshold limit values for chemical substances and physical agents and biological exposure indices. Second printing. American Conference of Governmental Industrial Hygienists. Cincinnati, OH.

*Allied Chemical Corp. 1978. Properties, applications and handling of 4,4´-methyleneedianiline. Material Safety Data Sheet.


*ATSDR. 1989. Decision guide for identifying substance-specific data needs related to toxicological profiles. Agency for Toxic Substances and Disease Registry, Division of Toxicology, Atlanta, GA.

*ATSDR/CDC. 1990. Subcommittee report on biological indicators of organ damage. Agency for Toxic Substances and Disease Registry, Centers for Disease Control and Prevention, Atlanta, GA.


*Cited in text


*Brunmark P, Dalene M, Skarping G. 1995. Gas chromatography negative ion chemical ionization mass spectrometry of hydrolyzed human urine and blood plasma for the biomonitoring of occupational exposure to 4,4 methylenebisaniline. Department of Occupational and Environmental Medicine, University Hospital S-221 85 Lund, Sweden.


8. REFERENCES


*CMA. 1982. Summary of information on 4,4,-methylenedianiline collected by a survey of the chemical manufacturers associations’s methylenedianiline program with cover letter dated 11-04-82. EPA OTS Dot. # 40-8261190.


*DuPont. 1975. Ten-day subacute exposure of rabbit to methylene dianiline. Dot ID. 878220288. (Unpublished study)

*DuPont. 1976a. Skin absorption studies in rabbits treated with 4,4´-diaminodiphenylmethane (MDA). A. pathological and clinical effects of a 10-day subacute study. Dot ID. 878220289. (Unpublished study)

*DuPont. 1976b. Eye toxicity of aniline, 4,4´-methylenedianiline (MDA). Dot ID: 878220284. (Unpublished study)


Endo Y, Hara I. 1991. DNA-detection in rats administered with 4,4´-methylenedianiline or 4,4´-methylenebis (2-chloroaniline). Sangyo Igaku 33:430-431. [Japanese].


EPA. 1995a. ASTER Ecotoxicity Profile on 4,4,´-Methylenedianiline. U.S. Environmental Protection Agency, Environmental Research Laboratory, Duluth, MN.


Cancer, Part II: Exposure monitoring and molecular dosimetry, Kailua-Kona, Hawaii, U.S.A.,

*FDA. 1977a. Indirect food additives: Adhesives and components of coatings. Substances for use
as components of coatings-resinous and polymeric coatings. U.S. Food and Drug Administration. 21
CFR 175.300

*FDA. 1977b. Indirect food additives: Polymer. Substances for use as basic components of single
and repeated use food contact surface-polyurethane resins U.S. Food and Drug Administration. 21
CFR 177.1680

*FDA. 1977c. Indirect food additives: Polymer. Substances for use as basic components of articles
intended for repeated use-4,4-isopropylidenediphenolepichlorohydrin thermosetting epoxy resins. U.S.
Food and Drug Administration. 21 CFR 177. 2280


Results of 70 coded chemicals tested for the National Toxicology Program. Environ Molecular
Mutagenesis 23(3):208-227.

on liver, kidney and bladder carcinogenesis in rats ingesting N-ethyl-N-hydroxyethylnitrosamine or


intragastric doses of aromatic and heterocyclic nitro or amino derivatives in young female Sprague-

*Gunderson EC, Anderson CC. 1988. A sampling and analytical method for airborne m-phenylenediamine
(MPDA) and 4,4´-methylenedianiline (MDA). Am Ind Hyg Assoc J 49 (10):531-538.

agents on induction of preneoplastic and neoplastic lesions in a medium-term multi-organ carcinogenesis


*HazDat. 1997. Database. Agency for Toxic Substances and Disease Registry (ATSDR), Atlanta, GA.


*Holland JM, Smith LH, Frome E, ET AL. 1987. Test of Carcinogenicity in Mouse Skin: Methylenedianiline, gamma Glycidoxytrimethyloxysilane, gamma Aminopropyltriethoxysilane and a mixture of M-phenylenediamine, methylenedianiline, and diglycidylether of Bisphenol-A. Govt Reports Announcements & Index (GRA&I), Issue 23.


8. REFERENCES


*NIOSH. 1997. NIOSH pocket guide to chemical hazards. National Institute for Occupational Safety and Health, Cincinnati, OH.


8. REFERENCES


9. GLOSSARY

**Acute Exposure**—Exposure to a chemical for a duration of 14 days or less, as specified in the Toxicological Profiles.

**Adsorption Coefficient** ($K_{oc}$)—The ratio of the amount of a chemical adsorbed per unit weight of organic carbon in the soil or sediment to the concentration of the chemical in solution at equilibrium.

**Adsorption Ratio** ($K_d$)—The amount of a chemical adsorbed by a sediment or soil (i.e., the solid phase) divided by the amount of chemical in the solution phase, which is in equilibrium with the solid phase, at a fixed solid/solution ratio. It is generally expressed in micrograms of chemical sorbed per gram of soil or sediment.

**Bioconcentration Factor** ($BCF$)—The quotient of the concentration of a chemical in aquatic organisms at a specific time or during a discrete time period of exposure divided by the concentration in the surrounding water at the same time or during the same period.

**Cancer Effect Level** ($CEL$)—The lowest dose of chemical in a study, or group of studies, that produces significant increases in the incidence of cancer (or tumors) between the exposed population and its appropriate control.

**Carcinogen**—A chemical capable of inducing cancer.

**Ceiling Value**—A concentration of a substance that should not be exceeded, even instantaneously.

**Chronic Exposure**—Exposure to a chemical for 365 days or more, as specified in the Toxicological Profiles.

**Developmental Toxicity**—The occurrence of adverse effects on the developing organism that may result from exposure to a chemical prior to conception (either parent), during prenatal development, or postnatally to the time of sexual maturation. Adverse developmental effects may be detected at any point in the life span of the organism.

**Embryotoxicity and Fetotoxicity**—Any toxic effect on the conceptus as a result of prenatal exposure to a chemical; the distinguishing feature between the two terms is the stage of development during which the insult occurred. The terms, as used here, include malformations and variations, altered growth, and in utero death.

**EPA Health Advisory**—An estimate of acceptable drinking water levels for a chemical substance based on health effects information. A health advisory is not a legally enforceable federal standard, but serves as technical guidance to assist federal, state, and local officials.

**Immediately Dangerous to Life or Health** (IDLH)—The maximum environmental concentration of a contaminant from which one could escape within 30 min without any escape-impairing symptoms or irreversible health effects.
**Intermediate Exposure**—Exposure to a chemical for a duration of 15-364 days, as specified in the Toxicological Profiles.

**Immunologic Toxicity**—The occurrence of adverse effects on the immune system that may result from exposure to environmental agents such as chemicals.

**In vitro**—Isolated from the living organism and artificially maintained, as in a test tube.

**In vivo**—Occurring within the living organism.

**Lethal Concentration_{(LO)} (LC_{LO})**—The lowest concentration of a chemical in air which has been reported to have caused death in humans or animals.

**Lethal Concentration_{(50)} (LC_{50})**—A calculated concentration of a chemical in air to which exposure for a specific length of time is expected to cause death in 50% of a defined experimental animal population.

**Lethal Dose_{(LO)} (LD_{LO})**—The lowest dose of a chemical introduced by a route other than inhalation that is expected to have caused death in humans or animals.

**Lethal Dose_{(50)} (LD_{50})**—The dose of a chemical which has been calculated to cause death in 50% of a defined experimental animal population.

**Lethal Time_{(50)} (LT_{50})**—A calculated period of time within which a specific concentration of a chemical is expected to cause death in 50% of a defined experimental animal population.

**Lowest-Observed-Adverse-Effect Level (LOAEL)**—The lowest dose of chemical in a study, or group of studies, that produces statistically or biologically significant increases in frequency or severity of adverse effects between the exposed population and its appropriate control.

**Malformations**—Permanent structural changes that may adversely affect survival, development, or function.

**Minimal Risk Level**—An estimate of daily human exposure to a dose of a chemical that is likely to be without an appreciable risk of adverse noncancerous effects over a specified duration of exposure.

**Mutagen**—A substance that causes mutations. A mutation is a change in the genetic material in a body cell. Mutations can lead to birth defects, miscarriages, or cancer.

**Neurotoxicity**—The occurrence of adverse effects on the nervous system following exposure to chemical.

**No-Observed-Adverse-Effect Level (NOAEL)**—The dose of chemical at which there were no statistically or biologically significant increases in frequency or severity of adverse effects seen between the exposed population and its appropriate control. Effects may be produced at this dose, but they are not considered to be adverse.
Octanol-Water Partition Coefficient (K_{ow})-The equilibrium ratio of the concentrations of a chemical in n-octanol and water, in dilute solution.

Permissible Exposure Limit (PEL)-An allowable exposure level in workplace air averaged over an 8-hour shift.

q_{1}*-The upper-bound estimate of the low-dose slope of the dose-response curve as determined by the multistage procedure. The q_{1}* can be used to calculate an estimate of carcinogenic potency, the incremental excess cancer risk per unit of exposure (usually \( \mu g/L \) for water, mg/kg/day for food, and pg/m\(^3\) for air).

Reference Dose (RfD)-An estimate (with uncertainty spanning perhaps an order of magnitude) of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime. The RfD is operationally derived from the NOAEL (from animal and human studies) by a consistent application of uncertainty factors that reflect various types of data used to estimate RfDs and an additional modifying factor, which is based on a professional judgment of the entire database on the chemical. The RfDs are not applicable to nonthreshold effects such as cancer.

Reportable Quantity (RQ)-The quantity of a hazardous substance that is considered reportable under CERCLA. Reportable quantities are (1) 1 pound or greater or (2) for selected substances, an amount established by regulation either under CERCLA or under Sect. 3 11 of the Clean Water Act. Quantities are measured over a 24-hour period.

Reproductive Toxicity-The occurrence of adverse effects on the reproductive system that may result from exposure to a chemical. The toxicity may be directed to the reproductive organs and/or the related endocrine system. The manifestation of such toxicity may be noted as alterations in sexual behavior, fertility, pregnancy outcomes, or modifications in other functions that are dependent on the integrity of this system.

Short-Term Exposure Limit (STEL)-The maximum concentration to which workers can be exposed for up to 15 min continually. No more than four excursions are allowed per day, and there must be at least 60 min between exposure periods. The daily TLV-TWA may not be exceeded.

Target Organ Toxicity-This term covers a broad range of adverse effects on target organs or physiological systems (e.g., renal, cardiovascular) extending from those arising through a single limited exposure to those assumed over a lifetime of exposure to a chemical.

Teratogen-A chemical that causes structural defects that affect the development of an organism.

Threshold Limit Value (TLV)-A concentration of a substance to which most workers can be exposed without adverse effect. The TLV may be expressed as a TWA, as a STEL, or as a CL.

Time-Weighted Average (TWA)-An allowable exposure concentration averaged over a normal 8-hour workday or 40-hour workweek.

Toxic Dose (TD_{50})-A calculated dose of a chemical, introduced by a route other than inhalation, which is expected to cause a specific toxic effect in 50% of a defined experimental animal population.
**Uncertainty Factor (UF)**-A factor used in operationally deriving the MRL or RfD from experimental data. UFs are intended to account for (1) the variation in sensitivity among the members of the human population, (2) the uncertainty in extrapolating animal data to the case of human, (3) the uncertainty in extrapolating from data obtained in a study that is of less than lifetime exposure, and (4) the uncertainty in using LOAEL data rather than NOAEL data. Usually each of these factors is set equal to 10.
APPENDIX A
ATSDR MINIMAL RISK LEVEL

The Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) [42 U.S.C. 9601 et seq.], as amended by the Super-fund Amendments and Reauthorization Act (SARA) [Pub. L. 99-4991, requires that the Agency for Toxic Substances and Disease Registry (ATSDR) develop jointly with the U.S. Environmental Protection Agency (EPA), in order of priority, a list of hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL); prepare toxicological profiles for each substance included on the priority list of hazardous substances; and assure the initiation of a research program to fill identified data needs associated with the substances.

The toxicological profiles include an examination, summary, and interpretation of available toxicological information and epidemiologic evaluations of a hazardous substance. During the development of toxicological profiles, Minimal Risk Levels (MRLs) are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration for a given route of exposure. An MRL is an estimate of the daily human exposure to a hazardous substance that is likely to be without appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are based on noncancer health effects only and are not based on a consideration of cancer effects. These substance-specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors to identify contaminants and potential health effects that may be of concern at hazardous waste sites. It is important to note that MRLs are not intended to define clean-up or action levels.

MRLs are derived for hazardous substances using the no-observed-adverse-effect level/uncertainty factor approach. They are below levels that might cause adverse health effects in the people most sensitive to such chemical-induced effects. MRLs are derived for acute (1-14 days), intermediate (15-364 days), and chronic (365 days and longer) durations and for the oral and inhalation routes of exposure. Currently, MRLs for the dermal route of exposure are not derived because ATSDR has not yet identified a method suitable for this route of exposure. MRLs are generally based on the most sensitive chemical-induced end point considered to be of relevance to humans. Serious health effects (such as irreparable damage to the liver or kidneys, or birth defects) are not used as a basis for
establishing MRLs. Exposure to a level above the MRL does not mean that adverse health effects will occur.

MRLs are intended only to serve as a screening tool to help public health professionals decide where to look more closely. They may also be viewed as a mechanism to identify those hazardous waste sites that are not expected to cause adverse health effects. Most MRLs contain a degree of uncertainty because of the lack of precise toxicological information on the people who might be most sensitive (e.g., infants, elderly, nutritionally or immunologically compromised) to the effects of hazardous substances. ATSDR uses a conservative (i.e., protective) approach to address this uncertainty consistent with the public health principle of prevention. Although human data are preferred, MRLs often must be based on animal studies because relevant human studies are lacking. In the absence of evidence to the contrary, ATSDR assumes that humans are more sensitive to the effects of hazardous substance than animals and that certain persons may be particularly sensitive. Thus, the resulting MRL may be as much as a hundredfold below levels that have been shown to be nontoxic in laboratory animals.

Proposed MRLs undergo a rigorous review process: Health Effects/MRL Workgroup reviews within the Division of Toxicology, expert panel peer reviews, and agencywide MRL Workgroup reviews, with participation from other federal agencies and comments from the public. They are subject to change as new information becomes available concomitant with updating the toxicological profiles. Thus, MRLs in the most recent toxicological profiles supersede previously published levels. For additional information regarding MRLs, please contact the Division of Toxicology, Agency for Toxic Substances and Disease Registry, 1600 Clifton Road, Mailstop E-29, Atlanta, Georgia 30333.
APPENDIX A

MINIMAL RISK LEVEL WORKSHEET

Chemical Name: 4,4'-Methyleneedianiline
CAS Number: 101-77-9
Date: November 1997
Profile Status: Final Draft Post Public
Route: [X] Inhalation  [X] Oral
Duration: [X] Acute  [ ] Intermediate  [ ] Chronic
Graph Key: 6r
Species: Rat

Minimal Risk Level: 0.2  [X] mg/kg/day  [ ] ppm


Experimental design: Groups of male Sprague-Dawley rats (3–5/group) (175–300 g body weight) were administered orally a single dose of 0 (vehicle alone) or 25, 50, 75, 100, 125, or 225 mg 4,4'-methyleneedianiline/kg in corn oil. Twenty-four hours after dosing, the rats were anesthetized, the bile duct was cannulated, bile was collected for 30 minutes and then the rats were sacrificed; indicators of liver injury were assessed. Additional groups of rats received a dose of 100 mg/kg and were sacrificed at various intervals for histopathological examination of the liver. The possible involvement of the cytochrome P-450 system in the toxicity of 4,4'-methylenedianiline was investigated. The inhibitors of monoxygenase (MO) function used were aminobenzotriazol (ABT) and SKF-525A (both administered intraperitoneally 2 hours before 4,4'-methylenedianiline). The inducers of MO function used were phenobarbital (PB) and β-naphthoflavone (BNF) (both administered intraperitoneally daily for 3 days before 4,4'-methylenedianiline).

Effects noted in study and corresponding doses: Administration of 4,4'-methylenedianiline caused a dose-dependent change in all markers of hepatic parenchyma injury: increased serum alanine aminotransferase (ALT) and gamma-glutamyl transferase, increased serum bilirubin, decreased bile flow, and increased relative liver weight. The minimal effective dose was between 25 and 75 mg/kg. Histologically, a dose of 100 mg/kg caused hepatocellular necrosis with hemorrhage and moderate neutrophil infiltration. Lesions associated with the portal triads consisted of bile ductular necrosis, portal edema with fibrin exudate, and neutrophil infiltration. The earliest change identified (at 4 hours) was bile ductular necrosis. A segmental necrotizing vasculitis of the portal vein was also observed. The severity of the effects continued to increase over a 16-hour period. Time-course experiments showed that the first significant biochemical markers of liver injury appeared about 8 hours after dosing. Pretreatment with ABT ameliorated the hepatic effects of 4,4'-methylenedianiline, but SKF-525A did not. This according to the authors, may have reflected a difference in the spectrum of cytochrome P-450 isozymes inhibited by the two agents. Pretreatment with PB had no effect after a 100 mg/kg dose of 4,4'-methylenedianiline, but attenuated the hepatotoxicity of a 50 mg/kg dose. BNF had a small attenuating effect.

Dose and end point used for MRL derivation: 25 mg/kg; minimal liver effects.

[ ] NOAEL  [X] LOAEL
Uncertainty Factors used in MRL derivation:

- [X] 3 for use of a minimal LOAEL
- [X] 10 for extrapolation from animals to humans
- [X] 10 for human variability

A modifying factor of 0.5 was used in the derivation of the MRL in order to account for the possibility of increased absorption of 4,4'-methylenedianiline due to the corn oil vehicle.

Was a conversion factor used from ppm in food or water to a mg/body weight dose?
No

If an inhalation study in animals, list conversion factors used in determining human equivalent dose:
NA

Was a conversion used from intermittent to continuous exposure?
No

Other additional studies or pertinent information that lend support to this MRL: Several other studies support the findings of Bailie et al. (1993). For example, Bailie et al. (1994) observed cholestasis, biliary epithelial injury, and hepatic parenchymal damage in the livers from rats treated with a single dose of 5 mg 4,4'-methylenedianiline/kg. Schmidt et al. (1980) reported bile duct necrosis and increased serum transaminases in rats also after a single dose of 50 mg/kg. The liver is a known target of 4,4'-methylenedianiline in humans and animals. An outbreak of toxic hepatitis was described in a group of individuals who ate bread contaminated with 4,4'-methylenedianiline (Kopelman et al. 1966). Liver toxicity has also been reported in humans and animals exposed by the dermal route (Brooks et al. 1979; DuPont 1976a; McGill and Motto 1974; Williams et al. 1974)

Agency Contact (Chemical Manager): Zemoria Rosemond
MINIMAL RISK LEVEL WORKSHEET

Chemical Name: 4,4'-Methylenedianiline
CAS Number: 101-77-9
Date: November 1997
Profile Status: Final Draft Post Public
Route: [ ] Inhalation [X] Oral
Duration: [ ] Acute [X] Intermediate [ ] Chronic
Graph Key: 23r
Species: Rat

Minimal Risk Level: 0.08 [X] mg/kg/day [ ] ppm


Experimental design: Groups of male and female Wistar rats (10/sex/group) were administered 0 (vehicle alone) 8.3, or 83 mg 4,4'-methylenedianiline by gavage in propylene glycol once a day for 12 weeks. End points evaluated included body weight, serum protein profile, hemoglobin levels and erythrocyte counts, and gross and histopathological appearance of liver, kidneys, and spleen.

Effects noted in study and corresponding doses: The only reported effects observed with the 8.3 mg/kg/day dose of 4,4'-methylenedianiline were unspecified histological lesions in the liver of one animal and unspecified lesions in the spleens from all rats. Administration of 83 mg 4,4'-methylenedianiline/kg/day did not affect body weight, hemoglobin levels, or red blood cell count. Analysis of electrophoretic patterns of serum showed an increase in beta-globulin and a decrease in the albumin fraction. On gross examination, the liver and kidneys appeared enlarged and their relative weights were markedly increased. Flatulence and intestinal occlusion were also described. Microscopical examination of the liver revealed intense degenerative lesions in all animals consisting of atrophy of the parenchyma accompanied by hyperplasia of the stroma, particularly at portal areas. The 8.3 mg/kg/day is considered a NOAEL although the authors noted unspecified spleen lesions in all rats. The significance of these lesions is unknown since a 13-week study conducted by NTP (1983) observed no gross or histopathological alterations in the spleen, thymus, and lymph nodes from rats treated with up to 141 mg 4,4'-methylenedianiline/kg/day in the drinking water. No lesions were observed in the same organs from mice treated similarly with up to 116 mg/kg/day (NTP 1983).

Dose and end point used for MRL derivation: 8.3 mg/kg/day; liver effects.

[X] NOAEL  [ ] LOAEL

Uncertainty Factors used in MRL derivation:

[ ] 10 for use of a LOAEL  
[X] 10 for extrapolation from animals to humans  
[X] 10 for human variability
Was a conversion factor used from ppm in food or water to a mg/body weight dose?
No

If an inhalation study in animals, list conversion factors used in determining human equivalent dose:
NA

Was a conversion used from intermittent to continuous exposure?
No

Other additional studies or pertinent information that lend support to this MRL: The liver is a known target for 4,4'-methyleneedianiline toxicity in humans and animals regardless of the route of exposure. The LOAEL of 83 mg/kg/day for liver effects is consistent with LOAELs identified in a number of intermediate-duration studies (Fukushima et al. 1979, 1981; Hagiwara et al. 1993; Miyamoto et al. 1977; NTP 1983). These LOAELs ranged from 67 to 100 mg/kg/day. With the exception of the NTP (1983) study, all of these studies tested only one dose level, therefore, no NOAELs could be established. The NTP (1983) study defined a NOAEL of 35 mg/kg/day for hepatic effects in rats and 58 mg/kg/day for mice.

Agency Contact (Chemical Manager): Zemoria Rosemond
APPENDIX B

USER’S GUIDE

Chapter 1

Public Health Statement

This chapter of the profile is a health effects summary written in non-technical language. Its intended audience is the general public especially people living in the vicinity of a hazardous waste site or chemical release. If the Public Health Statement were removed from the rest of the document, it would still communicate to the lay public essential information about the chemical.

The major headings in the Public Health Statement are useful to find specific topics of concern. The topics are written in a question and answer format. The answer to each question includes a sentence that will direct the reader to chapters in the profile that will provide more information on the given topic.

Chapter 2

Tables and Figures for Levels of Significant Exposure (LSE)

Tables (2-1, 2-2, and 2-3) and figures (2-1 and 2-2) are used to summarize health effects and illustrate graphically levels of exposure associated with those effects. These levels cover health effects observed at increasing dose concentrations and durations, differences in response by species, minimal risk levels (MRLs) to humans for noncancer end points, and EPA’s estimated range associated with an upper-bound individual lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. Use the LSE tables and figures for a quick review of the health effects and to locate data for a specific exposure scenario. The LSE tables and figures should always be used in conjunction with the text. All entries in these tables and figures represent studies that provide reliable, quantitative estimates of No-Observed-Adverse-Effect Levels (NOAELs), Lowest-Observed-Adverse-Effect Levels (LOAELs), or Cancer Effect Levels (CELs).

The legends presented below demonstrate the application of these tables and figures. Representative examples of LSE Table 2-1 and Figure 2-1 are shown. The numbers in the left column of the legends correspond to the numbers in the example table and figure.

LEGEND

See LSE Table 2-1

(1) Route of Exposure. One of the first considerations when reviewing the toxicity of a substance using these tables and figures should be the relevant and appropriate route of exposure. When sufficient data exists, three LSE tables and two LSE figures are presented in the document. The three LSE tables present data on the three principal routes of exposure, i.e., inhalation, oral, and dermal (LSE Table 2-1, 2-2, and 2-3, respectively). LSE figures are limited to the inhalation (LSE Figure 2-1) and oral (LSE Figure 2-2) routes. Not all substances will have data on each route of exposure and will not therefore have all five of the tables and figures.
(2) **Exposure Period** Three exposure periods - acute (less than 15 days), intermediate (15-364 days), and chronic (365 days or more) are presented within each relevant route of exposure. In this example, an inhalation study of intermediate exposure duration is reported. For quick reference to health effects occurring from a known length of exposure, locate the applicable exposure period within the LSE table and figure.

(3) **Health Effect** The major categories of health effects included in LSE tables and figures are death, systemic, immunological, neurological, developmental, reproductive, and cancer. NOAELs and LOAELs can be reported in the tables and figures for all effects but cancer. Systemic effects are further defined in the "System" column of the LSE table (see key number 18).

(4) **Key to Figure** Each key number in the LSE table links study information to one or more data points using the same key number in the corresponding LSE figure. In this example, the study represented by key number 18 has been used to derive a NOAEL and a Less Serious LOAEL (also see the 2 "18r" data points in Figure 2-1).

(5) **Species** The test species, whether animal or human, are identified in this column. Section 2.5, "Relevance to Public Health," covers the relevance of animal data to human toxicity and Section 2.3, "Toxicokinetics," contains any available information on comparative toxicokinetics. Although NOAELs and LOAELs are species specific, the levels are extrapolated to equivalent human doses to derive an MRL.

(6) **Exposure Frequency/Duration** The duration of the study and the weekly and daily exposure regimen are provided in this column. This permits comparison of NOAELs and LOAELs from different studies. In this case (key number 18), rats were exposed to 1,1,2,2-tetrachloroethane via inhalation for 6 hours per day, 5 days per week, for 3 weeks. For a more complete review of the dosing regimen refer to the appropriate sections of the text or the original reference paper, i.e., Nitschke et al. 1981.

(7) **System** This column further defines the systemic effects. These systems include: respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, and dermal/ocular. "Other" refers to any systemic effect (e.g., a decrease in body weight) not covered in these systems. In the example of key number 18, 1 systemic effect (respiratory) was investigated.

(8) **NOAEL** A No-Oberved-Adverse-Effect Level (NOAEL) is the highest exposure level at which no harmful effects were seen in the organ system studied. Key number 18 reports a NOAEL of 3 ppm for the respiratory system which was used to derive an intermediate exposure, inhalation MRL of 0.005 ppm (see footnote "b").

(9) **LOAEL** A Lowest-Oberved-Adverse-Effect Level (LOAEL) is the lowest dose used in the study that caused a harmful health effect. LOAELs have been classified into "Less Serious" and "Serious" effects. These distinctions help readers identify the levels of exposure at which adverse health effects first appear and the gradation of effects with increasing dose. A brief description of the specific endpoint used to quantify the adverse effect accompanies the LOAEL. The respiratory effect reported in key number 18 (hyperplasia) is a Less serious LOAEL of 10 ppm. MRLs are not derived from Serious LOAELs.

(10) **Reference** The complete reference citation is given in chapter 8 of the profile.
(11) **CEL**. A Cancer Effect Level (CEL) is the lowest exposure level associated with the onset of carcinogenesis in experimental or epidemiologic studies. CELs are always considered serious effects. The LSE tables and figures do not contain NOAELs for cancer, but the text may report doses not causing measurable cancer increases.

(12) **Footnotes**. Explanations of abbreviations or reference notes for data in the LSE tables are found in the footnotes. Footnote "b" indicates the NOAEL of 3 ppm in key number 18 was used to derive an MRL of 0.005 ppm.

**LEGEND**

See Figure 2-1

LSE figures graphically illustrate the data presented in the corresponding LSE tables. Figures help the reader quickly compare health effects according to exposure concentrations for particular exposure periods.

(13) **Exposure Period**. The same exposure periods appear as in the LSE table. In this example, health effects observed within the intermediate and chronic exposure periods are illustrated.

(14) **Health Effect**. These are the categories of health effects for which reliable quantitative data exists. The same health effects appear in the LSE table.

(15) **Levels of Exposure**. Concentrations or doses for each health effect in the LSE tables are graphically displayed in the LSE figures. Exposure concentration or dose is measured on the log scale “y” axis. Inhalation exposure is reported in mg/m³ or ppm and oral exposure is reported in mg/kg/day.

(16) **NOAEL**. In this example, 18r NOAEL is the critical endpoint for which an intermediate inhalation exposure MRL is based. As you can see from the LSE figure key, the open-circle symbol indicates to a NOAEL for the test species-rat. The key number 18 corresponds to the entry in the LSE table. The dashed descending arrow indicates the extrapolation from the exposure level of 3 ppm (see entry 18 in the Table) to the MRL of 0.005 ppm (see footnote “b” in the LSE table).

(17) **CEL**. Key number 38r is 1 of 3 studies for which Cancer Effect Levels were derived. The diamond symbol refers to a Cancer Effect Level for the test species-mouse. The number 38 corresponds to the entry in the LSE table.

(18) **Estimated Upper-Bound Human Cancer Risk Levels**. This is the range associated with the upper-bound for lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. These risk levels are derived from the EPA’s Human Health Assessment Group’s upper-bound estimates of the slope of the cancer dose response curve at low dose levels ($q_1^*$).

(19) **Key to LSE Figure**. The Key explains the abbreviations and symbols used in the figure.
TABLE 2-1. Levels of Significant Exposure to [Chemical x] – Inhalation

<table>
<thead>
<tr>
<th>Key to figure*</th>
<th>Species</th>
<th>Exposure frequency/duration</th>
<th>System</th>
<th>NOAEL (ppm)</th>
<th>LOAEL (effect)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Less serious (ppm)</td>
<td>Serious (ppm)</td>
</tr>
<tr>
<td>2</td>
<td>INTERMEDIATE EXPOSURE</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Systemic</td>
<td>↓</td>
<td>↓</td>
<td>↓</td>
<td>3&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>4</td>
<td>18</td>
<td>Rat</td>
<td>13 wk 5d/wk 6hr/d</td>
<td>Resp</td>
<td></td>
</tr>
</tbody>
</table>

CHRONIC EXPOSURE

<table>
<thead>
<tr>
<th>Cancer</th>
<th>Species</th>
<th>Exposure frequency/duration</th>
<th>System</th>
<th>NOAEL (ppm)</th>
<th>LOAEL (effect)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>38</td>
<td>Rat</td>
<td>18 mo 5d/wk 7hr/d</td>
<td></td>
<td>(CEL, multiple organs)</td>
<td>Wong et al. 1982</td>
</tr>
<tr>
<td>39</td>
<td>Rat</td>
<td>89–104 wk 5d/wk 6hr/d</td>
<td></td>
<td>10</td>
<td>(CEL, lung tumors, nasal tumors)</td>
</tr>
<tr>
<td>40</td>
<td>Mouse</td>
<td>79–103 wk 5d/wk 6hr/d</td>
<td></td>
<td>10</td>
<td>(CEL, lung tumors, hemangiosarcomas)</td>
</tr>
</tbody>
</table>

<sup>a</sup> The number corresponds to entries in Figure 2-1.

<sup>b</sup> Used to derive an intermediate inhalation Minimal Risk Level (MRL) of 5 x 10<sup>3</sup> ppm; dose adjusted for intermittent exposure and divided by an uncertainty factor of 100 (10 for extrapolation from animal to humans, 10 for human variability).
Figure 2-1. Levels of Significant Exposure to [Chemical X] – Inhalation

Acute
(≤14 days)

Systemic

Death
Respiratory
Hematological

Death
Respiratory
Hematological
Hepatic
Reproductive
Cancer

10^4
10^5
10^6
10^7

Key

- Rat
- Mouse
- Rabbit
- Guinea Pig
- Monkey

- LOAEL for serious effects (animals)
- LOAEL for less serious effects (animals)
- NOAEL (animals)
- CEL - Cancer Effect Level

18r 30r 31r 33r 35m 37m 38r 40m 39r

- Minimal risk level for effects other than cancer

- The number next to each point corresponds to entries in the accompanying table.

* Doses represent the lowest dose tested per study that produced a tumorigenic response and do not imply the existence of a threshold for the cancer end point.
Chapter 2 (Section 2.5)

Relevance to Public Health

The Relevance to Public Health section provides a health effects summary based on evaluations of existing toxicologic, epidemiologic, and toxicokinetic information. This summary is designed to present interpretive, weight-of-evidence discussions for human health end points by addressing the following questions.

1. What effects are known to occur in humans?

2. What effects observed in animals are likely to be of concern to humans?

3. What exposure conditions are likely to be of concern to humans, especially around hazardous waste sites

The section covers end points in the same order they appear within the Discussion of Health Effects by Route of Exposure section, by route (inhalation, oral, dermal) and within route by effect. Human data are presented first, then animal data. Both are organized by duration (acute, intermediate, chronic). In vitro data and data from parenteral routes (intramuscular, intravenous, subcutaneous, etc.) are also considered in this section. If data are located in the scientific literature, a table of genotoxicity information is included.

The carcinogenic potential of the profiled substance is qualitatively evaluated, when appropriate, using existing toxicokinetic, genotoxic, and carcinogenic data. ATSDR does not currently assess cancer potency or perform cancer risk assessments. Minimal risk levels (MRLs) for noncancer end points (if derived) and the end points from which they were derived are indicated and discussed.

Limitations to existing scientific literature that prevent a satisfactory evaluation of the relevance to public health are identified in the Data Needs section.

Interpretation of Minimal Risk Levels

Where sufficient toxicologic information is available, we have derived minimal risk levels (MRLs) for inhalation and oral routes of entry at each duration of exposure (acute, intermediate, and chronic). These MRLs are not meant to support regulatory action; but to acquaint health professionals with exposure levels at which adverse health effects are not expected to occur in humans. They should help physicians and public health officials determine the safety of a community living near a chemical emission, given the concentration of a contaminant in air or the estimated daily dose in water. MRLs are based largely on toxicological studies in animals and on reports of human occupational exposure.

MRL users should be familiar with the toxicologic information on which the number is based. Chapter 2.5, “Relevance to Public Health,” contains basic information known about the substance. Other sections such as 2.7, “Interactions with Other Substances,” and 2.8, “Populations that are Unusually Susceptible” provide important supplemental information.

MRL users should also understand the MRL derivation methodology. MRLs are derived using a modified version of the risk assessment methodology the Environmental Protection Agency (EPA) provides (Barnes and Dourson 1988) to determine reference doses for lifetime exposure (RfDs).
To derive an MRL, ATSDR generally selects the most sensitive endpoint which, in its best judgement, represents the most sensitive human health effect for a given exposure route and duration. ATSDR cannot make this judgement or derive an MRL unless information (quantitative or qualitative) is available for all potential systemic, neurological, and developmental effects. If this information and reliable quantitative data on the chosen endpoint are available, ATSDR derives an MRL using the most sensitive species (when information from multiple species is available) with the highest NOAEL that does not exceed any adverse effect levels. When a NOAEL is not available, a lowest-observed-adverse-effect level (LOAEL) can be used to derive an MRL, and an uncertainty factor (UF) of 10 must be employed. Additional uncertainty factors of 10 must be used both for human variability to protect sensitive subpopulations (people who are most susceptible to the health effects caused by the substance) and for interspecies variability (extrapolation from animals to humans). In deriving an MRL, these individual uncertainty factors are multiplied together. The product is then divided into the inhalation concentration or oral dosage selected from the study. Uncertainty factors used in developing a substance-specific MRL are provided in the footnotes of the LSE Tables.
APPENDIX C

ACRONYMS, ABBREVIATIONS, AND SYMBOLS

ACGIH  American Conference of Governmental Industrial Hygienists
ADME  Absorption, Distribution, Metabolism, and Excretion
atm  atmosphere
ATSDR  Agency for Toxic Substances and Disease Registry
BCF  bioconcentration factor
BSC  Board of Scientific Counselors
C  Centigrade
CDC  Centers for Disease Control
CEL  Cancer Effect Level
CERCLA  Comprehensive Environmental Response, Compensation, and Liability Act
CFR  Code of Federal Regulations
CLP  Contract Laboratory Program
cm  centimeter
CNS  central nervous system
d  day
DHEW  Department of Health, Education, and Welfare
DHHS  Department of Health and Human Services
DOL  Department of Labor
ECG  electrocardiogram
EEG  electroencephalogram
EPA  Environmental Protection Agency
EKG  see ECG
F  Fahrenheit
F_1  first filial generation
FAO  Food and Agricultural Organization of the United Nations
FEMA  Federal Emergency Management Agency
FIFRA  Federal Insecticide, Fungicide, and Rodenticide Act
fpm  feet per minute
ft  foot
FR  Federal Register
g  gram
GC  gas chromatography
gen  generation
HPLC  high-performance liquid chromatography
hr  hour
IDLH  Immediately Dangerous to Life and Health
IARC  International Agency for Research on Cancer
ILO  International Labor Organization
in  inch
Kd  adsorption ratio
kg  kilogram
kkg  metric ton
K_{OC}  organic carbon partition coefficient
K_{OW}  octanol-water partition coefficient
<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>L</td>
<td>liter</td>
</tr>
<tr>
<td>LC</td>
<td>liquid chromatography</td>
</tr>
<tr>
<td>LC_{50}</td>
<td>lethal concentration, 50% kill</td>
</tr>
<tr>
<td>LD_{50}</td>
<td>lethal dose, 50% kill</td>
</tr>
<tr>
<td>LOAEL</td>
<td>lowest-observed-adverse-effect level</td>
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<tr>
<td>LSE</td>
<td>Levels of Significant Exposure</td>
</tr>
<tr>
<td>m</td>
<td>meter</td>
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<tr>
<td>mg</td>
<td>milligram</td>
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<tr>
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<td>minute</td>
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<tr>
<td>mL</td>
<td>milliliter</td>
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<td>millimeter</td>
</tr>
<tr>
<td>mmHg</td>
<td>millimeters of mercury</td>
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<tr>
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<td>millimole</td>
</tr>
<tr>
<td>mo</td>
<td>month</td>
</tr>
<tr>
<td>mppcf</td>
<td>millions of particles per cubic foot</td>
</tr>
<tr>
<td>MRL</td>
<td>Minimal Risk Level</td>
</tr>
<tr>
<td>MS</td>
<td>mass spectrometry</td>
</tr>
<tr>
<td>NIEHS</td>
<td>National Institute of Environmental Health Sciences</td>
</tr>
<tr>
<td>NIOSH</td>
<td>National Institute for Occupational Safety and Health</td>
</tr>
<tr>
<td>NIOSHTIC</td>
<td>NIOSH's Computerized Information Retrieval System</td>
</tr>
<tr>
<td>ng</td>
<td>nanogram</td>
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<td>nm</td>
<td>nanometer</td>
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<td>National Health and Nutrition Examination Survey</td>
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<td>NOAEL</td>
<td>no-observed-adverse-effect level</td>
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<td>NOES</td>
<td>National Occupational Exposure Survey</td>
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<td>National Occupational Hazard Survey</td>
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<td>National Priorities List</td>
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<td>National Research Council</td>
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<td>National Technical Information Service</td>
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<tr>
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<td>National Toxicology Program</td>
</tr>
<tr>
<td>OSHA</td>
<td>Occupational Safety and Health Administration</td>
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<tr>
<td>PEL</td>
<td>permissible exposure limit</td>
</tr>
<tr>
<td>pg</td>
<td>picogram</td>
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<tr>
<td>pmol</td>
<td>picomole</td>
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<tr>
<td>PMR</td>
<td>proportionate mortality ratio</td>
</tr>
<tr>
<td>ppb</td>
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<td>parts per million</td>
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<td>recommended exposure limit</td>
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<td>RfD</td>
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<td>Registry of Toxic Effects of Chemical Substances</td>
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<td>SCE</td>
<td>sister chromatid exchange</td>
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<td>SIC</td>
<td>Standard Industrial Classification</td>
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<td>standard mortality ratio</td>
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<td>Definition</td>
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<td>STEL</td>
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<td>STORAGE and RETRIEVAL</td>
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<td>threshold limit value</td>
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<td>TSCA</td>
<td>Toxic Substances Control Act</td>
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<td>Toxics Release Inventory</td>
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<td>TWA</td>
<td>time-weighted average</td>
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<td>World Health Organization</td>
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